

**DETERMINATION OF *IN VITRO* AND *IN SILICO* INDEXES FOR THE MODELLING OF
BLOOD-BRAIN BARRIER PARTITIONING OF DRUGS VIA MICELLAR AND
IMMOBILIZED ARTIFICIAL MEMBRANE LIQUID CHROMATOGRAPHY.**

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ABSTRACT

In the present work, 79 structurally unrelated analytes were taken into account and their chromatographic retention coefficients, measured by Immobilized Artificial Membrane Liquid Chromatography (IAM-LC), and by Micellar Liquid Chromatography (MLC) employing sodium dodecyl sulfate (SDS) as surfactant, were determined. Such indexes, along with topological and physico-chemical parameters calculated *in silico*, were subsequently used for the development of Blood-Brain Barrier passage-predictive statistical models using partial least square (PLS) regression.. Highly significant relationships were observed either using IAM ($r^2 (n-1) = 0.78$) or MLC ($r^2 (n-1) = 0.83$) derived indexes along with *in silico* descriptors. This hybrid approach proved fast and effective in the development of highly predictive BBB passage oriented models and, therefore, it can be of interest for pharmaceutical industries as a high-throughput BBB penetration oriented screening method. Finally, it shed new light into the molecular mechanism involved in the BBB uptake of therapeutics.

Keywords: Immobilized Artificial Membrane; Micellar Liquid Chromatography; Blood-Brain Barrier passage; Quantitative structure–activity relationships.

1.0 INTRODUCTION

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35 Pharmaceutical drug development is still a highly inefficient process: over one third of the failures
36 in drug candidate development is estimated to occur due to unsatisfactory pharmacokinetic
37 properties¹, mainly regarding absorption, metabolism and toxicity and the attrition rates for Central
38 Nervous System (CNS) active drugs are even higher². In fact, before reaching the blood circulation,
39 a drug diffuses through the biological barriers separating the circulating blood from the interstitial
40 fluid that surrounds the tissues. For orally administered drugs, this barrier is the intestinal
41 epithelium whereas the passage of drugs designed to act at the CNS level is further regulated by the
42 Blood-Brain Barrier (BBB). The BBB is one of the most complex and extensively studied
43 biological barriers, and its function is to preserve mammalian brain integrity against possible
44 injurious substances. It is made of endothelial cells, narrowly adherent one to the other to form tight
45 junctions, restricting the passage of solutes^{3,4}. Indeed, drug transport is strongly limited by this
46 peculiar biological structure to pure passive transcellular diffusion. In fact, the paracellular route,
47 i.e. the passage of actives through the gaps between each endothelial cell, is completely hindered.
48 Apart from active transport mechanisms, whose occurrence is difficult to predict on a solely
49 chemical structure basis, drugs can therefore cross the BBB only by the passive transcellular route.
50 Plenty of *in vivo*, *ex vivo*, and *in vitro* methods are available for measuring BBB partitioning of
51 analytes. Historically, one of the most used and reputed method is the determination of log BB
52 values⁵. Log BB is defined as:

53

$$\log BB = \log \frac{C_{Brain}}{C_{Blood}}$$

55

56 in which C_{Brain} is the concentration that the analyte realizes in the brain tissues, and C_{Blood} is the
57 concentration that it realizes in the blood. However, this method involves the use of animal models,

usually rodents, and does not provide any mechanistic information about the nature of the passage; furthermore, the method is time-consuming and potential source of ethical issues.

Methods based on the employment of cultured cell lines can also be effective; however, astrocytes cell cultures are often difficult to grow and recreating an *in vitro* environment similar to the *in vivo* BBB can be challenging even for the most experienced scientists. Caco-2 model based methods may also be an alternative; however, apart from the structural dissimilarities with the other cell cultures⁶, they are difficult to standardize, complicating comparisons of data determined in different laboratories.

In silico methods, generally based on the calculation of physico-chemical parameters, yield various advantages. They are much faster to perform, allowing the screening of large libraries of compounds (even solutes not yet synthesized); in addition, they can assist in the elucidation of the molecular mechanisms involved in membrane permeation. However, they also suffer from several limitations including the aspect that they are unable to take into account all phenomena actually occurring *in vivo*⁷.

In vitro methods based on the use of biomimetic stationary phases coupled with high performance liquid chromatography (HPLC) have been used to surrogate BBB permeation data^{8,9}. The main advantages are that they are much more reproducible and easier to perform and, albeit conceptually simple, they can be incidentally able to provide an in-depth understanding of the mechanisms involved in membrane barrier passage. Such biomimetic stationary phases include, for instance, Immobilized Artificial Membranes (IAM). IAM stationary phases are based on analogues of phosphatidylcholine, which is the major component of biological membranes, and chromatographic retention coefficients of the analytes on such stationary phases are assumed as direct measures of their phospholipophilicity¹⁰, i.e. their affinity for phospholipids. Such measures have been proven to be able to mirror various phenomena underlying membrane barrier passage^{8,11–13}. In addition, other chromatographic indexes, whose drug BBB-penetration predictivity has been demonstrated^{14–16}, include those achieved by Micellar Liquid Chromatography (MLC) technique. This technique is

84 based on the addition of surfactants to an aqueous mobile phase at concentrations higher than their
85 critical micelle concentrations (CMC)¹⁷ resulting in the formation of micelles acting as a partition
86 phase. Both IAM and MLC chromatographic indexes, mainly if combined with *in silico* calculated
87 descriptors, have demonstrated effectiveness in the prediction of BBB drug penetration¹⁶. However,
88 the methods proposed are still too time-consuming to meet the demands of pharmaceutical
89 companies and their suitability should be confirmed on larger set of analytes.

90 The aim of the present work has been the development of drug BBB penetration oriented statistical
91 models based on analytical indexes, achieved on biomimetic conditions by medium/high-
92 throughput methods, along with *in silico* calculated descriptors. To the best of our knowledge, this
93 is the study based on the highest number of compounds among those employing IAM and MLC
94 data to predict drug pharmacokinetic properties.

95 Therefore, particular attention is set on:

- 96 *i)* the setup of medium/high-throughput methods for the achievement of both IAM and MLC
97 indexes;
98 *ii)* the validation of such parameters by developing statistical models for the prediction of BBB
99 penetration (log BB) by using the chromatographic indexes along with *in silico* calculated
100 descriptors;
101 *iii)* the elucidation of the molecular mechanism involved in BBB passive diffusion of drugs;
102 *iv)* the possibility of taking into account, by molecular docking studies, the occurrence of active
103 efflux mechanisms.

104 In the present work, 79 structurally unrelated analytes have been taken into account and their
105 chromatographic retention coefficients, measured by high-throughput IAM-LC and MLC methods,
106 the latter employing sodium dodecyl sulfate (SDS) as surfactant, were determined. Such indexes
107 have subsequently been used for the development of BBB-passage predictive statistical models
108 using partial least squares (PLS) automatic regression along with physico-chemical parameters,
109 calculated *in silico*. Such hybrid approach was aimed at combining the speediness in the

110 achievement of computational chemistry derived topological and physico-chemical parameters with
111 the improved predictivity of the *in vitro* methods.

112

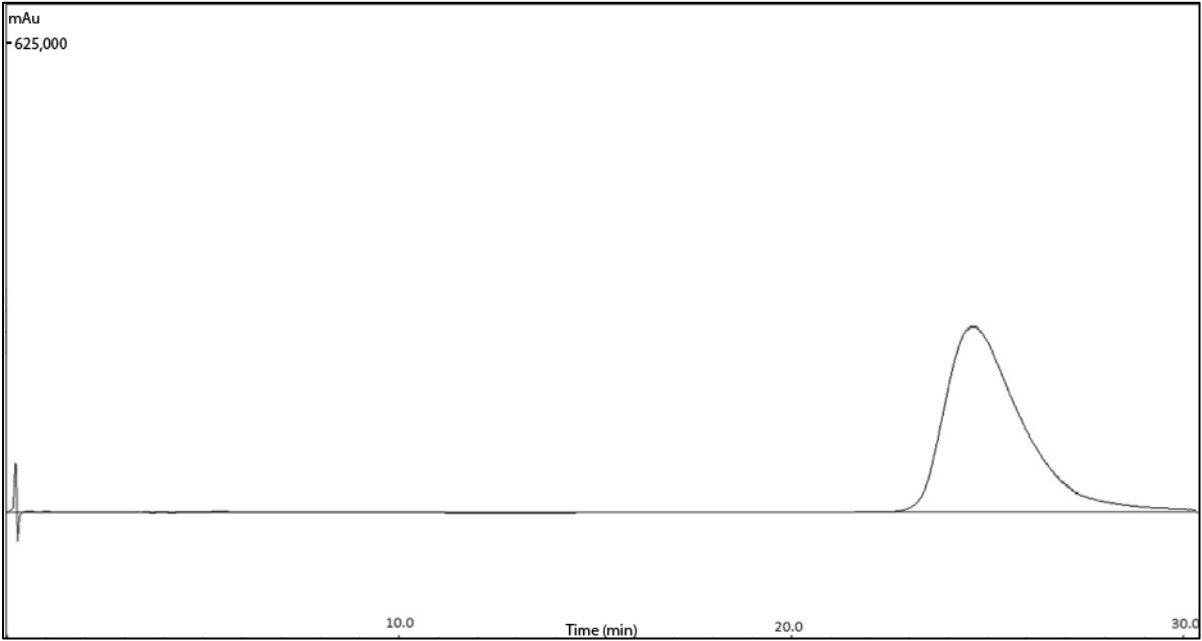
113 2.0 RESULTS AND DISCUSSION

114 The IAM-LC and MLC chromatographic retention coefficients, as well as the pKa, log BB values,
115 UV wavelengths of the experimental determinations and suppliers, are presented in Table 1. In
116 MLC, the highest retained compound (triprolidine) eluted within 33.0 minutes, whereas in IAM-LC
117 the maximum run time was 37.0 minutes (fluphenazine). However, by performing either the MLC
118 or the IAM-LC analytical methods, 85% of the compounds of the dataset eluted within 15.0 minutes
119 and a preliminary estimate, as an order of magnitude, of the retention times expected can be easily
120 performed based on the calculation of $\log D^{7.4}$ values of each compound present in the dataset. Two
121 chromatographic runs for each technique are reported in Figure 1 (MLC) and Figure 2 (IAM-LC).
122 The log BB values span a very large range (from -2.00 to +1.51) as the analytes to be included in
123 the dataset were selected to include both CNS inactive (e.g. norfloxacin, nitrofurantoin) and CNS
124 active (e.g. fluphenazine, desipramine) drugs. The P-gp affinities, expressed in $\text{kcal}\cdot\text{mol}^{-1}$, of the
125 drugs considered are listed in Table 2A and Table 2B. They were incorporated in each of the
126 following steps to model even the BBB passage of analytes undergoing P-gp efflux mechanisms.

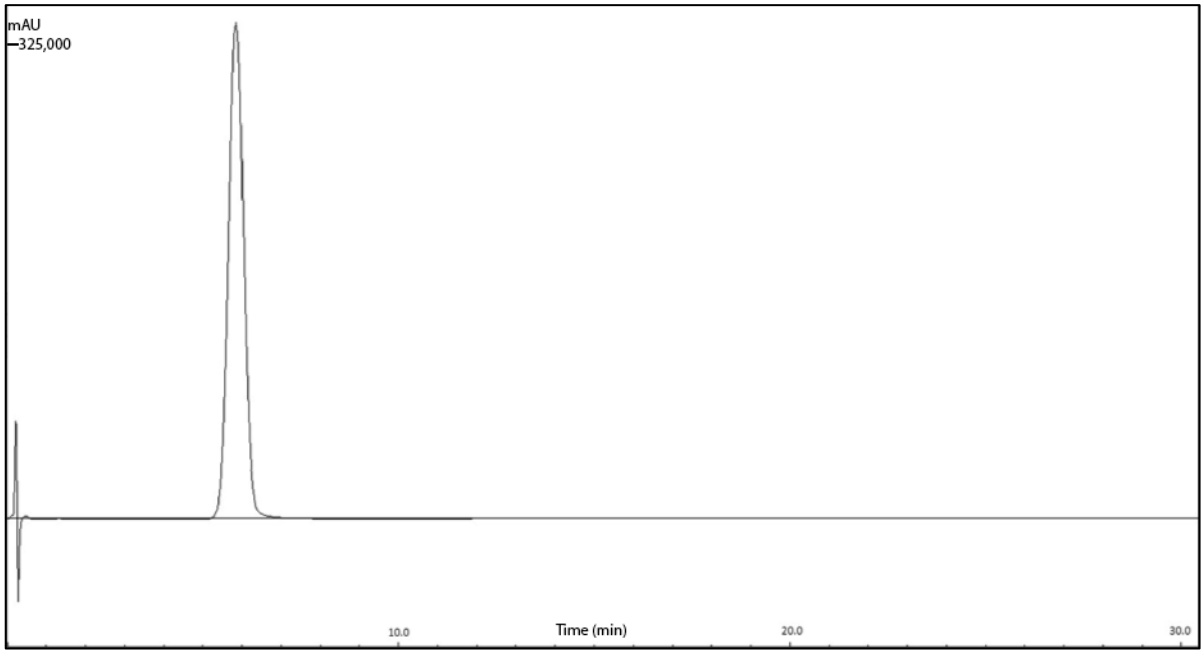
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129 **Figure 1.** MLC chromatograms achieved for the analytes A) Imipramine ($50\text{ }\mu\text{g mL}^{-1}$ in methanol)
130 and B) Ethylbenzene ($50\text{ }\mu\text{g mL}^{-1}$ in methanol) employing a mobile phase consisting of an aqueous
131 solution of $0.05\text{ mol}\cdot\text{L}^{-1}$ sodium dodecyl sulfate. The detailed experimental conditions are reported
132 in paragraph 4.0.

133
134
135 A



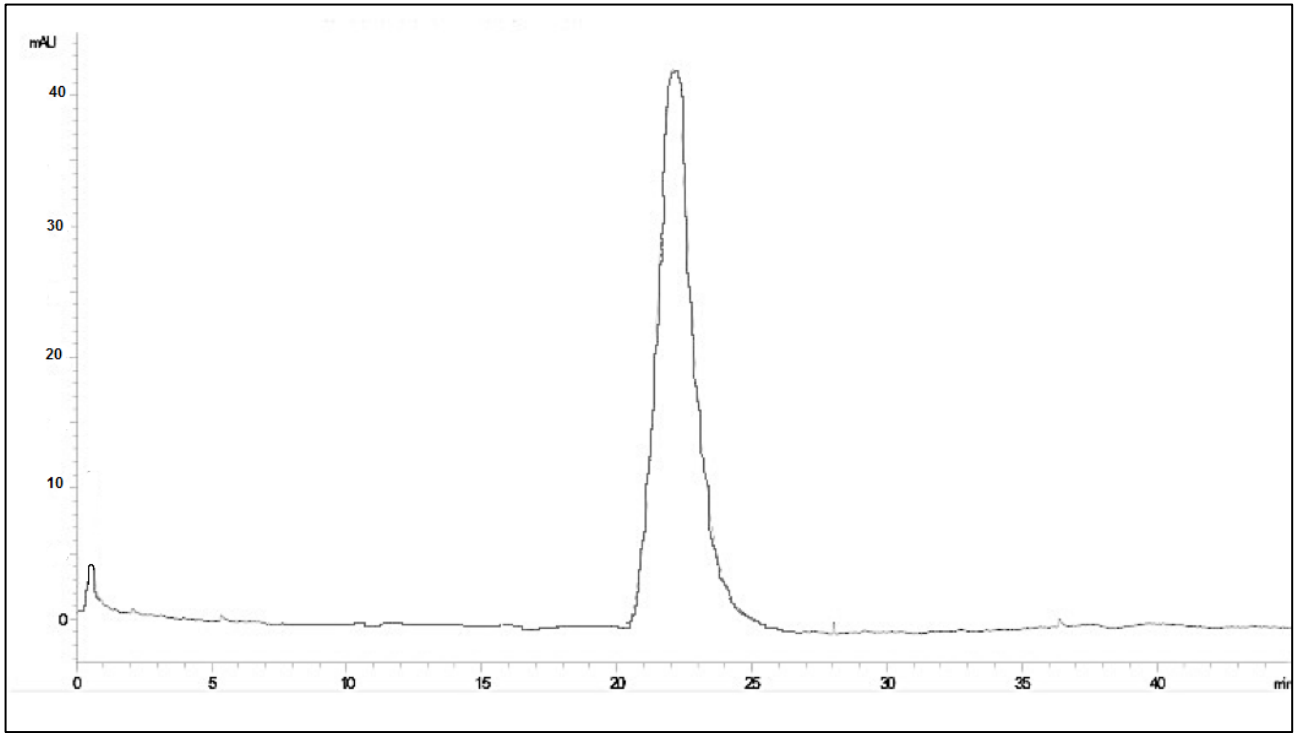
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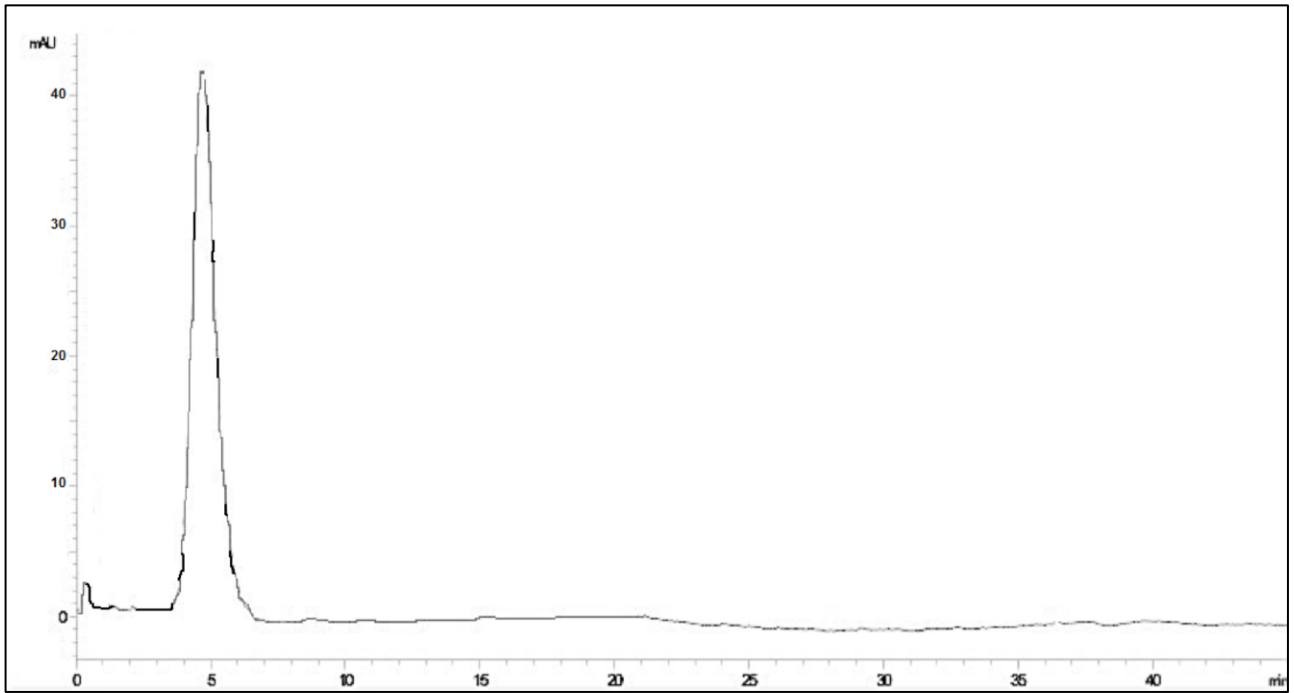
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142 **Figure 2.** IAM chromatograms achieved for the analytes A) Paroxetine (50 $\mu\text{g mL}^{-1}$ in methanol)
143 and B) Diclofenac (50 $\mu\text{g mL}^{-1}$ in methanol). The mobile phase was a solution 70/30 v/v
144 Dulbecco's phosphate-buffered saline (DPBS) / methanol. The detailed experimental conditions are
145 reported in paragraph 4.0.

146 A



147
148
149 B



150

151 **Table 1.** pKa values, $\log k_w^{\text{SDS}}$, $\log k_{30\% \text{MeOH}}^{\text{IAM}}$ indexes, log BB values, experimental UV
152 wavelengths and suppliers for the analytes taken into account.

153

Analyte	pKa	$\log k_w^{\text{SDS}}$	$\log k_{30\% \text{MeOH}}^{\text{IAM}}$	log BB	UV wavelength (nm)	Supplier
2-(Methylamino)pyridine	-	1.611	-0.164	-0.30 ¹⁸	254	Sigma-Aldrich
2,2,2-trifluoroethyl vinyl ether	-	0.929	-0.142	0.13 ¹⁸	210	Sigma-Aldrich
2,6-diisopropylphenol	-	1.688	1.097	0.91 ¹⁹	210	TCI
Acetaminophen	9.69	-0.092	-0.204	-1.00 ¹⁹	254	Europe Acros
Acetylsalicylic acid	3.50	-0.301	-0.274	-1.30 ¹⁹	230	Organics Acros
Aminopyrine	5.03	1.486	-0.206	0.00 ¹⁹	254	Organics Sigma-Aldrich
Amitriptyline	9.17	2.230	1.606	1.30 ¹⁹	254	TCI
Amobarbital	7.48/11.15*	1.208	0.059	0.04 ¹⁹	210	Europe Sigma-Aldrich
Antipyrine	1.44	1.059	-0.277	-0.10 ¹⁹	240	Acros Organics
Atenolol	9.19	1.156	-0.162	-1.00 ¹⁹	270	Acros Organics
Benzene	-	1.202	0.036	0.37 ²⁰	210	TCI
Betahistine	7.84	0.125	-0.193	-0.30 ²¹	254	Europe Acros
Caffeine	0.60	0.910	-0.284	-0.06 ¹⁹	210	Organics Acros
Carbamazepine	-	1.191	0.210	0.00 ¹⁹	210	Organics Acros
Celecoxib	9.38	1.461	1.613	-1.00 ²¹	254	Organics Acros
Chlorambucil	4.60	0.787	0.308	-1.70 ¹⁹	254	Organics TCI
Chlorpromazine	9.50	2.169	2.038	1.36 ²¹	254	Europe TCI
Cimetidine	7.01	1.003	-0.177	-1.42 ¹⁹	210	Europe TCI
Citalopram	9.22	1.832	1.005	0.48 ²¹	210	Europe TCI
Clonidine	8.08	1.436	0.171	0.11 ¹⁹	270	Europe TCI
Clozapine	7.90	1.784	1.529	0.60 ²¹	254	Europe TCI
Cotinine	-	1.424	-0.260	-0.32 ¹⁹	260	Europe TCI
Cyclobenzaprine	8.47	2.092	1.607	1.08 ²¹	230	Europe TCI
Desipramine	10.28	2.144	1.536	1.20 ¹⁹	254	Europe Sigma-Aldrich
Diclofenac	3.99	0.602	0.024	-1.70 ²¹	254	Organics Acros
Diphenhydramine	8.86	2.077	0.858	1.20 ²¹	210	Organics Acros
Domperidone	9.68	1.937	1.562	-0.78 ¹⁹	270	Organics TCI
						Europe

Donepezil	8.54*	1.968	0.858	0.89 ²²	210	Acros Organics
Eserine	8.17	1.656	0.030	0.08 ¹⁹	240	TCI Europe
Ethosuximide	9.27	0.545	-0.228	0.04 ²¹	210	Acros Organics
Ethylbenzene	-	1.588	0.600	0.26 ²⁰	210	Acros Organics
Fluphenazine	7.84/2.08*	2.207	2.066	1.51 ¹⁹	254	Sigma- Aldrich
Haloperidol	8.29	2.366	1.483	1.34 ²²	254	TCI Europe
Halothane	-	1.215	0.152	0.35 ²⁰	210	Sigma- Aldrich
Hexobarbital	8.20	1.284	-0.008	0.10 ¹⁹	254	Sigma- Aldrich
Hydroxyzine	7.52/1.58*	2.038	1.337	0.90 ²¹	210	Sigma- Aldrich
Ibuprofen	4.24	0.626	0.090	-0.18 ¹⁹	270	Acros Organics
Imipramine	9.52	2.190	1.452	1.30 ¹⁹	240	Acros Organics
Indomethacin	4.13	0.647	-0.257	-1.26 ¹⁹	210	TCI Europe
Ketorolac	3.84	-0.097	-0.500	-2.00 ²¹	300	TCI Europe
Lamotrigine	5.36	1.316	-0.006	0.48 ²³	220	Acros Organics
Levofloxacin	6.20/5.45*	1.388	-0.099	-0.70 ²¹	290	TCI Europe
Metanol	-	0.000	-0.447	0.02 ²³	210	Sigma- Aldrich
Metoclopramide	9.71	1.610	0.346	0.08 ²¹	210	TCI Europe
Metoprolol	9.56	1.771	0.198	1.15 ²²	220	TCI Europe
Mianserin	6.92	2.152	1.456	0.99 ¹⁹	280	TCI Europe
Naproxen	4.14	0.153	-0.090	-1.70 ²¹	254	Acros Organics
Nicotine	8.11	1.969	-0.139	0.40 ²⁰	210	Acros Organics
Nitrofurantoin	7.05	-0.074	-0.447	-2.00 ²¹	254	Acros Organics
Norfloxacin	8.68/5.77*	1.332	-0.062	-1.00 ²¹	280	Acros Organics
Nortriptyline	10.13	2.169	1.639	1.04 ²¹	254	TCI Europe
Olanzapine	7.80	1.825	0.843	0.80 ²¹	254	TCI Europe
Omeprazole	9.29/4.77*	1.591	-0.229	-0.82 ¹⁹	300	TCI Europe
Oxazepam	-	1.420	0.707	0.61 ¹⁹	230	Sigma- Aldrich
Paroxetine	9.77	2.104	1.796	0.48 ²¹	210	TCI Europe
Pentobarbital	8.18	1.243	0.103	0.12 ¹⁹	210	Sigma- Aldrich
Phenylbutazone	4.34	0.996	0.273	-0.52 ¹⁹	240	Acros Organics
Phenytoin	8.28	1.311	0.382	-0.04 ¹⁹	210	Acros Organics

Pindolol	9.54	0.811	0.312	0.30 ²¹	210	Sigma-Aldrich
Primidone	-	0.710	-0.152	-0.07 ²¹	210	TCI
Promazine	9.36	2.030	1.643	1.23 ²⁰	254	Europe Sigma-Aldrich
Promethazine	9.00	2.040	1.613	1.30 ²⁴	254	TCI
Propranolol	9.16	2.028	0.992	0.85 ²¹	290	Europe Acros
Quinidine	8.56	2.245	0.982	0.33 ²²	230	Organics Acros
Ranitidine	8.33	1.233	-0.239	-1.23 ¹⁹	230	Organics TCI
Rifampicin	1.70	1.900	0.990	-1.52 ²¹	230	Europe TCI
Ropinirole	10.17	1.685	0.326	0.25 ¹⁹	254	Europe Sigma-Aldrich
Salicylic acid	2.82	-0.280	-0.302	-1.10 ¹⁹	300	Acros
Theobromine	-	0.347	-0.284	-0.28 ¹⁹	270	Organics Acros
Theophylline	-	0.447	-0.218	-0.29 ¹⁹	270	Organics Acros
Toluene	-	1.459	0.330	0.37 ²⁰	210	Organics Acros
Tramadol	9.41	1.692	0.256	0.70 ²¹	210	Organics Sigma-Aldrich
Trazodone	7.30	2.223	0.780	0.30 ²¹	210	Sigma-Aldrich
Tripolidine	8.64	2.493	0.789	0.78 ²¹	230	Sigma-Aldrich
Valproic acid	4.54	0.001	-0.279	-0.84 ¹⁹	210	Acros
Venlafaxine	9.67	1.900	0.429	0.48 ²⁵	230	Organics Acros
Verapamil	8.68	2.271	1.169	-0.52 ¹⁹	210	Organics Acros
Zidovudine	9.40	0.271	-0.264	-1.00 ²⁰	270	Organics Acros
Zolmitriptan	9.55	0.974	-0.159	-1.40 ²¹	220	Organics TCI
						Europe

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155 * calculated by Marvin Sketch 15.1 software

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158 **Table 2A.** Minimum and most populated values, expressed in kcal mol⁻¹, of the cluster affinities of
 159 the analytes for the first four (from 1 to 4) discrete binding sites located on the P-gp.

160
 161

Analyte	P-gp 1 Min	P-gp 1 MP	P-gp 2 Min	P-gp 2 MP	P-gp 3 Min	P-gp 3 MP	P-gp 4 Min	P-gp 4 MP
2-(Methylamino)pyridine	-3.03	-3.03	-3.61	-3.61	-3.62	-3.62	-3.63	-3.63
2,2,2-trifluoroethyl vinyl ether	-1.72	-1.72	-2.09	-2.09	-1.85	-1.85	-1.98	-1.94
2,6-diisopropylphenol	-4.42	-4.42	-5.36	-5.36	-5.55	-5.55	-5.36	-5.36
Acetaminophen	-3.30	-3.30	-4.72	-4.72	-4.13	-4.08	-4.85	-4.85
Acetylsalicylic acid	-3.57	-3.57	-4.47	-4.36	-4.48	-4.48	-4.22	-3.95
Aminopyrine	-4.43	-4.30	-5.66	-5.66	-5.63	-5.63	-5.70	-5.70
Amitriptyline	-6.09	-5.02	-7.29	-7.15	-6.39	-6.39	-7.22	-7.22
Amobarbital	-4.14	-4.06	-5.30	-5.00	-4.83	-4.83	-5.05	-5.05
Antipyrine	-4.33	-4.33	-5.61	-5.61	-5.31	-5.31	-5.61	-5.44
Atenolol	-3.81	-3.34	-5.74	-3.99	-3.82	-3.40	-4.83	-4.70
Benzene	-2.72	-2.72	-3.31	-3.31	-3.31	-3.31	-3.31	-3.31
Betahistine	-3.06	-3.06	-3.68	-3.68	-3.32	-3.14	-3.70	-3.70
Caffeine	-3.80	-3.80	-4.60	-4.60	-4.23	-4.23	-4.33	-4.33
Carbamazepine	-5.84	-5.82	-7.09	-7.09	-6.12	-6.12	-7.07	-7.07
Celecoxib	-4.30	-4.30	-7.01	-6.98	-4.73	-4.12	-7.18	-7.18
Chlorambucil	-3.77	-3.77	-5.13	-4.81	-5.19	-5.19	-5.16	-5.16
Chlorpromazine	-5.38	-4.84	-7.24	-7.24	-6.57	-6.57	-7.26	-7.26
Cimetidine	-3.27	-2.95	-4.74	-4.60	-4.10	-4.02	-4.70	-4.64
Citalopram	-4.75	-4.45	-6.41	-6.14	-4.93	-4.93	-7.16	-6.86
Clonidine	-4.34	-4.34	-5.41	-5.41	-5.44	-5.44	-5.40	-5.40
Clozapine	-5.10	-5.05	-7.01	-7.01	-5.36	-5.36	-7.00	-6.98
Cotinine	-3.93	-3.87	-5.14	-4.79	-4.92	-4.92	-4.82	-4.82
Cyclobenzaprine	-6.32	-5.14	-7.18	-7.04	-6.94	-6.94	-7.14	-7.14
Desipramine	-5.75	-5.43	-6.80	-6.62	-6.26	-6.08	-6.53	-6.53
Diclofenac	-5.23	-4.96	-6.34	-6.13	-6.49	-6.19	-6.14	-6.05
Diphenhydramine	-4.35	-3.91	-5.63	-5.63	-5.14	-5.14	-5.61	-5.61
Domperidone	-5.23	-4.82	-7.12	-6.98	-6.39	-6.39	-7.18	-7.18
Donepezil	-6.05	-6.05	-7.70	-7.46	-6.69	-6.67	-7.72	-7.65
Eserine	-4.88	-4.88	-6.01	-5.87	-5.45	-5.45	-5.91	-5.91
Ethosuximide	-3.62	-3.62	-4.22	-4.22	-4.47	-4.39	-4.42	-4.42
Ethylbenzene	-3.34	-3.34	-4.22	-4.22	-4.07	-4.07	-4.07	-4.07
Fluphenazine	-4.81	-3.58	-6.75	-6.75	-4.30	-4.12	-6.60	-5.82
Haloperidol	-4.35	-4.17	-6.25	-6.25	-5.60	-5.60	-7.23	-6.20
Halothane	-2.12	-2.12	-2.76	-2.75	-2.74	-2.72	-2.66	-2.64
Hexobarbital	-4.85	-4.79	-6.02	-6.02	-5.13	-4.97	-6.03	-6.03
Hydroxyzine	-4.24	-3.67	-6.41	-5.43	-4.05	-3.66	-6.19	-5.84
Ibuprofen	-4.37	-4.37	-5.52	-5.43	-5.88	-5.88	-5.43	-5.34
Imipramine	-5.34	-5.34	-6.68	-6.68	-5.76	-4.64	-6.68	-6.13
Indomethacin	-5.67	-5.53	-7.02	-7.02	-7.28	-7.28	-7.37	-7.37
Ketorolac	-5.22	-5.22	-6.61	-6.61	-6.55	-6.41	-6.62	-6.62
Lamotrigine	-4.49	-3.92	-5.84	-5.84	-4.88	-4.56	-5.36	-5.36
Levofloxacin	-4.45	-4.45	-5.80	-5.54	-5.80	-5.23	-6.07	-5.85
Metanol	-1.40	-1.33	-1.43	-1.43	-1.33	-1.33	-1.40	-1.40

Metoclopramide	-3.47	-3.47	-5.19	-3.92	-3.71	-3.07	-4.52	-4.13
Metoprolol	-3.58	-3.35	-4.63	-4.06	-3.54	-2.87	-4.39	-4.18
Mianserin	-5.23	-5.23	-7.06	-7.06	-6.05	-5.98	-7.11	-7.11
Naproxen	-4.82	-4.82	-5.99	-5.99	-6.03	-5.91	-6.03	-6.03
Nicotine	-4.02	-4.02	-4.70	-4.69	-4.50	-4.50	-4.70	-4.70
Nitrofurantoin	-4.10	-4.10	-5.33	-5.33	-5.18	-5.18	-5.32	-5.32
Norfloxacin	-3.83	-3.83	-5.59	-5.59	-5.51	-5.51	-5.75	-5.63
Nortriptyline	-6.35	-6.35	-7.07	-6.86	-6.44	-6.44	-7.00	-7.00
Olanzapine	-4.60	-4.60	-6.71	-6.62	-5.47	-5.29	-6.68	-6.68
Omeprazole	-5.26	-5.20	-6.96	-6.76	-6.65	-6.41	-7.16	-6.92
Oxazepam	-5.29	-5.29	-6.90	-6.90	-6.16	-6.16	-6.89	-6.89
Paroxetine	-4.94	-4.94	-6.95	-6.84	-6.42	-5.67	-6.83	-6.29
Pentobarbital	-3.88	-3.88	-4.76	-4.76	-4.41	-4.41	-4.91	-4.91
Phenylbutazone	-5.53	-5.53	-7.27	-7.27	-6.02	-5.12	-7.45	-6.69
Phenytoin	-5.00	-5.00	-6.56	-6.56	-5.39	-5.00	-6.55	-6.55
Pindolol	-4.17	-4.13	-5.43	-4.86	-4.56	-4.26	-5.47	-4.60
Primidone	-4.55	-4.55	-5.19	-5.19	-4.88	-4.71	-5.50	-5.50
Promazine	-5.58	-5.58	-6.79	-6.50	-6.32	-6.32	-6.50	-5.84
Promethazine	-4.99	-4.80	-6.78	-6.49	-6.60	-5.83	-6.51	-6.42
Propranolol	-4.60	-4.38	-6.33	-5.54	-4.79	-4.22	-6.10	-5.39
Quinidine	-5.72	-5.72	-7.43	-7.43	-5.26	-4.56	-7.77	-7.77
Ranitidine	-2.77	-2.65	-4.11	-3.91	-4.42	-2.88	-4.31	-3.48
Rifampicin	-7.10	-6.50	-4.48	-4.32	-6.80	-5.95	-7.22	-6.59
Ropinirole	-4.02	-4.01	-6.30	-5.55	-4.72	-3.94	-5.94	-5.52
Salicylic acid	-3.09	-3.09	-3.69	-3.69	-4.00	-4.00	-3.74	-3.71
Theobromine	-3.47	-3.47	-4.54	-4.54	-4.08	-4.08	-4.54	-4.54
Theophylline	-3.63	-3.63	-4.43	-4.43	-3.88	-3.87	-4.43	-4.43
Toluene	-3.08	-3.08	-3.81	-3.81	-3.77	-3.77	-3.74	-3.74
Tramadol	-4.89	-4.89	-5.94	-5.20	-5.15	-3.77	-5.71	-5.30
Trazodone	-5.37	-5.00	-7.30	-5.74	-5.79	-6.47	-7.09	-6.06
Triprolidine	-5.03	-4.92	-7.19	-7.11	-5.36	-5.36	-6.96	-6.88
Valproic acid	-2.78	-2.78	-3.56	-3.56	-3.81	-3.81	-3.43	-3.43
Venlafaxine	-4.81	-4.54	-6.07	-5.82	-4.54	-4.09	-6.47	-6.08
Verapamil	-3.93	-3.54	-6.56	-6.37	-5.28	-4.48	-6.58	-6.14
Zidovudine	-3.45	-3.21	-5.14	-5.14	-3.56	-3.27	-5.20	-5.20
Zolmitriptan	-4.48	-4.32	-6.32	-6.01	-5.56	-5.56	-6.38	-5.73

163 **Table 2B.** Minimum and most populated values, expressed as kcal mol⁻¹, of the cluster affinities of
164 the analytes for the second four (from 5 to 8) discrete binding sites located on the P-gp.

165

Analyte	P-gp 5 Min	P-gp 5 MP	P-gp 6 Min	P-gp 6 MP	P-gp 7 Min	P-gp 7 MP	P-gp 8 Min	P-gp 8 MP
2-(Methylamino)pyridine	-3.03	-3.03	-3.40	-3.40	-3.53	-3.53	-3.22	-3.22
2,2,2-trifluoroethyl vinyl ether	-1.72	-1.64	-2.05	-2.05	-2.18	-2.18	-1.93	-1.93
2,6-diisopropylphenol	-4.42	-4.22	-5.17	-5.17	-5.56	-5.56	-4.67	-4.67
Acetaminophen	-3.81	-3.81	-4.20	-4.20	-4.10	-4.10	-3.37	-3.37
Acetylsalicylic acid	-3.86	-3.86	-3.91	-3.91	-4.42	-4.42	-3.68	-3.68
Aminopyrine	-5.20	-5.18	-5.36	-4.91	-5.60	-5.60	-4.47	-4.47
Amitriptyline	-6.00	-5.92	-6.85	-6.85	-7.49	-7.49	-5.63	-5.63
Amobarbital	-4.51	-4.51	-4.40	-4.05	-4.99	-4.99	-4.23	-4.23
Antipyrine	-4.78	-4.78	-4.77	-4.77	-5.58	-5.58	-4.31	-4.30
Atenolol	-4.03	-4.03	-4.82	-4.60	-4.86	-4.86	-3.37	-3.37
Benzene	-3.19	-3.19	-3.26	-3.26	-3.31	-3.31	-3.01	-3.01
Betahistine	-2.93	-2.93	-3.54	-3.54	-3.52	-3.52	-3.41	-3.41
Caffeine	-3.64	-3.64	-3.76	-3.76	-4.56	-4.56	-3.76	-3.76
Carbamazepine	-5.87	-5.87	-6.16	-6.16	-6.83	-6.83	-5.31	-5.31
Celecoxib	-5.93	-5.31	-5.82	-5.82	-7.97	-7.97	-5.59	-5.04
Chlorambucil	-4.30	-4.30	-4.57	-4.56	-5.63	-5.63	-3.93	-3.93
Chlorpromazine	-5.66	-5.00	-6.75	-6.57	-7.23	-7.01	-5.16	-5.16
Cimetidine	-3.72	-3.45	-4.58	-3.80	-4.76	-3.88	-3.41	-3.20
Citalopram	-5.63	-5.59	-5.85	-5.69	-6.44	-6.44	-4.93	-4.86
Clonidine	-4.06	-3.98	-4.47	-4.40	-5.34	-5.34	-4.31	-4.31
Clozapine	-5.44	-5.18	-6.64	-6.64	-7.05	-7.05	-4.98	-4.95
Cotinine	-4.52	-4.52	-4.48	-4.48	-4.86	-4.76	-4.29	-4.29
Cyclobenzaprine	-5.70	-5.68	-7.12	-6.86	-7.35	-7.23	-5.36	-5.28
Desipramine	-5.74	-5.74	-6.44	-6.44	-6.42	-5.73	-4.78	-4.78
Diclofenac	-5.23	-5.23	-5.96	-5.46	-6.32	-6.32	-4.65	-4.37
Diphenhydramine	-5.07	-5.07	-5.41	-4.78	-5.42	-5.42	-3.89	-3.54
Domperidone	-6.04	-5.71	-5.73	-5.73	-7.69	-7.69	-6.26	-6.13
Donepezil	-7.11	-6.10	-7.07	-7.07	-7.83	-7.77	-6.58	-6.58
Eserine	-5.52	-5.50	-5.57	-5.57	-5.92	-5.92	-4.57	-4.54
Ethosuximide	-3.62	-3.62	-4.16	-4.16	-4.34	-4.34	-3.76	-3.35
Ethylbenzene	-3.58	-3.58	-3.96	-3.96	-4.21	-4.21	-3.71	-3.71
Fluphenazine	-4.39	-4.39	-5.23	-3.06	-6.46	-5.16	-5.79	-5.79
Haloperidol	-5.68	-5.14	-5.35	-4.41	-7.32	-7.31	-5.48	-5.35
Halothane	-2.37	-2.37	-2.57	-2.55	-2.86	-2.86	-2.41	-2.35
Hexobarbital	-5.21	-4.99	-5.22	-5.22	-5.51	-5.51	-4.56	-4.50
Hydroxyzine	-5.26	-4.29	-5.29	-5.29	-6.18	-5.89	-4.57	-4.50
Ibuprofen	-4.84	-4.84	-4.91	-4.69	-5.41	-5.41	-4.55	-4.55
Imipramine	-5.83	-5.18	-6.69	-6.32	-6.67	-6.67	-4.97	-4.97

Indomethacin	-5.92	-5.92	-6.44	-6.44	-7.22	-6.91	-5.26	-5.26
Ketorolac	-5.73	-5.68	-5.71	-5.63	-6.36	-6.35	-5.01	-4.74
Lamotrigine	-4.13	-4.09	-4.77	-4.77	-5.30	-5.23	-4.32	-3.66
Levofloxacin	-5.40	-5.14	-3.46	-3.46	-6.37	-6.37	-4.71	-4.71
Metanol	-1.47	-1.47	-1.38	-1.38	-1.37	-1.37	-1.42	-1.42
Metoclopramide	-3.63	-3.36	-4.40	-4.02	-4.95	-4.95	-3.55	-2.80
Metoprolol	-3.49	-3.46	-4.25	-4.25	-4.60	-4.39	-3.57	-3.42
Mianserin	-5.89	-5.89	-6.21	-6.21	-7.07	-7.07	-5.63	-5.63
Naproxen	-5.26	-5.26	-5.48	-5.48	-5.80	-5.80	-4.72	-4.71
Nicotine	-3.77	-3.77	-4.42	-4.20	-4.67	-4.67	-4.24	-4.13
Nitrofurantoin	-4.37	-4.30	-4.62	-4.31	-5.24	-5.24	-3.90	-3.67
Norfloxacin	-4.46	-4.44	-3.73	-2.70	-5.85	-5.83	-4.99	-4.99
Nortriptyline	-5.98	-5.98	-7.20	-7.20	-7.09	-7.07	-5.30	-5.05
Olanzapine	-5.42	-5.42	-6.24	-6.24	-6.48	-6.48	-5.06	-5.06
Omeprazole	-5.24	-5.22	-6.47	-6.47	-7.26	-6.79	-5.28	-4.22
Oxazepam	-5.96	-5.94	-6.61	-6.61	-6.81	-6.70	-5.02	-5.02
Paroxetine	-5.71	-5.03	-6.14	-4.23	-7.49	-6.50	-4.97	-4.97
Pentobarbital	-4.29	-4.18	-4.35	-4.22	-4.84	-4.84	-3.75	-3.60
Phenylbutazone	-6.29	-6.19	-7.19	-7.19	-7.33	-6.78	-5.57	-5.44
Phenytoin	-5.80	-5.80	-5.42	-5.38	-6.25	-6.25	-4.67	-4.64
Pindolol	-3.61	-3.58	-5.72	-5.72	-5.29	-5.13	-4.03	-3.81
Primidone	-4.43	-4.31	-4.99	-4.99	-5.23	-5.23	-4.26	-4.08
Promazine	-5.47	-4.86	-6.22	-5.94	-6.47	-6.05	-4.71	-4.50
Promethazine	-5.55	-4.99	-6.07	-5.87	-6.62	-6.40	-4.72	-4.68
Propranolol	-4.42	-4.42	-5.99	-5.19	-5.67	-5.62	-4.30	-4.30
Quinidine	-5.68	-5.68	-6.74	-5.62	-7.92	-7.78	-5.32	-4.72
Ranitidine	-2.99	-2.99	-3.61	-3.34	-3.76	-2.86	-2.12	-1.63
Rifampicin	-5.13	-3.28	-6.66	-6.00	-7.44	-7.00	-3.14	-3.14
Ropinirole	-4.25	-4.17	-5.98	-5.98	-6.22	-6.22	-4.11	-3.39
Salicylic acid	-3.09	-3.09	-3.42	-3.42	-3.85	-3.85	-3.31	-3.31
Theobromine	-3.46	-3.46	-3.49	-3.49	-4.29	-4.29	-3.80	-3.80
Theophylline	-3.63	-3.63	-3.72	-3.72	-4.45	-4.45	-3.66	-3.66
Toluene	-3.42	-3.42	-3.79	-3.79	-3.93	-3.93	-3.39	-3.39
Tramadol	-4.46	-4.46	-5.36	-5.36	-5.62	-5.46	-4.26	-4.26
Trazodone	-6.06	-6.26	-6.86	-6.86	-7.40	-7.29	-6.06	-5.47
Triprolidine	-6.48	-6.48	-6.40	-5.53	-7.28	-7.28	-5.38	-5.26
Valproic acid	-3.03	-3.03	-3.10	-3.10	-3.76	-3.76	-2.90	-2.90
Venlafaxine	-4.64	-4.33	-5.60	-5.23	-6.48	-6.48	-4.40	-4.40
Verapamil	-5.16	-4.84	-4.74	-3.51	-6.80	-6.53	-4.32	-3.72
Zidovudine	-3.94	-3.93	-4.49	-4.49	-4.55	-4.55	-3.90	-3.19
Zolmitriptan	-4.78	-3.70	-6.13	-6.13	-6.15	-6.15	-4.42	-4.42

167 2.1 MLC INDEXES IN LOG BB PREDICTION

168 MLC indexes were used to develop BBB passage potential predicting models along with either
169 static or conformational properties. At first, all the analytes were assumed as having zero atomic
170 charges, even the ones supporting one or more ionizable functions. The equations along with the
171 statistical validation are reported in Table 3. In the equations thereby reported, r^2 is the multiple
172 regression coefficient, q^2 is the r^2 validated by Leave-One-Out (LOO) optimization, SE is the error
173 standard deviation, F represents the Fischer regression statistical value, PC is the Amemiya
174 predictive criterion and ExRow is the analyte excluded for maximizing the predictive strength of
175 the statistical model. If not differently indicated, every regression was developed by employing four
176 different independent variables (MLC indexes + three other physico-chemical descriptors).
177 Surprisingly, even if over two thirds of the analytes support one or more ionizable functions, fairly
178 good relationships, as the one expressed by equations (1) and (2), are obtained even not taking into
179 account the presence of electric charges. This may be attributed to the fact that, although the
180 molecular mechanisms involved in MLC are multiple and complex, the occurrence of
181 analyte/micelles electrostatic interactions plays a pivotal role in the global retention and it appears
182 reasonable to assume that such interactions are encoded in MLC indexes. It should be also
183 highlighted that, in these specific cases, being VirtualLogP values calculated starting from the
184 analytes assumed in their forms having zero atomic charges, such values can be reasonably assumed
185 as estimates of their $\log P^N$ values. Subsequently, the analytes supporting extensively ionizable
186 functions (i.e. carboxy groups, for acids primary, secondary and tertiary amines for bases) were
187 assumed as completely charged, regardless of the relative abundance of the charged species at the
188 physiological pH. Considering the ionizable analytes as entirely charged species slightly worsened
189 the relationships (equations (3) and (4)). It should be pointed out that verapamil, the analyte
190 excluded to maximize the predictive strength of the statistical model is a well-known P-gp
191 substrate²⁶. P-gp is an ATP-dependent efflux pump, with broad substrate specificity which pumps
192 many foreign substances out of cells²⁷. Although it is widely expressed in the intestinal epithelium,

193 liver cells and proximal tubule of the kidney, P-gp is also localized in the capillary endothelial cells
194 composing the BBB and is responsible, for some classes of actives, of multi-drug resistance.
195 Eventually, a weighted average of the static properties at physiological pH (7.4), according to the
196 pKa of each compound, was performed. For zwitterions, the static properties were calculated for
197 each microspecies possibly present at pH 7.4 and their relative abundances, calculated by the
198 software Marvin Sketch 15.1 for Mac OS X²⁸, were also used to perform the weighted averages.
199 The relative abundances of the microspecies present at pH 7.4 are reported in the Supporting
200 Information section for the ampholytes levofloxacin (page S-2), norfloxacin (page S-4) and
201 omeprazole (page S-6). This approach was adapted to mirror more closely what actually occurs *in*
202 *vivo*. Performing the weighted average of the properties benefited noticeably the relationships as
203 described by equations (5) and (6). It is also interesting to note how, according to the above
204 reported relationships, the BBB penetration of drugs will be enhanced for highly retained
205 compounds in MLC, how it is hindered by the occurrence of drug/membrane polar (Psa)/
206 electrostatic (Dipole) interactions, and how the transport through the barrier seems favored for
207 bases (Charge). However, by taking into account the analytes assumed as static, the properties are
208 derived considering them in their minimum energy conformations, i.e. after minimization. Indeed,
209 several authors²⁹ reported that such conformations are not always the ones actually involved in
210 membrane barrier passage. Therefore, a conformational analysis *in vacuum* was carried out for each
211 analyte included in the data set by using the *Boltzmann Jump* method that generates at random 1000
212 possible conformations by exploring the conformational space of the rotatable dihedral angles. The
213 conformational analysis was first performed on the analytes assumed as having zero atomic
214 charges, then on the analytes assumed as completely charged and finally taking into account a
215 weighted average of the properties at the experimental pH 7.4, according to the pKa of each analyte.
216 In the following models the conformational properties were considered separately to look into the
217 predictive strength of the models. As it is evident from Table 3, the use of conformational
218 properties instead of the static ones slightly worsened the relationships. This aspect is quite

219 interesting as the calculation of conformational properties can be rather time-consuming especially
220 if the compound libraries to screen are wide and the computers employed are not sufficiently
221 powerful. Conversely, the static properties are much faster to calculate. Performing the weighted
222 average of the conformational properties yielded the most predictive models (equations (11) and
223 (12)) and in those relationships, verapamil again behaved as an *outlier*, suggesting that such models
224 would not be able to mirror the penetration of analytes undergoing some sort of active transport, in
225 this case P-gp mediated efflux. It is interesting to point out how, among the ionized properties
226 employed for the statistical method development (equations (9) and (10)), no one depends
227 noticeably on ionization. Furthermore, the conformational analysis demonstrated how it is the PSA
228 Max, i.e. the maximum value that the PSA assumes by exploring the conformational space *in*
229 *vacuum* of each analyte, that best relates with log BB values as those values are incorporated in
230 each model based on conformational properties (equations (7), (8), (9), (10), (11) and (12)).

231

232 **Table 3.** Statistical validation of the models developed employing $\log k_w^{\text{SDS}}$ values of the dataset
233 $(n=79)$ along with three other physico-chemical descriptors.

234

MOLECULAR DESCRIPTORS	r^2	q^2	SE	F	PC	r^2 (n-1)	SE (n-1)	F (n-1)	PC (n-1)	EX-ROW	EQUATIONS	EQ No
STATIC												
ZERO ATOMIC CHARGES PROPERTIES	0.69	0.65	0.521	41.42	21.656	0.71	0.510	44.06	20.535	-	log BB = - 0.2693 + 0.8191 log k_w^{SDS} - 0.0162 Ps _a - 0.0824 VirtualLogP + 0.1456 HbDon	1
										2- (Methylamino) pyridine	log BB = - 0.2166 + 0.8383 log k_w^{SDS} - 0.0170 Ps _a - 0.0994 VirtualLogP + 0.1570 HbDon	2
IONIZED PROPERTIES	0.68	0.63	0.530	39.28	22.452	0.70	0.517	42.20	21.104	-	log BB = - 0.3460 + 0.6671 log k_w^{SDS} - 0.0104 Ps _a + 0.1425 Charge - 0.0138 Impropers	3
										Verapamil	log BB = - 0.4024 + 0.7071 log k_w^{SDS} - 0.0093 Ps _a + 0.1297 Charge - 0.0187 Impropers	4
WEIGHTED AVERAGE	0.72	0.68	0.498	47.05	19.795	0.73	0.486	50.29	18.617	-	log BB = - 0.3145 + 0.6825 log k_w^{SDS} - 0.0091 Ps _a - 0.0202 Dipole + 0.2042 Charge	5
										Verapamil	log BB = - 0.3145 + 0.6825 log k_w^{SDS} - 0.0091 Ps _a -	6

											0.0202 Dipole + 0.2042 Charge	
CONFORMATIONAL												
ZERO ATOMIC CHARGES PROPERTIES	0.69	0.64	0.526	40.25	22.081	0.71	0.509	44.37	20.426	-	log BB = - 0.4857 + 0.8201 log k_w^{SDS} - 0.0044 PSA Max - 0.0708 MD Max - 0.2671 MD sd	7
										Pindolol	log BB = - 0.4973 + 0.8307 log k_w^{SDS} - 0.0048 PSA Max - 0.0583 MD Max - 0.3274 MD sd	8
IONIZED PROPERTIES	0.65	0.62	0.549	69.77	23.504	0.67	0.538	74.60	22.304	-	log BB = - 0.5392 + 0.7898 log k_w^{SDS} - 0.0093 PSA Max	9
										Primidone	log BB = - 0.5338 + 0.8008 log k_w^{SDS} - 0.0099 PSA Max	10
WEIGHTED AVERAGE	0.68	0.63	0.534	38.57	22.729	0.70	0.520	41.69	21.283	-	log BB = - 0.4329 + 0.7969 log k_w^{SDS} - 0.0072 PSA Max - 0.0235 MD Min - 0.0485 MLP Average	11
										Verapamil	log BB = - 0.4911 + 0.8121 log k_w^{SDS} - 0.0068 PSA Max - 0.0233 MD Min - 0.0334 MLP Average	12

2.2 IAM INDEXES IN LOG BB PREDICTION

The same approach was extended to the IAM indexes. The equations along with the statistical validation coefficient are reported in Table 4. Indeed, taking into account either the properties of the analytes assumed as having zero atomic charges (equations (13) and (14)) or those of the analytes

assumed as completely charged (equations (15) and (16)) resulted in a BBB passage predictive strength inferior to that obtained by using MLC indexes. Such conclusions are supported by the lower correlation coefficients obtained. It is interesting to note how domperidone, the compound excluded in first best optimized model described by equation (14), is a well-known substrate of the P-gp²⁶, and is pumped out of cells by such efflux system despite its high biomembrane passive diffusion. Analogously to what was observed in the analysis of MLC indexes, performing the weighted average of the static properties resulted the winning strategy also for this set of experimental measures. In fact, a 0.72 r^2 ($n-1$), achieved on a set as large as 79 analytes, employing only four descriptors suggests that the model (equations (17) and (18)) is robust and reliable. However, these relationships are roughly comparable to those obtained by using MLC indexes (equations (5) and (6)). This evidence is indeed rather surprising, since the IAM stationary phase consists of analogues of phosphatidylcholine, the most abundant phospholipid expressed in the capillary endothelium acting as a barrier between the blood and the cerebrospinal fluids (CSF), and thus they would represent an ideal biomimetic system. Conversely, this kind of SDS based MLC should have drawbacks arising from the different chemical structure of SDS in comparison with membrane phospholipids. But this evidence would suggest that they are incidentally able to mirror the drug/membrane interactions involved *in vivo* thanks to the peculiar amphiphilic features of the anionic micelles. In fact, for some reasons, they seem to be able to model the passive BBB penetration of drugs fairly better than IAM indexes. Furthermore, the physico-chemical descriptors reported in equation (18) are the same as the ones in equation (6), supporting again the concept according to which the polar (Psa) /electrostatic (Dipole) interaction component plays a relevant role in hindering the BBB penetration of drugs. Again, bases seem to be favored in BBB entering and this is also consistent with the clinical experience. In fact, polar and extensively protonated at pH 7.4 basic compounds, such as amphetamine and methamphetamine, are known to have an appreciable CNS activity but it is much harder to recall similar cases for polar acids. The conformational analyses of the analytes neither assumed as having zero atomic charges, nor as

265 ionized benefitted the relationships. Moreover, even performing the weighted average was not
266 beneficial at all for the relationships (data not shown).

267

268 **Table 4.** Statistical validation of the models developed employing $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ values of the
269 dataset ($n=79$) along with three other physico-chemical descriptors.

270

MOLECULAR DESCRIPTORS	r^2	q^2	SE	F	PC	r^2 (n-1)	SE (n-1)	F (n-1)	PC (n-1)	EX-ROW	EQUATIONS	EQ No
STATIC												
ZERO ATOMIC CHARGES PROPERTIES	0.64	0.59	0.561	33.08	25.156	0.67	0.540	36.88	23.025	-	log BB = +0.6691 + 0.8369 $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ - 0.0166 P_{sa} - 0.1473 VirtualLogP + 0.1139 HbDon	13
										Domperidone	log BB = +0.6706 + 0.9057 $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ - 0.0161 P_{sa} - 0.1473 VirtualLogP + 0.1173 HbDon	14
IONIZED PROPERTIES	0.64	0.58	0.561	33.07	25.155	0.67	0.540	37.29	22.976	-	log BB = -0.3460 + 0.5276 $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ + 0.0680 HbAcc - 0.0164 P_{sa} - 0.3020 Charge	15
										Lamotrigine	log BB = 0.3429 + 0.5324 $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ + 0.1027 HbAcc + 0.3288 Charge - 0.0188 P_{sa}	16
WEIGHTED AVERAGE	0.70	0.65	0.515	42.79	21.169	0.72	0.494	47.40	19.219	-	log BB = +0.4388 + 0.5458 $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ - 0.0110 P_{sa} - 0.0190 Dipole + 0.4653 Charge	17
										Celecoxib	log BB = +0.3773 + 0.6063 $\log k_{30\% \text{ MeOH}}^{\text{IAM}}$ - 0.0097 P_{sa} - 0.0207 Dipole + 0.4182 Charge	18

271

272 2.3 IAM + MLC INDEXES IN LOG BB PREDICTION

273 In the present study, MLC and IAM indexes were, in a first instance, considered separately.
274 However, the evident differences in the elution order observed depict a rather different selectivity
275 between both techniques. For this reason, the development of the BBB entering potential statistical
276 models was also performed by considering both the chromatographic indexes at the same time,
277 along with three other molecular descriptors (five independent variables in total), starting from the
278 weighted average of the molecular properties. This strategy resulted in a markedly improved
279 predictive strength (equations (19) and (20)) as reported in Table 5. These relationships may
280 suggest that the molecular mechanism involved in IAM-LC and MLC are different but play both a
281 relevant role in BBB diffusion of drugs.

282 2.4 P-GP AFFINITIES IN LOG BB PREDICTION

283 As already mentioned, each analyte present in the dataset was docked into each discrete binding site
284 on the P-gp and the binding affinities were incorporated in the development of BBB passage
285 predictive statistical models. Indeed, recent functional studies have identified seven sometimes
286 overlapping binding sites accommodating substrates and inhibitors in the greasy, polyspecific
287 binding cavity of P-gp. These binding sites were demonstrated able to allosterically communicate in
288 a negative heterotropic manner. Moreover, an additional binding site was recognized on the exterior
289 of P-gp bounded by residues from the transmembrane helices 9, 12 and the elbow helix 2. This site
290 faces away from the transporter, lying close to the predicted membrane–water interface and
291 intramembranous substrate-entry portal.

292 On average , highly clustered results were achieved, meaning that the conformational search
293 procedure was exhaustive enough to ensure a coverage of the accessible conformational space. An
294 extensive cluster analysis (RMSD tolerance was set to 2.0 Å) was performed and the binding
295 affinity now reported in Table 2A and Table 2B are the minimum and the most populated binding
296 energies of the clusters. The errors of the estimates of free energies of binding were never higher

297 than $\pm 1.8 \text{ kcal mol}^{-1}$. However, from the relationships reported above, P-gp affinities do not seem
298 to have an appreciable role in BBB passage. However, this is not entirely true because the statistical
299 model development was carried out using only four independent variables, thus leading the
300 software to select only the four most relevant descriptors, among which P-gp affinities were not
301 included. Indeed, when five independent variables were set in the statistical method development,
302 the P-gp binding affinities (Table 5A and 5B) were used by the software to build up the models.
303 Equations (21) and (22), generated by IAM indexes and four static properties of the analytes,
304 assumed as having zero atomic charges, is an example as can be seen in Table 5. The AutoDock
305 GPF/DPF files for site 1 and 7, i.e. the ones actually involved in the statistical models (21) and (22),
306 are now provided as supplementary materials.

307 This is not surprising because among the considered analytes, the only ones known from the
308 literature to be substrates of P-gp are cimetidine, domperidone, ranitidine, rifampicin, quinidine and
309 verapamil²⁶, and they represent less than 5% of the dataset. Indeed, the compounds considered were
310 selected in the attempt to mirror as accurately and completely as possible the marketed drugs, in
311 terms of diverse chemical nature, molecular volume, CNS activity and molecular lipophilicity.
312 Since the active transport comprises only for a minority of drugs, whereby the drug uptake in
313 mainly driven by passive transcellular diffusion, the limited predictivity of the P-gp molecular
314 affinity may be dataset related. This approach suffers from some limitations, the most evident one
315 being the aspect that the receptor flexibility is not taken into account. The main reason behind it is
316 the large number of degrees of freedom that should be considered in this kind of calculations, thus
317 requiring remarkable computational power. However, neglecting the receptor flexibility could lead
318 to poor docking results in terms of binding pose prediction in real-world settings. Therefore, these
319 results must be regarded as a preliminary attempt to gain new insights and model the active efflux
320 of drugs pumped out of cells by P-gp, being neither exhaustive nor complete. Other experiments
321 have to be performed and docking conditions further calibrated in order to validate the proposed

322 model.

323

2.5 E-DRAGON DESCRIPTORS IN MAXIMIZING THE PREDICTIVE STRENGTH OF THE MODELS

In an attempt to further maximize the predictive strength of the models, IAM and MLC indexes were used in combination with E-Dragon descriptors³⁰. The E-Dragon software calculates more than 1,600 descriptors, including not only the simplest atom type, functional group and fragment counts, but also several topological and geometrical descriptors. The results and statistical method validation are reported in Table 5. Remarkably high correlation coefficients were achieved with either IAM ($r^2 = 0.78$, equation (26)) or MLC ($r^2 = 0.83$, equation (24)). As suggested by the similarly high q^2 values, those relationships are not affected by any over fitting. The plots of the experimental vs predicted log BB values (as predicted by equation (24)) are reported in Figure 3. Such relationship is based on MLC indexes, TPSA (Tot) i.e. the topological polar surface area using nitrogen, oxygen, sulphur, phosphorus polar contributions which differs from the TPSA (NO), included, for instance in equations (23), (25) and (26) that instead takes into account, in the topological polar surface area computation the nitrogen and oxygen contributions only. nRCOOH and D/Dr 05 are included in all the equations reported in Table 5. While the former is a functional group descriptor referring to the number of aliphatic carboxylic acids, the latter is a topological descriptor, named distance/detour ring index of order 5. It is based on operation over the distance/detour matrix D/Δ , a square symmetric matrix that contains the ratios of the lengths of the shortest to the longest path between any pair of vertices. It is calculated by the following equation:

$$D/\Delta = \frac{1}{2} \sum_{i=1}^A \sum_{j=1}^A (D/\Delta)_{ij}$$

351

Although the role that such a parameter could play in the BBB partition is unclear, being its interpretation quite difficult, it cannot be excluded that it might affect the molecular flexibility of the analytes. However, the models obtained starting from E-Dragon descriptors would again support the view according to which the BBB penetration of drugs would be enhanced for highly

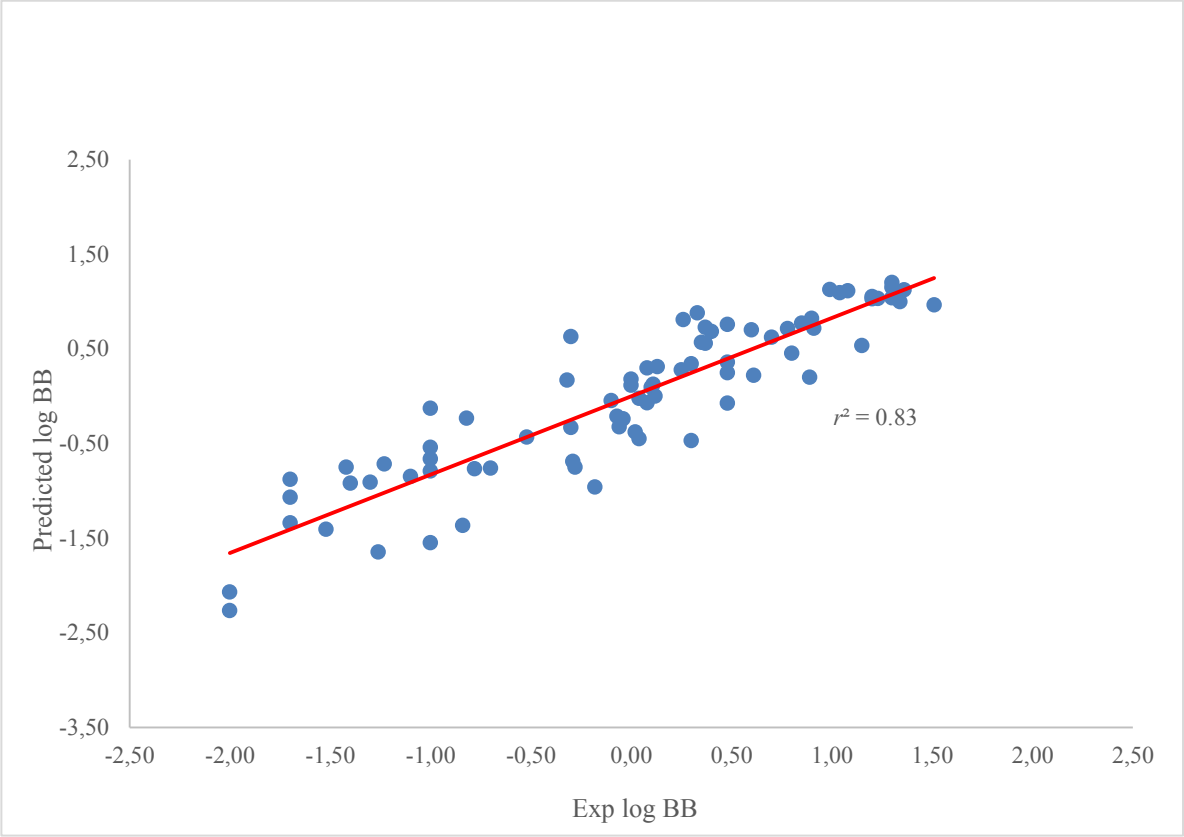
355

retained compounds either in IAM or MLC and hindered for compounds having greater PSA and supporting one or even more acidic functions. To further validate the proposed method, the datasets were divided randomly into 16 pairs of training and test sets. For each pair, the multiple linear regression was performed and the equations derived from the training sets were subsequently used to predict the log BB values of the test sets. Such value set was used to evaluate the regression coefficient (r^2), the standard error (SE) of the estimates and the Fischer coefficients. The results of this additional validation are shown in Table 6.

Table 6. Validation of the best model employing four descriptors for log BB prediction.

Model Validation					
Trial	Training set			Test set	
	r^2	SE	F	r^2	SE
1	0.87	0.320	57.459	0.75	0.496
2	0.85	0.412	46.645	0.71	0.445
3	0.84	0.411	45.880	0.71	0.498
4	0.84	0.390	45.696	0.74	0.488
5	0.84	0.415	43.969	0.75	0.433
6	0.84	0.394	43.432	0.75	0.450
7	0.83	0.389	41.407	0.77	0.455
8	0.81	0.427	35.093	0.76	0.445
9	0.80	0.444	35.085	0.78	0.411
10	0.80	0.438	34.173	0.77	0.430
11	0.80	0.426	33.984	0.79	0.453
12	0.80	0.482	33.303	0.79	0.372
13	0.80	0.444	33.181	0.78	0.424
14	0.79	0.444	32.284	0.79	0.413
15	0.77	0.462	28.660	0.81	0.404
16	0.75	0.476	25.656	0.82	0.393

Figure 3. Experimental vs Predicted log BB values plot for the best model obtained in the present study (Eq. (24)).



378 3.0 CONCLUSION

379
380 Highly significant (r^2 ($n-1$) up to 0.83) statistical methods for the BBB entering potential of drugs
381 were achieved by applying the proposed method, which incidentally shed new light into BBB
382 penetration of drugs. In fact, the BBB passage was found related to the analyte charges, being
383 hindered for compounds supporting one or more acidic functions, and enhanced for bases.
384 Moreover, molecules with higher dipolar momentum and greater PSA seemed less prone to cross
385 the BBB. The relatively high number of analytes taken into account support statistically the
386 suitability of the method as early screening method to evaluate BBB passage, and consolidate the
387 novelty of the present work. In the modeling of drugs' BBB passage, both IAM and MLC indexes
388 are found advantageously suitable; however, their combination with physico-chemical descriptors is
389 highly beneficial for prediction. From a theoretical point of view, it should be considered that both
390 IAM and MLC indexes relate to BBB passage data despite the different interactions they depict as
391 confirmed by the lack of co-linearity between those two analytical indexes. Again, their
392 simultaneous use in the statistic models, here performed for the first time, improved their prediction
393 strength, thus suggesting that both play a relevant role in BBB passage although mirroring different
394 phenomena. The P-gp efflux has also been investigated, but our results indicate that it would affect
395 the overall BBB drug uptake only negligibly. However, this conclusion should be regarded
396 cautiously due to the aspect that only a few P-gp substrates were included in the set of analytes
397 considered. Furthermore, the molecular docking simulations suffer from several limitations, the
398 most important being the aspect that the receptor flexibility is not taken into account. This might
399 have played a role in the moderate predictivity of the *in silico* calculated P-gp binding affinities.
400 Finally, the proposed method is also suitable for pharmaceutical companies in the search for
401 accurate BBB penetration oriented screening methods as the chromatographic conditions were
402 carefully studied to obtain the indexes in a relatively short time such as to meet their demands.
403 Chromatographic indexes (MLC and IAM) were always included in the best statistical models; this

404 implies that the information encoded in such measures is original and cannot be satisfactorily
405 surrogated by other *in silico* descriptors. The molecular modeling performed was simple, easy-to-
406 perform and can be configured to run automatically in case of batch analyses. Furthermore, as the
407 method is rather cheap and relies on basic HPLC equipment, it offers potential for broad scale
408 application

409 **4.0 EXPERIMENTAL SECTION**

410

411 **4.1 CHEMICALS**

412

413 The solutes were obtained from Sigma-Aldrich (Machelen, Belgium), TCI-Europe (Zwijndrecht,
414 Belgium) and Thermofisher Acros Organics (Geel, Belgium) as listed in Table 1 and their purity
415 was equal to or higher than 98%.

416

417 **4.2 ANALYTICAL COLUMNS**

418 MLC and IAM experiments were performed on an Agilent Zorbax SB-C18 Rapid Resolution (3.5
419 μm , 50 mm x 2.1 mm; Santa Clara, CA, USA) and Regis IAM Fast Mini Screening (10 μm , 10 mm
420 \times 3.0 mm; Morton Grove, IL, USA) columns, respectively.

421 **4.3 APPARATUS**

422

423 **4.3.1 MLC-HPLC**

424 MLC chromatographic analysis was performed on an Alliance, Waters 2690 chromatograph
425 (Milford, MA, USA) with a quaternary pump and an automatic injector. A Waters 2487 dual-
426 wavelength absorbance ultraviolet detector was used. The applied detection wavelengths for the
427 various solutes were always in the range between 210 and 300 nm as listed in Table 1. Data

428 acquisition and processing were performed using a PeakSimple Chromatography Data System
429 (model 202) and PeakSimple software (SRI Instruments, Torrance, CA, USA). The temperature of
430 the analysis was controlled by a Polaratherm series 9000 unit (Selerity Technologies, Salt Lake
431 City, USA) and set at 37 °C. The flow rate was 1.0 mL min⁻¹ and the injection volume was 20 µL.

432 4.3.2. IAM-HPLC

433 IAM based chromatographic analysis was performed on an Agilent Capillary 1200 system (Santa
434 Clara, CA, USA). The system included a capillary pump, a micro vacuum degasser and an
435 automatic injector. An Agilent 1200 Series variable wavelength detector was used and set at the
436 maximum absorbance wavelength of each analyte. The IAM-HPLC experiments were carried out
437 at room temperature (20 ± 2 °C), the flow rate was 300 µL min⁻¹ and the injection volume was 1 µL.

438 4.4 MOBILE PHASE AND SAMPLE PREPARATION

439 MLC mobile phases were composed of aqueous solutions of 0.05 mol·L⁻¹ sodium dodecyl sulfate
440 (SDS) (Acros). Water (18.2 MΩ·cm⁻¹) was purified and deionized in house via a Milli-Q plus
441 instrument from Millipore (Bedford, New Hampshire, USA). pH was adjusted to pH 7.4 by
442 phosphate buffer, prepared with 0.05 mol·L⁻¹ disodium hydrogen phosphate (Sigma–Aldrich) and
443 potassium dihydrogen phosphate (Sigma–Aldrich). To reproduce the osmotic pressure of biological
444 fluids, NaCl (9.20 g·L⁻¹) (Sigma–Aldrich) was added to the micellar mobile phase. IAM mobile
445 phases consisted of a solution 70/30 v/v Dulbecco's phosphate-buffered saline (DPBS) / methanol
446 (HPLC-grade; Biosolve, Valkenswaard, The Netherlands). DPBS was composed of 2.7 mmol·L⁻¹
447 KCl, 1.5 mmol·L⁻¹ potassium dihydrogen phosphate, 137.0 mmol·L⁻¹ NaCl, and 8.1 mmol·L⁻¹
448 disodium hydrogen phosphate (Sigma–Aldrich). Such solution had a pH value of 7.40 ± 0.05, and
449 no pH adjustment was performed. All mobile phases were vacuum-filtered through 0.20 µm nylon
450 membranes (Grace, Lokeren, Belgium) before use. Different mobile phases and elution programs
451 were tested starting from 100% aqueous phase; however, in IAM-LC the latter condition did not
452 allow the elution of the most lipophilic bases in a reasonable amount of time. Stock solutions of all

453 drugs were prepared by dissolving 10 mg in 1 mL of methanol except for i) quinidine and
454 theobromine, for which stock concentrations of 1 mg·mL⁻¹ and 200 µg·mL⁻¹, respectively, were
455 used, ii) caffeine and theophylline, which were dissolved in water (10 mg·mL⁻¹), iii) domperidone,
456 which was dissolved in dimethyl sulfoxide (10 mg·mL⁻¹) and iv) chlorpromazine, which was
457 dissolved in acetonitrile. Stock solutions were stored at 4 °C, except for atenolol, zidovudine,
458 chlorambucil and rifampicin, which were stored at -20 °C. Working solutions were freshly
459 prepared at the beginning of each day by dilution, with the mobile phase, of the stock solutions to
460 50 µg·mL⁻¹ for all the analytes, except for valproic acid and halothane that were diluted to 250
461 µg·mL⁻¹.

462 4.5 DATA SOURCES

463 Log BB values were taken from the literature¹⁸⁻²⁴. pKa values were obtained from the literature²¹
464 except for amobarbital, donepezil, fluphenazine, hydroxyzine, ketorolac, paroxetine and ropinirole,
465 whose values were calculated by the software Marvin Sketch 15.1 for Mac OS X²⁸.

466 4.6 SOFTWARE

467

468 4.6.1 MOLECULAR MODELING

469 Molecular modeling was performed by the software Vega ZZ 3.0.5 for Windows-based PCs³¹. The
470 starting three-dimensional structures of the considered molecules were downloaded from PubChem
471 database³² and they were considered in both zero atomic charge and ionized form. The Gasteiger –
472 Marsili³³ method, along with CHARMM^{34,35,36} force field, was applied to calculate the atomic
473 charges required to perform the next molecular mechanics calculations. An extensive
474 conformational analysis was carried out *in vacuum* by using the Boltzmann Jump method
475 (MonteCarlo procedure) implemented in AMMP software³⁷ which generates 1000 geometries for
476 each compound by randomly rotating the rotors and the obtained lowest energy conformation was
477 further optimized by performing a PM7 semi-empirical calculation with MOPAC 2012 program³⁸

478 (keywords: PM7 PRECISE MMOK). A cluster analysis was performed to select the most populated
479 conformation states. Physico-chemical and topological/geometrical properties (Virtual logP³⁹,
480 lipole⁴⁰, volume, polar surface area, surface accessible to the solvent, gyration radius, ovality, mass,
481 number of atoms, angles, dihedrals, etc) were calculated by VEGA ZZ software and, finally, all
482 molecules were inserted into a Microsoft Access database.

483 The QSPR models were obtained by the automatic stepwise approach implemented in “Automatic
484 linear regression” script of VEGA ZZ software, calculating regression models, including from 1 to
485 5 independent variables. The predictive strength of the best equation was evaluated by leave-one-
486 out (LOO) cross validation and the regression coefficients were calculated to evaluate the set in
487 terms of standard deviation of errors, angular coefficient, intercept and r^2 of the trend line of the
488 chart of the predicted *vs.* experimental activities. Descriptors with too low regression coefficient (r^2
489 < 0.1) were excluded and collinear descriptors were detected by evaluating the variance inflation
490 factor (VIF) whose threshold value was set to 5. A further validation of the model having the
491 highest predictive strength was performed *via* model validator script, included in Vega.

492

493 4.6.2 MOLECULAR DOCKING

494 Molecular docking calculations were carried out using AutoDock 4.2 software⁴¹. The 3.4 Å
495 resolution P-glycoprotein (P-gp) crystallographic structure (mouse P-glycoprotein 3, gene: MD1A,
496 PDB code: 4Q9H) was downloaded from Protein Data Bank (PDB) Database. Gasteiger partial
497 charges were calculated on ligand atoms. Polar hydrogens were added to P-gp and Gasteiger³³
498 partial charges were calculated using AutoDock Tools⁴². Simulation boxes were centered on the
499 ligands in the structures of P-gp-ligand complexes (PDB codes: 4Q9I, 4Q9J, 4Q9K, 4Q9L) as
500 reported in the literature²⁷. The simulation boxes were adjusted to accommodate the ligand in each
501 complex and the sizes were between 26x26x26 Å and 30x26x30 Å. 100 runs for each simulation
502 were performed and the Lamarckian Genetic Algorithm (number of energy evaluation: 2.5×10^6)
503 for the docking simulations was used. The choice was based on the aspect that this protocol

504 provides the most efficient search for general applications, and is typically effective for systems
505 with about 10 rotatable bonds in the ligand. The acidic compounds having $pK_a < 7.4$ and the basic
506 ones having $pK_a > 7.4$ were considered in their dissociated forms. Gasteiger-Marsili³³ electric
507 charges were supplied. Amphoteric drugs were assumed in their prevalent forms as calculated by
508 the software MarvinSketch²⁸. For the analytes supporting one or more stereocenters, the following
509 arrangements were undertaken. When the drugs were administered as racemic mixture (Atenolol,
510 Citalopram, Donepezil, Eserine, Halothane, Hexobarbital, Hydroxyzine, Ibuprofen, Ketorolac,
511 Mianserin, Nicotine, Omeprazole, Oxazepam, Pindolol, Promethazine, Venlafaxine and
512 Verapamil), each stereoisomer was docked into each site of the P-gp and the binding energies
513 presented are the averages of those of the stereoisomers included in the mixtures. On the contrary,
514 when the log BB values referred to a specific stereoisomer (Rifampicine, Zidovudine,
515 Levofloxacin) as that was the one administered in the log BB determinations, only this was docked.
516 When a new stereocenter was created because of protonation, as for instance occurs for tertiary
517 amines supporting different groups, both configurations were tested. The consistency of the results
518 was analyzed by clustering spatially the docked conformations. This step was necessary because of
519 the stochastic nature of the search methods, that are used to predict optimal docked conformations.

520 4.7 PROCESSING

521 The chromatographic retention coefficients of each analytes were calculated by using the following
522 expression:

$$523 \quad k = \frac{t_r - t_0}{t_0}$$

524 in which t_r is the retention time of the compound of interest and t_0 the retention time of a non-
525 retained compound (acetone). All reported log k values are the average of at least three
526 measurements; for each log k value the 95% confidence interval associated with each value never

527 exceeded 0.04.

528 Three different sets of properties were generated. At first, all the analytes were considered as
529 uncharged (having full charge equal to 0), subsequently analytes having acidic or basic functions
530 were considered fully ionized and zwitterions were considered with both the acidic and basic
531 functions in their charged forms. Eventually, a weighted average of the static properties at pH 7.4
532 according to the pKa of each analyte was performed; for zwitterions, the relative abundance of each
533 microspecies (zero atomic charges, zwitterion, anion and cation) in solution at the physiological pH
534 (7.4) was calculated by the software Marvin Sketch 15.1 for Mac OS X²⁸. This approach was also
535 extended to the conformational analysis performed *in vacuum*, yielding three different sets of
536 conformational properties, i.e. i) conformational properties of the forms of the analytes having zero
537 atomic charges, ii) conformational properties of the ionized forms of the analytes, and iii) average
538 of the conformational properties at pH 7.4 according to the pKa of each analyte and the calculated
539 microspecies distribution for zwitterions. For each of the properties taken into account (Molecular
540 lipophilicity potential (MLP)³⁹, lipole⁴⁰, volume, polar surface area, superficial area, gyration
541 radius, ovality, volume diameter, dipolar moment, etc), minimum and maximum value, average,
542 range and standard deviation for each population of conformers were calculated and incorporated in
543 the statistical models. An additional deal of molecular descriptor were calculated by the software E-
544 Dragon³⁰.

545 **Conflict of interest disclosure**

546 The authors declare no competing financial interest.

547

548 **LIST OF ABBREVIATIONS**

549

550 *CNS* Central Nervous System; *D/Dr05* distance/detour ring index of order 5; *Improvers* Number of
551 improper angles (out of plane); *HbDon* Number of H-bond donor groups; *HbAcc* Number of H-

552 bond acceptor groups; *IAM* Immobilized artificial membrane; *MD* Dipole Moment (Debye); *MLC*
553 Micellar liquid chromatography; *MLP* Molecular Lipophilicity Potential; *nRCOOH* number of
554 carboxylic group (aliphatic); *PLS* Partial Least Squares; *Psa* Polar Surface Area (\AA^2); *P-gp* P-
555 glycoprotein; *SDS* sodium dodecyl sulphate; *TPSA (NO)* topological polar surface area using N,O
556 polar contributions; *TPSA (Tot)* topological polar surface area using N,O,S,P polar contributions.
557

558 **Supporting Information**

559

560 - Figure 1 Page S-2;

561 - Figure 2 Page S-4;

562 - Figure 3 Page S-6;

563 - raw data in the spreadsheet file “data.xlsx”.

564 - AutoDock GPF file for site 1

565 - AutoDock DPF file for site 1

566 - AutoDock GPF file for site 7

567 - AutoDock DPF file for site 7

568

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