## Supplementary Table 1. Interaction of COI1 and TIR1 with SCF subunits

	TIR1	COI1
ASK	<b>Direct Interaction: (positive)</b> The Arabidopsis TIR1- ASK1 complex was crystallized, revealing interaction with ASK1 at the N-terminal F-Box domain (Tan <i>et al.</i> , 2007).	<b>Direct Interaction: (positive)</b> The Arabidopsis COI1-ASK1 complex was crystallized, demonstrating that ASK1 interacts with COI1 at the N-terminal F-Box domain (Sheard <i>et al.</i> , 2010).
	<ul> <li>Stability: (positive) Steady-state levels of TIR1 increase in the presence of ASK1 (Dezfulian <i>et al.</i>, 2016).</li> <li>Auxin Response: (positive) The <i>ask1-1</i> mutant has</li> </ul>	<b>Stability: (positive)</b> <i>ask1-1</i> mutants have reduced levels of COI1 in immunoblot analysis, using $\alpha$ -COI1; whereas ASK1 being present in a yeast pBridge expression system leads to an increased COI1 protein levels (Yan <i>et al.</i> , 2013).
	a reduced auxin response (Gray <i>et al.,</i> 1999).	<b>JA Response: (positive)</b> The <i>ask1-1</i> mutant has reduced expression levels of <i>vsp2</i> in the presence of MeJA (Yan <i>et al.</i> , 2013).
CULLIN	<ul> <li>Direct Interaction: (positive) Immunoprecipitates and subsequent western blot analysis provide evidence that TIR1 interacts with CUL1 through TIR1's N-terminal H1 helix (Yu <i>et al.</i>, 2015).</li> <li>Stability: (negative) Inhibition of CUL1 interaction leads to increased TIR1 protein levels based on comparing histochemical staining in tagged GUS mutant and GUS wild-type lines (Yu <i>et al.</i>, 2015).</li> <li>Auxin Response: (positive) The <i>cul1-7</i> mutant is less responsive to 2,4-D (synthetic auxin) treatment compared to the wild type and also degrades IAA1-</li> </ul>	<ul> <li>Direct Interaction: (?) The interaction is assumed to be indirect, via the SCF<sup>COI1</sup> complex. Although, it cannot be ruled out that COI1 may interact directly with CUL1, through its F-Box H1 helix (Zheng <i>et al.</i>, 2002b). CUL1 is able to be co-immunoprecipitated with COI1 (Xu <i>et al.</i>, 2002).</li> <li>Stability: (positive) The <i>axr6-1/+</i> and <i>axr6-2/+</i> CUL1 heterozygous mutants show reduced COI1 protein levels (Yan <i>et al.</i>, 2013).</li> <li>JA Response: (positive) When treated with MeJA, AtCUL1 mutants, <i>axr6-1</i> and <i>axr6-2</i>, show a reduced expression of JA responsive genes compared to wild-</li> </ul>
	LUC at a slower rate (Gilkerson <i>et al.,</i> 2009).	type lines (Ren <i>et al.,</i> 2005).
RBX	<b>Direct Interaction: (negative)</b> Indirect Interaction with RBX1 through CULLIN1 of the SCF <sup>TIR1</sup> complex (Gray <i>et al.</i> , 2002).	<b>Direct Interaction: (negative)</b> COI1 indirectly interacts with RBX1 when assembled into the SCF <sup>COI1</sup> complex (Zheng <i>et al.</i> , 2002b).
	<b>Stability: (?)</b> We could not find any substantial evidence that RBX1 changes the TIR1 protein levels.	<b>Stability: (?)</b> We could not find any substantial evidence that RBX1 changes the COI1 protein levels.
	<b>Auxin Response: (dynamic)</b> As part of the SCF complex, RBX1 facilitates substrate ubiquitination (Zheng <i>et al.</i> , 2002a). However, overexpression of <i>RBX1</i> leads to a reduced auxin response (Gray <i>et al.</i> , 2002).	<b>JA Response: (positive)</b> when induced, <i>AtkDx1</i> (RNAI) mutants show a reduction in the expression of JA responsive genes (Xu <i>et al.</i> , 2002). In <i>RBX1</i> overexpression lines, the JA response is unaffected (Gray <i>et al.</i> , 2002).

ALF4	<ul> <li>Direct Interaction: (negative) Indirect interaction through the SCF<sup>TIR1</sup> complex. ALF4 interacts with RBX1 in Y2H and co-immunoprecipitation experiments (Bagchi <i>et al.</i>, 2018).</li> <li>Stability: (negative) In an <i>alf4</i> mutant, there is an increase of TIR1 abundance (Bagchi <i>et al.</i>, 2018).</li> <li>Auxin Response: (negative) ALF4 reduces the ability of RBX1 and E2 to ubiquitinate substrates (Bagchi <i>et al.</i>, 2018).</li> </ul>	
CAND1	<ul> <li>Direct Interaction: (negative) The interaction is indirect through CULLIN (Zheng <i>et al.</i>, 2002a).</li> <li>Stability: (negative) Diminishing CULLIN-CAND1 interaction increases SCF<sup>TIR1</sup> abundance (Zhang <i>et al.</i>, 2008).</li> <li>Auxin Response: (dynamic) The loss-of-function mutants display a reduced auxin response, this is thought to be based on dynamic cycling being necessary for proper SCF function (Zhang <i>et al.</i>, 2008).</li> </ul>	<ul> <li>Direct Interaction: (negative) The interaction is indirect through CULLIN (Zheng <i>et al.</i>, 2002a).</li> <li>Stability: (no effect) There is no reduction in the COI1 protein levels in <i>cand1-1</i> mutants (Feng <i>et al.</i>, 2004).</li> <li>JA Response: (positive) The <i>cand1-1</i> mutant has some JA response defects, whereas the <i>coi1-1 cand1-1</i> double mutant is fully insensitive (Feng <i>et al.</i>, 2004).</li> </ul>
JAZ		<ul> <li>Direct Interaction: (positive)</li> <li>The COI1-ASK1-JAZ1 degron peptide complex was crystallized with either JA-Ile or COR (Sheard <i>et al.</i>, 2010). Furthermore COI1 has shown interaction with a number of JAZ proteins although interaction with JAZ7 and JAZ8, which lack the canonical Jaz motif, has not been reported to our knowledge (Pauwels and Goossens, 2011; Shyu <i>et al.</i>, 2012).</li> <li>Stability: (no effect) When COI1 protein is co-expressed with JAZ1, it does not lead to increased COI1 protein levels (Yan <i>et al.</i>, 2013).</li> <li>JA response: (negative) JAZ protein interaction with COI1 leads to their ubiquitination and subsequent degradation, which prompts the JA response (Chini <i>et al.</i>, 2007; Thines <i>et al.</i>, 2007).</li> </ul>

Aux/IAA	<ul> <li>Direct Interaction: (positive) TIR1-ASK1 complex was crystallized with Aux/IAA substrate peptides (Tan <i>et al.</i>, 2007).</li> <li>Stability: (?) We did not find experimental evidence for an effect of Aux/IAAs on the stability of TIR1 proteins.</li> <li>Auxin Response: (positive) Aux/IAA protein interaction with TIR1 leads to their degradation and de-repression of ARFs, enabling the auxin response (Tan <i>et al.</i>, 2007).</li> </ul>	
MED25		<ul> <li>Direct Interaction: (positive) MED25 immunoprecipitates with COI1 (An <i>et al.</i>, 2017).</li> <li>Stability: (positive) The <i>med25-4</i> mutant displays reduced COI1 levels at the promoters of JA responsive genes (An <i>et al.</i>, 2017).</li> <li>JA Response: (positive) <i>med25-4</i> mutants have a reduced ability to degrade JAZ proteins in the presence of JA-Ile (An <i>et al.</i>, 2017).</li> </ul>
Ubiquitination	<ul> <li>Direct Interaction: (positive) TIR1 was demonstrated to be polyubiquitinated (Stuttmann et al., 2009).</li> <li>Stability: (negative): Proteasome inhibitors stabilize TIR1 protein (Stuttmann et al., 2009; Yu et al., 2015).</li> <li>Auxin Response (negative) Once TIR1 is associated into the SCF<sup>TIR1</sup> complex results in eventual proteasomal degradation of TIR1 (Stuttmann et al., 2009; Yu et al., 2015).</li> </ul>	<ul> <li>Direct Interaction: (positive) COI1 is thought to be ubiquitinated on the K297 residue, leading to its degradation (Yan <i>et al.</i>, 2013).</li> <li>Stability: (negative) COI1 is thought to be ubiquitinated on K297, leading to its degradation. The 26 proteasome component mutant, <i>rpt5a-4</i>, has increased COI1 protein levels (Yan <i>et al.</i>, 2013).</li> <li>JA Response: (negative) The 26 proteasome component mutant, <i>rpt5a-4</i>, shows an enhanced JA sensitivity (Yan <i>et al.</i>, 2013).</li> </ul>
S-Nitrosylation	<ul> <li>Direct Interaction: (positive) Biotinylated TIR1 was detected in a biotin switch assays strongly supporting that a TIR1 residue (candidates being C140 or C480) has SNO modification (Terrile <i>et al.</i>, 2012).</li> <li>Stability: (no effect) Mutating the C140 residue did not result in a decrease in TIR1 protein stability (Dezfulian <i>et al.</i>, 2016).</li> <li>Auxin Response: (positive) An amino acid change of C140A led to abolishment of interaction with Aux/IAA proteins (Terrile <i>et al.</i>, 2012).</li> </ul>	

Rubbylation	<b>Direct Interaction: (negative)</b> Rubbylation occurs on CUL1 in the SCF complex (Gray <i>et al.,</i> 2002).	<b>Direct Interaction: (negative)</b> Rubbylation occurs on CUL1 in the SCF complex (del Pozo <i>et al.,</i> 2002; Gray <i>et al.,</i> 2002).
	Stability: (no effect) RUB modification does not	
	seem to affect the stability of the SCF complex (del Pozo <i>et al.,</i> 2002).	<b>Stability: (no effect)</b> RUB modification does not seem to affect the stability of the SCF complex (del Pozo <i>et al.,</i> 2002).
	Auxin Response: (dynamic) An axr1 mutant,	
	defective in RUB activation, lacks the ability for RUB modification of CUL1 and leads to decreases in the auxin response (del Pozo <i>et al.</i> , 2002). Overexpression of <i>RBX1</i> leads to hypermodification of CUL1 by RUB, which disrupts its function and leads to an impaired auxin response (Gray <i>et al.</i> , 2002).	<b>JA Response: (positive)</b> <i>axr1</i> mutants are slightly JA insensitive, indicating that RUB modification is necessary for the JA response (del Pozo <i>et al.</i> , 2002; Xu <i>et al.</i> , 2002). The <i>axr1-3</i> mutant shows a reduced JA root growth inhibition (Schwechheimer <i>et al.</i> , 2002).
COP9	<b>Direct Interaction: (negative)</b> COP9 interacts with SCF <sup>TIR1</sup> subunits and specifically interacts with AtCUL1 in a yeast two-hybrid analysis (Schwechheimer <i>et al.</i> , 2001). CSN forms a complex	<b>Direct Interaction: (negative)</b> CSN forms a complex with CULLIN (Cavadini <i>et al.</i> , 2016). COP9 interacts with SCF <sup>COI1</sup> (Feng <i>et al.</i> , 2003).
	with CULLIN (Cavadini et al., 2016).	<b>Stability: (no effect)</b> We could not find any experimental evidence demonstrating a change in COI1 protein
	Stability: (positive) A COP9 signalosome	abundance. In CSN mutants, the integrity of the SCF
	subunit mutant, <i>csn2-5</i> , has reduced TIR1 protein levels (Stuttmann <i>et al.</i> , 2009).	complex was not affected (Schwechheimer <i>et al.,</i> 2002).
		JA Response: (positive) Plants with antisense CSN5 show
	Auxin Response: (positive) Plants with reduced	a reduced JA response (Schwechheimer et al., 2002).
	COP9 show a reduced auxin response (Schwechheimer <i>et al.</i> , 2001).	Genome expression profiles demonstrate that functional COP9 is crucial for the JA response (Feng <i>et al.</i> , 2003).

## References

An C, Li L, Zhai Q, et al. 2017. Mediator subunit MED25 links the jasmonate receptor to transcriptionally active chromatin. Proceedings of the National Academy of Sciences of the United States of America **114**, E8930-E8939.

Bagchi R, Melnyk CW, Christ G, et al. 2018. The Arabidopsis ALF4 protein is a regulator of SCF E3 ligases. EMBO Journal 37, 255-268.

Cavadini S, Fischer ES, Bunker RD, et al. 2016. Cullin-RING ubiquitin E3 ligase regulation by the COP9 signalosome. Nature 531, 598-603.

Chini A, Fonseca S, Fernández G, et al. 2007. The JAZ family of repressors is the missing link in jasmonate signalling. Nature 448, 666-671.

del Pozo JC, Dharmasiri S, Hellmann H, Walker L, Gray WM, Estelle M. 2002. AXR1-ECR1–dependent conjugation of RUB1 to the Arabidopsis cullin AtCUL1 is required for auxin response. Plant Cell **14**, 421-433.

**Dezfulian MH, Jalili E, Roberto DKA, Moss BL, Khoo K, Nemhauser JL, Crosby WL.** 2016. Oligomerization of SCF<sup>TIR1</sup> is essential for Aux/IAA degradation and auxin signaling in Arabidopsis. PLoS Genetics **12**, e1006301.

**Feng S, Ma L, Wang X, Xie D, Dinesh-Kumar SP, Wei N, Deng XW.** 2003. The COP9 signalosome interacts physically with SCF<sup>COI1</sup> and modulates jasmonate responses. Plant Cell **15**, 1083-1094.

**Feng S, Shen Y, Sullivan JA, Rubio V, Xiong Y, Sun T-p, Deng XW.** 2004. Arabidopsis CAND1, an unmodified CUL1-interacting protein, is involved in multiple developmental pathways controlled by ubiquitin/proteasome-mediated protein degradation. Plant Cell **16**, 1870-1882.

**Gilkerson J, Hu J, Brown J, Jones A, Sun T-p, Callis J.** 2009. Isolation and characterization of *cul1-7*, a recessive allele of *CULLIN1* that disrupts SCF function at the C terminus of CUL1 in *Arabidopsis thaliana*. Genetics **181**, 945-963.

Gray WM, del Pozo JC, Walker L, et al. 1999. Identification of an SCF ubiquitin–ligase complex required for auxin response in Arabidopsis thaliana. Genes & Development 13, 1678-1691.

Gray WM, Hellmann H, Dharmasiri S, Estelle M. 2002. Role of the Arabidopsis RING-H2 protein RBX1 in RUB modification and SCF function. Plant Cell 14, 2137-2144.

Pauwels L, Goossens A. 2011. The JAZ proteins: a crucial interface in the jasmonate signaling cascade. Plant Cell 23, 3089-3100.

Ren C, Pan J, Peng W, et al. 2005. Point mutations in Arabidopsis *Cullin1* reveal its essential role in jasmonate response. Plant Journal 42, 514-524.

Schwechheimer C, Serino G, Callis J, Crosby WL, Lyapina S, Deshaies RJ, Gray WM, Estelle M, Deng X-W. 2001. Interactions of the COP9 signalosome with the E3 ubiquitin ligase SCF<sup>TIR1</sup> in mediating auxin response. Science **292**, 1379-1382.

Schwechheimer C, Serino G, Deng X-W. 2002. Multiple ubiquitin ligase-mediated processes require COP9 signalosome and AXR1 function. Plant Cell 14, 2553-2563.

Sheard LB, Tan X, Mao H, et al. 2010. Jasmonate perception by inositol-phosphate-potentiated COI1–JAZ co-receptor. Nature 468, 400–405.

Shyu C, Figueroa P, DePew CL, Cooke TF, Sheard LB, Moreno JE, Katsir L, Zheng N, Howe GA. 2012. JAZ8 lacks a canonical degron and has an EAR motif that mediates transcriptional repression of jasmonate responses in *Arabidopsis*. Plant Cell **24**, 536-550.

Stuttmann J, Lechner E, Guérois R, Parker JE, Nussaume L, Genschik P, Noël LD. 2009. COP9 signalosome- and 26S proteasomedependent regulation of SCF<sup>TIR1</sup> accumulation in *Arabidopsis*. Journal of Biological Chemistry **284**, 7920-7930.

Tan X, Calderon-Villalobos LIA, Sharon M, Zheng C, Robinson CV, Estelle M, Zheng N. 2007. Mechanism of auxin perception by the TIR1 ubiquitin ligase. Nature 446, 640-645.

**Terrile MC, París R, Calderón-Villalobos LIA, Iglesias MJ, Lamattina L, Estelle M, Casalongué CA.** 2012. Nitric oxide influences auxin signaling through *S*-nitrosylation of the Arabidopsis TRANSPORT INHIBITOR RESPONSE 1 auxin receptor. Plant Journal **70,** 492-500.

**Thines B, Katsir L, Melotto M**, et al. 2007. JAZ repressor proteins are targets of the SCF<sup>CO11</sup> complex during jasmonate signalling. Nature **448**, 661-665.

Xu L, Liu F, Lechner E, Genschik P, Crosby WL, Ma H, Peng W, Huang D, Xie D. 2002. The SCF<sup>COI1</sup> ubiquitin-ligase complexes are required for jasmonate response in Arabidopsis. Plant Cell **14**, 1919-1935.

Yan J, Li H, Li S, Yao R, Deng H, Xie Q, Xie D. 2013. The *Arabidopsis* F-box protein CORONATINE INSENSITIVE1 is stabilized by SCF<sup>COI1</sup> and degraded via the 26S proteasome pathway. Plant Cell **25**, 486-498.

Yu H, Zhang Y, Moss BL, Bargmann BOR, Wang R, Prigge M, Nemhauser JL, Estelle M. 2015. Untethering the TIR1 auxin receptor from the SCF complex increases its stability and inhibits auxin response. Nature Plants 1, 14030.

**Zhang W, Ito H, Quint M, Huang H, Noël LD, Gray WM.** 2008. Genetic analysis of CAND1-CUL1 interactions in *Arabidopsis* supports a role for CAND1-mediated cycling of the SCF<sup>TIR1</sup> complex. Proceedings of the National Academy of Sciences of the United States of America **105**, 8470-8475.

Zheng J, Yang X, Harrell JM, et al. 2002a. CAND1 binds to unneddylated CUL1 and regulates the formation of SCF ubiquitin E3 ligase complex. Molecular Cell 10, 1519-1526.

**Zheng N, Schulman BA, Song L, et al.** 2002b. Structure of the Cul1-Rbx1-Skp1-F box<sup>Skp2</sup> SCF ubiquitin ligase complex. Nature **416,** 703-709.