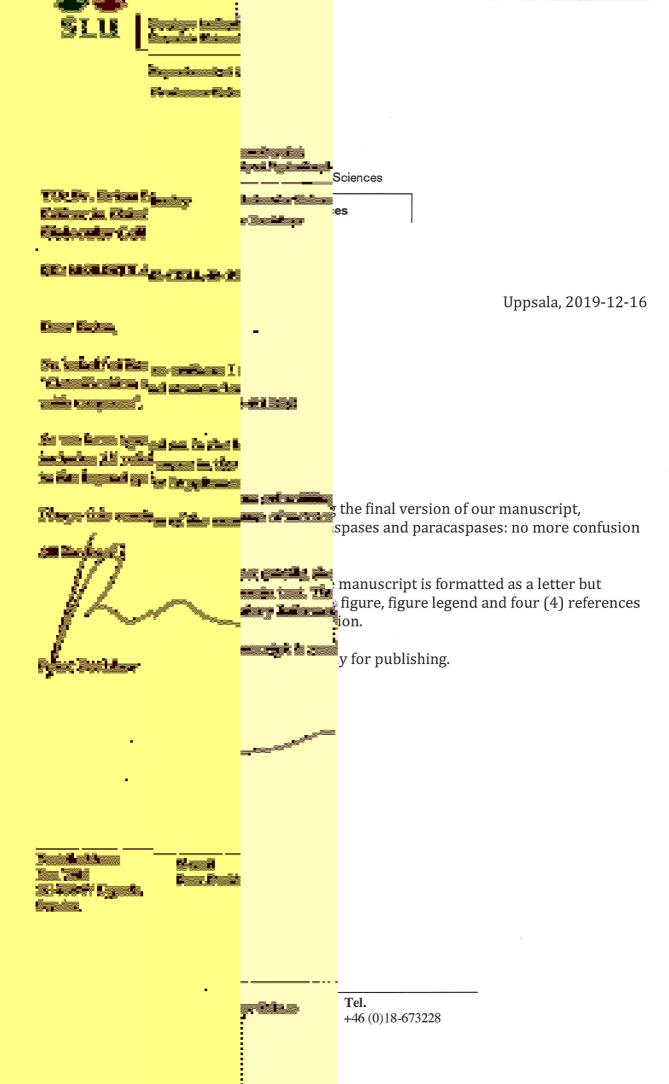
Molecular Cell

Classification and nomenclature of metacaspases and paracaspases: no more confusion with caspases --Manuscript Draft--

Manuscript Number:	MOLECULAR-CELL-D-19-01986R1
Full Title:	Classification and nomenclature of metacaspases and paracaspases: no more
T dil Tido	confusion with caspases
Article Type:	Letter to the Editor
Keywords:	Caspases; Metacaspases; Paracaspases; Proteases; Classification; Nomenclature
Corresponding Author:	Peter Bozhkov Swedish University of Agricultural Sciences Uppsala, SWEDEN
First Author:	Elena Minina
Order of Authors:	Elena Minina
	Jens Staal
	Vanina Alvarez
	John Berges
	Ilana Berman-Frank
	Rudy Beyaert
	Kay Bidle
	Frédéric Bornancin
	Magali Casanova
	Juan Cazzulo
	Chang Jae Choi
	Nuria Coll
	Vishva Dixit
	Marko Dolinar
	Nicolas Fasel
	Christiane Funk
	Patrick Gallois
	Kris Gevaert
	Emilio Gutierrez-Beltran
	Stephan Hailfinger
	Marina Klemencic
	Eugene Koonin
	Daniel Krappmann
	Anna Linusson
	Mauricio Maca uanria Coll

Jeremy Mottram
Thomas Nyström
Heinz Osiewacz
Christopher Overall
Kailash Pandey
Jürgen Ruland
Guy Salvesen
Yigong Shi
Andrei Smertenko
Simon Stael
Jerry Ståhlberg
Maria Suarez
Margot Thome
Hannele Tuominen
Frank Van Breusegem
Renier van der Hoorn
Assaf Vardi
Boris Zhivotovsky
Eric Lam
Peter Bozhkov
Not required for Letters as far as I know.
Response



Reviewer #1: This is a well reasoned, compelling, and important commentary on nomenclature of the caspases, paracaspases, and metacaspases. The authors are correct in pointing out that the names of the proteases themselves are problematic and have led to unfounded assertions and over-interpretations.

As an optional point, the authors might consider some discussion of the concept of induced proximity as it applies to caspase (and probably) paracaspase activation platforms, and whether this principle applies to metacaspases. In the context of apoptosis, induced proximity by activation platforms is the basis of the signaling pathways that orchestrate cell death; understanding when and how these can apply within the families might be helpful. Again, however, I think this is optional.

Response to reviewer:

proximity model for the activation of metacaspases, but are afraid that this interesting and important topic is beyond the scope of the classification and nomenclature document, that we have made as concise and dry as possible.

Classification and nomenclature of metacaspases and paracaspases: no more confusion with caspases

Elena A. Minina^{1,2,*}, Jens Staal³, Vanina E. Alvarez⁴, John A. Berges⁵, Ilana Berman-Frank⁶, Rudi Beyaert³, Kay D. Bidle⁷, Frédéric Bornancin⁸, Magali Casanova⁹, Juan J. Cazzulo⁴, Chang Jae Choi¹⁰, Nuria S. Coll¹¹, Vishva M. Dixit¹², Marko Dolinar¹³, Nicolas Fasel¹⁴, Christiane Funk¹⁵, Patrick Gallois¹⁶, Kris Gevaert¹⁷, Emilio Gutierrez-Beltran¹⁸, Stephan Hailfinger¹⁹, Marina Klemenčič¹³, Eugene V. Koonin²⁰, Daniel Krappmann²¹, Anna Linusson¹⁵, Maurício F. M. Machado²², Frank Madeo²³, Lynn A. Megeney²⁴, Panagiotis N. Moschou^{25,26,27}, Jeremy C. Mottram²⁸, Thomas Nyström²⁹, Heinz D. Osiewacz³⁰, Christopher M. Overall³¹, Kailash C. Pandey³², Jürgen Ruland^{33,34,35}, Guy S. Salvesen³⁶, Yigong Shi³⁷, Andrei Smertenko³⁸, Simon Stael^{17,39}, Jerry Ståhlberg¹, María Fernanda Suárez⁴⁰, Margot Thome¹⁴, Hannele Tuominen⁴¹, Frank Van Breusegem³⁹, Renier A. L. van der Hoorn⁴², Assaf Vardi⁴³, Boris Zhivotovsky^{44,45}, Eric Lam⁴⁶, Peter V. Bozhkov^{1,*}

¹Department of Molecular Sciences, Uppsala BioCenter, Swedish University of Agricultural Sciences and Linnean Center for Plant Biology, Uppsala, Sweden

²COS, Heidelberg University, Heidelberg, Germany

³VIB Center for Inflammation Research; Department of Biomedical Molecular Biology, Ghent University; Ghent, Belgium

⁴Instituto de Investigaciones Biotecnológicas, Universidad Nacional de San Martin, San Martin, Buenos Aires, Argentina

⁵Department of Biological Sciences and School of Freshwater Sciences, University of Wisconsin-Milwaukee, Milwaukee, WI, USA

⁶Department of Marine Biology, Leon H. Charney School of Marine Sciences, University of Haifa, Israel

⁷Department of Marine and Coastal Sciences, Rutgers University, New Brunswick, NJ, USA

⁸Novartis Institutes for BioMedical Research, Basel, Switzerland

⁹Aix-Marseille Univ, CNRS, LISM, Institut de Microbiologie de la Méditerranée, Marseille, France

¹⁰The University of Texas at Austin, Marine Science Institute, Port Aransas, TX, USA

- ¹¹Centre for Research in Agricultural Genomics (CRAG), CSIC-IRTA-UAB-UB, Campus UAB, Bellaterra, Barcelona, Spain
- ¹²Department of Physiological Chemistry, Genentech, South San Francisco, CA, USA
- ¹³University of Ljubljana, Faculty of Chemistry and Chemical Technology, Ljubljana, Slovenia
- ¹⁴Department of Biochemistry, University of Lausanne, Epalinges, Switzerland
- ¹⁵Department of Chemistry, Umeå University, Umeå, Sweden
- ¹⁶Faculty of Biology, Medicine and Health, University of Manchester, Manchester, UK
- ¹⁷VIB Center for Medical Biotechnology; Department of Biomolecular Medicine, Ghent University; Ghent, Belgium
- ¹⁸Instituto de Bioquímica Vegetal y Fotosíntesis, Universidad de Sevilla and Consejo Superior de Investigaciones Científicas, Sevilla, Spain
- ¹⁹Interfaculty Institute for Biochemistry, Eberhard Karls University, Tübingen, Germany
- ²⁰National Center for Biotechnology Information, National Library of Medicine, Bethesda, MD, USA
- ²¹Research Unit Cellular Signal Integration, Helmholtz Zentrum München German Research Center for Environmental Health, Neuherberg, Germany
- ²²Interdisciplinary Center for Biochemical Research, University of Mogi das Cruzes, Mogi das Cruzes, Brazil
- ²³Institute of Molecular Biosciences, NAWI Graz, University of Graz; BioTechMed Graz, Graz, Austria
- ²⁴Sprott Centre for Stem Cell Research, Ottawa Hospital Research Institute and Departments of Medicine and Cellular and Molecular Medicine, University of Ottawa, Ottawa, ON, Canada
- ²⁵Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology Hellas, Heraklion, Greece
- ²⁶Department of Biology, University of Crete, Greece
- ²⁷Department of Plant Biology, Uppsala BioCenter, Swedish University of Agricultural Sciences and Linnean Center for Plant Biology, Uppsala, Sweden
- $^{28} York\ Biomedical\ Research\ Institute,\ Department\ of\ Biology,\ University\ of\ York,\ York,\ UK$
- ²⁹Institute for Biomedicine, Sahlgrenska Academy, Centre for Ageing and Health AgeCap, University of Gothenburg, Gothenburg, Sweden

- ³⁰Institute for Molecular Biosciences, Faculty of Biosciences, Goethe University, Frankfurt/Main, Germany
- ³¹Departments of Oral Biological and Medical Sciences / and Biochemistry and Molecular Biology, Life Sciences Institute, University of British Columbia, Vancouver, BC, Canada
- ³² Protein Biochemistry and Engineering Laboratory, ICMR-National Institute of Malaria Research, New Delhi, India
- ³³Institute of Clinical Chemistry and Pathobiochemistry, School of Medicine, Technical University of Munich, Munich, Germany
- ³⁴German Cancer Consortium (DKTK), partner site Munich, Germany
- ³⁵German Center for Infection Research (DZIF), partner site Munich, Germany
- ³⁶Sanford Burnham Prebys Medical Discovery Institute, La Jolla, CA, USA
- ³⁷School of Life Sciences, Westlake University, Xihu District, Hangzhou Zhejiang Province, China
- ³⁸Institute of Biological Chemistry, Washington State University, Pullman, WA, USA
- ³⁹Department of Plant Biotechnology and Bioinformatics, Ghent University; VIB-UGent Center for Plant Systems Biology; Ghent, Belgium
- ⁴⁰Departamento de Biologia Molecular y Bioquimica, Facultad de Ciencias, Universidad de Malaga, Campus de Teatinos, Malaga, Spain
- ⁴¹Umeå Plant Science Centre, Department of Plant Physiology, Umeå University, Umeå, Sweden
- ⁴²Department of Plant Sciences, University of Oxford, Oxford, UK

USA

- ⁴³Department of Plant and Environmental Sciences, Weizmann Institute of Science, Rehovot, Israel
- ⁴⁴Division of Toxicology, Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden
- ⁴⁵Faculty of Fundamental Medicine, MV Lomonosov Moscow State University, Moscow, Russia
 ⁴⁶Department of Plant Biology, Rutgers the State University of New Jersey, New Brunswick, NJ
- *Correspondence: Peter.Bozhkov@slu.se (P.V.B), Alena.Minina@slu.se (E.A.M.)

Metacaspases and paracaspases are proteases that were first identified as containing a caspase-like structural fold (Uren et al., 2000). Like caspases, meta- and paracaspases are multifunctional proteins regulating diverse biological phenomena, such as aging, immunity, proteostasis and programmed cell death. The broad phylogenetic distribution of meta- and paracaspases across all kingdoms of life and large variation of their biochemical and structural features complicate classification and annotation of the rapidly growing number of identified homologs. Establishment of an adequate classification and unified nomenclature of meta- and paracaspases is especially important to avoid frequent confusion of these proteases with caspases - a tenacious misnomer that unfortunately does not appear to decline with time. This letter represents a consensus opinion of researchers studying different aspects of caspases, meta- and paracaspases in various organisms, ranging from microbes to plants and animals.

Classification of meta- and paracaspases

The current classification of proteases provided by the MEROPS database clusters caspases, metaand paracaspases to the same family, C14, within the CD clan (https://www.ebi.ac.uk/merops/). All members of C14 family are annotated to possess aspartate P1 cleavage specificity, and the family is further split into two subfamilies: C14A (caspases) and C14B (meta- and paracaspases).

Importantly, the MEROPS approach of grouping proteases into families or subfamilies is based on statistically significant similarities of the amino acid sequence within the peptidase domain or part thereof, without considering their biochemical properties (Rawlings et al., 2018). Being valuable for high-throughput protease classification, this approach, however, has substantial drawbacks if implemented without further adjustment. Indeed, in contradiction with the MEROPS description, none of the meta- or paracaspases characterized so far cleave after an aspartate residue. Instead, paracaspases are arginine-specific (Coornaert et al., 2008; Hachmann et al., 2012; Rebeaud et al., 2008), whereas metacaspases

caspase-specific probes for studying meta- and paracaspases that is commonly found in the literature and leads to false conclusions.

Apart from substrate specificity, caspases, meta- and paracaspases feature other fundamental differences (**Figure S1A**). For example, active metacaspases are monomers and their activation usually requires millimolar concentrations of calcium (Hander et al., 2019; McLuskey et al., 2012; Wong et al., 2012). In contrast, active caspases and paracaspases are calcium-independent dimers (Hachmann et al., 2012; Weismann et al., 2012; Yu et al., 2011). This indicates that upstream pathways regulating activation of caspases, meta- and paracaspases are likewise different.

In the past two decades we have learned about important differences between caspases, meta- and paracaspases. Thus, simple extrapolation of features typical for caspases to all other members of

numerals (e.g. type I metacaspases). As for the conserved protein structures, they will be referred to as the p20-like region, the p10-like region, the linker region and the N-terminal pro-domain, matching the nomenclature of caspases (**Figure S1A**; Alnemri et al., 1996). The p20, p10 and linker regions have been previously defined for the caspase group of the C14 family (Fuentes-Prior and Salvesen, 2004) and can be easily identified in meta- and paracaspase homologs based on a hidden Markov model (HMM) alignment with the C14 peptidase domain (**Figure S1B**). Notably, although not always clearly stated in the literature, most known members of the C14 family contain the linker region. Furthermore, type II metacaspases are distinguished by a long linker between the p20 and p10 regions and an additional linker within the p10 region (**Figure S1A**), which are frequently referred to as a single linker.

We suggest to consider the active form of meta- or paracaspases being a monomer if it is a cleaved or intact polypeptide chain derived from a single translational event, and a dimer if it comprises uncut or processed products of two translational events.

We propose to establish a unified nomenclature of meta- and paracaspases in order (i) to facilitate comparison of orthologs from different organisms and (ii) to make it suitable for annotating homologs of species with partially sequenced genomes. Thus, we suggest using simple root symbols such as MCA for metacaspases and PCA for paracaspases. When naming individual family members, these root symbols will be preceded by the abbreviated Latin name of the species and followed by a hyphen, Latin number representing the type and then a small alpha character indicating in alphabetical order the number of the homolog of this type in a given genome (**Figure S1C**). Proenzymes that require proteolytic processing for activation could be annotated with a prefix "pro-", e.g. pro-AtMCA-Ia for the metacaspase 1 of type I from

classification and unified nomenclature of meta- and paracaspases will facilitate a more comprehensive exchange of relevant findings within the scientific community and help to bridge already existing knowledge with newly discovered homologs, thus promoting mechanistic understanding of these ancient, evolutionarily conserved proteases.

Supplemental Information

Supplemental Information includes one figure and can be found with this article online at

Acknowledgements

This work was supported by Knut and Alice Wallenberg Foundation. We apologize to colleagues whose work has not been cited due to space limitation.

References

Alnemri, E., Livingston, D., Nicholson, D., Salvesen, G., Thornberry, N., Wong, W., and Yuan, J. (1996). Cell 87, 171.

Coornaert, B., Baens, M., Heyninck, K., Bekaert, T., Haegman, M., Staal, J., Sun, L., Chen, Z.J., Marynen, P., and Beyaert, R. (2008). Nat. Immunol. 9, 263–271.

Fuentes-Prior, P., and Salvesen, G.S. (2004). Biochem. J. 384, 201–232.

Hachmann, J., Snipas, S.J., Van Raam, B.J., Cancino, E.M., Houlihan, E.J., Poreba, M., Kasperkiewicz, P., Drag, M., and Salvesen, G.S. (2012). Biochem. J. 443, 287–295.

Hander, T., Fernández-Fernández, Á.D., Kumpf, R.P., Willems, P., Schatowitz, H., Rombaut, D., Staes, A., Nolf, J., Pottie, R., Yao, P., et al. (2019). Science *363*, 1–10.

Klemenčič, M., Novinec, M., and Dolinar, M. (2015). Mol. Microbiol. 98, 142–150.

McLuskey, K., Rudolf, J., Proto, W.R., Isaacs, N.W., Coombs, G.H., Moss, C.X., and Mottram, J.C. (2012). Proc. Natl. Acad. Sci. U. S. A. 109, 7469–7474.

Rawlings, N.D., Barrett, A.J., Thomas, P.D., Huang, X., Bateman, A., and Finn, R.D. (2018). Nucleic Acids Res. 46, D624–D632.

Rebeaud, F., Hailfinger, S., Posevitz-Fejfar, A., Tapernoux, M., Moser, R., Rueda, D., Gaide, O., Guzzardi, M., Iancu, E.M., Rufer, N., et al. (2008). Nat. Immunol. 9, 272–281.

Sundström, J.F., Vaculova, A., Smertenko, A.P., Savenkov, E.I., Golovko, A., Minina, E., Tiwari, B.S., Rodriguez-Nieto, S., Zamyatnin Jr., A.A., Välineva, T., et al. (2009). Nat. Cell Biol. *11*, 1347-1354.

Uren, A., O'Rourke, K., Aravind, L., Pisabarro, M.T., Seshagiri, S., Koonin, E. V, and Dixit, V.M. (2000). Mol. Cell *6*, 961–967.

Vercammen, D., De Cotte, B. Van, De Jaeger, G., Eeckhout, D., Casteels, P., Vandepoele, K., Vandenberghe, I., Van Beeumen, J., Inzé, D., and Van Breusegem, F. (2004). J. Biol. Chem. 279, 45329–45336.

Wiesmann, C., Leder, L., Blank, J., Bernardi, A., Melkko, S., Decock, A., D'Arcy, A., Villard, F., Erbel, P., Hughes, N., et al. (2012). J. Mol. Biol. 419, 4–21.

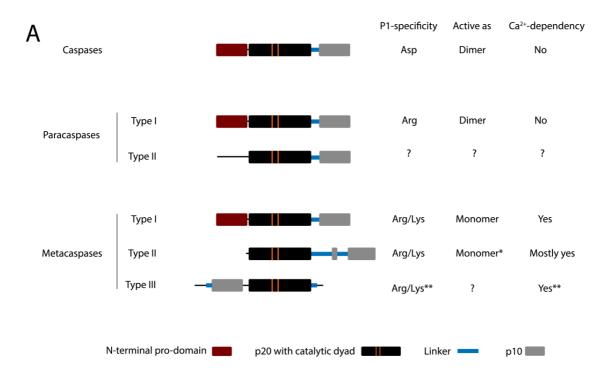
Wong, A.H.H., Yan, C., and Shi, Y. (2012). J. Biol. Chem. 287, 29251–29259.

Yu, J.W., Jeffrey, P.D., Ha, J.Y., Yang, X., and Shi, Y. (2011). Proc. Natl. Acad. Sci. U. S. A. 108, 21004–21009.

M lec la Cell
S le e al I f a i

Claificai ad e cla e f e aca a e a d a aca a e: e c f i i h ca a e

Ele a A. Mi i a, Je Saal, Va i a E. Al a e, Jh A. Be ge, Ila a Be a -F a k, R di Be ae, Ka D. Bidle, F d ic B a ci, Magali Ca a a, Ja J. Ca l, Cha g Jae Chi, N ia S. Cll, Vih a M. Dii, Mak Dlia, Nic la Fael, Chi ia e F k, Paick Galli, Ki Ge ae, Eili Gie e-Bel a, Seha Hailfige, Maia V Ege e V. Ki, Daiel Kaa, AaLi, Maci F. M. Machad, Fak Made, L A. Mege e, Paagii N. Mch, Je e C. Ma, ThaN, Hei D. Oie ac, Chi he M. Oe all, Kailah C. Pade, Jge R lad, GS. Sal ee, Yig g Shi, Adei Seek, Si Sael, Je Shlbeg, Maa Feada Se, Mag The, Haele Tie, Fak Va Beege, Reie A. L. ade H, Aaf Vadi, Bi Zhi k, Eic La, Pee V. Bhk



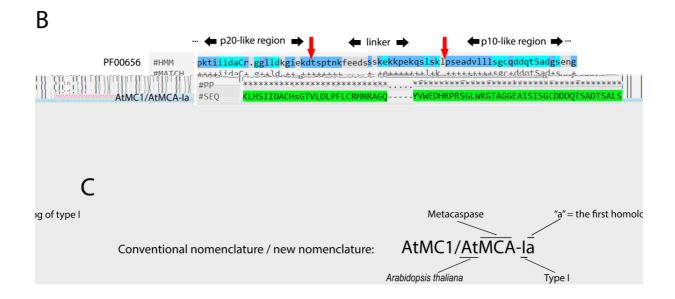


Fig e S1. Cla ifica i a d e cla e f e a- a d a aca a e .

(A) Comparison between caspases, metacaspases and paracaspases: domain composition and biochemical characteristics. *Bozhkov and Smertenko, unpublished data for mcII-Pa/PaMCA-IIb from *Picea abie* (Minina et al., 2017; Suarez et al., 2004). **Only two orthologs of the type III metacaspases have been characterized so far, GtMC2/GtMCA-IIIa from *G illa dia he a* (Klemenčič and Funk, 2018) and PtMC5/PtMCA-IIIc from *Phae dac l ic* (van

Creveld et al., 2018). (**B**) Part of the HMM alignment of the *A abid i halia a* metacaspase 1 with the C14 peptidase domain (PF00656), red arrows indicate borders between the p20-like region, linker and the p10-like region. (**C**) An example of the use of the new nomenclature for the *A. halia a* type I metacaspase. For homologs with well-established names we recommend to use the new nomenclature synonymously; this will significantly ease comparison with orthologs from species with partially sequenced genome.

S le e al Refe e ce

Klemenčič, M., and Funk, C. (2018). New Phytol. 218, 1179–1191.

Minina, E.A., Coll, N.S., Tuominen, H., and Bozhkov, P. V. (2017). Cell Death Differ. 24, 1314–1325.

Suarez, M.F., Filonova, L.H., Smertenko, A., Savenkov, E.I., Clapham, D.H., Von Arnold, S., Zhivotovsky, B., and Bozhkov, P. V. (2004). Curr. Biol. *14*, 339–340.

van Creveld, SG., Ben-Dor, S., Mizrachi, A., Alcolombri, U., Hopes, A., Mock, T., Rosenwasser, S., and Vardi, A. (2018). bioRxiv doi: https://doi.org/10.1101/444109

