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Use of selected amendments for reducing metal mobility and ecotoxicity in contaminated bottom sediments

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40 Abstract

The aims of the study were 1) to assess the suitability of selected amendments for 41 reducing the mobility of metals in sediments by evaluating their effects on metal sorption 42 capacity, and 2) to assess the ecotoxicity of sediment/amendment mixtures. Three different 43 amendments were tested: cellulose waste, biochar, and dolomite. The efficiency of metal 44 immobilization in mixtures was dependent on pH, which increased with concentrations of 45 amendment. The higher negative charge observed for dolomite and cellulose waste 46 corresponded with greater attraction of cations and enhanced metal sorption. For cellulose 47 waste, the highest values of the Q parameter were attributed to the presence of OH groups, 48 which corresponded with the highest immobilization of metals. Biochar reduced the negative 49 surface charge, which highlights the importance of additional factors such as high specific 50 surface area and volume of pores in metal immobilization. All amendments increased the SSA 51 and V_{N2}, indicating a higher number of sorption sites for metal immobilization. Most 52 bioassays established a reduction of the ecotoxicity for amendments. Mixtures with dolomite 53 54 (25%, 45% doses) and biochar (45% dose) were low toxic. Mixtures with cellulose waste were toxic or highly toxic. The mobility of metals from contaminated sediments can be 55 56 limited by reused industry side products, which could contribute to further closing the circular economy loop. 57

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59 Keywords: metal fractionation, industry side products, immobilisation, sorption capacity,

60 biotest

61 **1. Introduction**

Metal contamination of bottom sediments poses a significant ecological risk. Metals 62 can persist in sediments for decades, and they can easily enter the food chain, posing a threat 63 to all levels of the biosphere (Peng et al., 2018; Baran et. al., 2023). Metal contamination also 64 limits the options for using dredged sediments outside of their original waterbody, such as for 65 agriculture, reclamation, and civil engineering projects (Renella, 2021). Because of the threat 66 67 to the aquatic environment, contaminated sediments require remediation. However, this is a complex issue due to the variety in physical and chemical properties of sediments. Therefore, 68 there is an urgent need to conduct remediation studies to develop effective and sustainable 69 70 solutions (Peng et al., 2018; Polrot et al., 2021).

Metal immobilization is a simple, efficient, and widely used remediation method for 71 metal-contaminated bottom sediments (Palansooriya et al., 2022; Wang et al., 2021). It 72 73 involves changing the physical and chemical properties of the sediments to make the metals less mobile. This is done by modifying the sorption complex of the substrate by adding 74 materials that improve the physicochemical parameters responsible for metal immobilization 75 (Zhang et al., 2015; Peng et al., 2018; Martínez-Nicolás et al., 2020). The goal is to move 76 metals from mobile to less mobile forms or to immobilize them completely. It is important to 77 78 note that immobilization does not reduce the total metal content of the sediments, but it does make the metals less accessible and less harmful. 79

To remediate metal-contaminated bottom sediments, it is important to consider not only the total metal content, but also the forms and fractions of the metals present. This is because the mobility of metals in the environment depends on their form. Soluble and exchangeable metal forms are considered to be mobile, potentially bioavailable, and toxic (Wojtkowska and Bogucki, 2022). The mobility of metals in bottom sediments is governed by the properties of the sediment, the environmental conditions, and the type and form of the

metals (Wojtkowska, 2013, Peng et al., 2018). Strong and relatively permanent binding of 86 87 metals occurs in combination with carbonates, phosphates, sulfates, and sulfides. Incorporation into the lattice structure of minerals can also occur as a result of coagulation, 88 flocculation, precipitation, or co-precipitation into insoluble forms (Wojtkowska, 2013; Zhang 89 et al., 2014). In contrast, the colloid and organic matter contents of sediments are key factors 90 influencing the ecologically most important type of interaction with metals, which is 91 exchangeable sorption. This process is reversible and depends on pH, redox potential, ionic 92 strength, and the structure and content of organic matter. Under unfavorable conditions, 93 metals may re-mobilize and become more toxic (Boguta et al., 2016). In the case of bottom 94 95 sediments, sorption processes can also be mediated by the presence of dissolved natural organic matter, including humic and fulvic acids (Boguta et al., 2016). The 96 mobility/immobility of metals in sediments also shows a direct relationship with some 97 98 physical parameters, such as porosity and the surface area available for adsorption. These parameters are a function of sediment granulometry (Brito et al., 2020; Otunola and Ololade, 99 2020). 100

Metal immobilisation is a widely studied and promising approach for remediating 101 metal-contaminated bottom sediments (Peng et al., 2018; Ma et al., 2019; Wikström et. al., 102 103 2021). While various materials have been investigated for metal immobilization, environmentally friendly solutions are constantly being sought, and materials of natural or 104 waste origin are increasingly promoted (Lwin et al., 2018; Palansooriya et al., 2019; Wang et 105 al., 2021). Metal immobilization in bottom sediments involves using materials with good 106 sorption properties that can affect pH and form hardly soluble compounds with metal ions. 107 According to the literature, metal immobilization can be achieved by introducing into bottom 108 sediments various substances that permanently or temporarily bind metal contaminants, such 109 as limestone, gypsum, zeolites, phosphorus compounds, lignite, and natural minerals (Lwin et 110

al., 2018). Recently, industrial side products have shown promise as amendments for further 111 reducing the mobility and bioavailability of metals in soil/sediments (Lwin et al., 2018; Wang 112 et al., 2021). Industrial by-products have been widely used beneficially in agriculture and 113 reclamation, and most have alkaline properties that can increase soil pH, reduce bulk density, 114 increase aggregate stability and water holding capacity, and add basic soil cations. They also 115 contain many macro and micronutrients with useful properties for enhancing soil quality 116 (Lwin et al., 2018; Antonkiewicz et al., 2019). To effectively reduce metal solubility while 117 preserving the physicochemical properties suitable for a bottom sediment substrate, it is 118 important to match the distinct properties of the sediment and amendment. An important 119 120 factor determining the success of metal immobilization processes is the effectiveness of the substance used to immobilize metal ions. However, there is a lack of data on the effective 121 modification of the properties of contaminated bottom sediments by the application of 122 industrial by-products characterized by the potential to immobilize many metals and reduce 123 their ecotoxicity. 124

The goals of the study were 1) to assess the suitability of selected amendments (cellulose waste, biochar, and dolomite) for reducing the mobility of metals in sediments by evaluating their effects on metal sorption capacity, and 2) to assess the ecotoxicity of sediment/amendment mixtures. Evaluating the relationships between sorption capacity, metals mobility and ecotoxicity is useful for managing contaminated bottom sediments and reducing the risk of introducing metals to the environment.

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- 132 **2.** Materials and methods
- 133 **2.1. Materials and experimental setup**

Bottom sediment was collected from one of the contaminated zones (dam zone) of theChechło reservoir. Using an Ekman sampler, five sediment samples within a 30-meter radius

were collected from a depth of 0 to 15 centimeter and stored at 4°C for transport. In the 136 137 laboratory, the samples were combined, air-dried, and ground.

Three amendments were tested: cellulose waste (CW), biochar (BC), and dolomite 138 (DL). Cellulose production waste was acquired from the Paper and Cardboard Factory 139 Beskidy S.A. in Wadowice, Poland (Antonkiewicz et al., 2018). Cellulose waste with code 140 030310 includes "fibre rejects, fibre-, filler- and coating sludges from mechanical separation". 141 It belongs to chapter 03 of the European waste Catalogue concerning "wastes from wood 142 processing and the production of panels and furniture, pulp, paper and cardboard", from sub-143 chapter 03 on "wastes from pulp, paper and cardboard production and processing" (European 144 145 Commission, 2014). The biochar came from the company Fluid SA in Sędziszów, Poland. Biochar was made of renewable resources, i.e. plants, wood production residue in sawmills, 146 or specifically grown energy willow and miscanthus, through FLUID technology. As opposed 147 148 to the classical pyrolysis process, this technology is an innovative method of thermal treatment of every type of plant biomass and biomass residues in an oxygen-free atmosphere, 149 at temperatures below 300°C. Dolomite was obtained from Kopalnia Józefka Sp. z o.o. in 150 Górno, Poland. This waste material is a byproduct of grinding dolomite for use as aggregate 151 in the construction industry. Amendments were air dried prior to the experiment, ground (0-152 153 0.2 mm) and homogenized.

Before the experiment started, a chemical analysis of the sediments and materials was 154 preformed (Table 1). Bottom sediment was characterised by an acidic pH, whereas cellulose 155 waste, dolomite and biochar were alkaline. The highest pH value of the cellulose waste 156 resulted from technological processes where calcium compounds are used for hygienisation of 157 this waste. Biochar had the highest content of total organic carbon (TOC). A high content in 158 Cd, Zn and Pb were found in the bottom sediments, whereas biochar had the highest content 159 in Cr, Ni and Cu. 160

The experimental design covered 4 treatments with a combination of 95%, 75%, 55% 161 162 bottom sediment and 5%, 25%, 45% of each waste material respectively, and a control sample which was only bottom sediment (100%). The proportion of sediment and amendments was 163 calculated based on their dry weight. Each mixture was placed in a 500 cm³ propylene 164 container and mechanically mixed for 24 hours until homogeneous blending (36 rpm). Next, 165 mixtures were hydrated to 45% maximum water holding capacity, and kept in the dark at 166 20°C for 116 days. Water losses were supplemented with distilled water once a week. The 167 incubation experiment was conducted in triplicate for each mixture. 168

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170 **2.2. Physico-chemical analysis**

In the bottom sediment, in the additives, as well as in the developed mixtures, pH was determined potentiometrically with a pH meter in 1 mol KCl/dm³ after 24 h in a 2:5 suspension of material and distilled water. TOC was measured with a CNS analyser (Vario EL Cube, Elementar Analysensysteme, Germany).

Fourier-transform infrared spectroscopy (FTIR) was performed on all mixtures prepared as a thin, pressed pellet containing 1 mg of sample, dried at 105°C and homogenized with 200 mg of KBr. Spectra were recorded using a Tensor 27 FTIR spectrometer (Bruker Optics, Germany) in the spectral range of 400-4000 cm⁻¹, with resolution of 256 scans at 2 cm⁻¹.

180 Negative surface charge (Q) and distribution function of dissociation constants 181 f(pKapp) were calculated from the raw data of potentiometric titration according to Szatanik-182 Kloc et al. (2009). Potentiometric titration was performed using an automated Titrino 702 SM 183 instrument (Metrohm, Switzerland) in a pH range from 3.0 to 10.0 using 0.1 *M* NaOH based 184 on 0.1 M NaCl as a titrant. Samples were prepared as suspensions of 100 mg of the dried solid mixture in 25 ml of 0.1 M NaCl, adjusted to pH 3.0, left on 24h and readjusted (if necessary)
to pH 3.0 on the next day.

Pore size distributions from 0.003 to 360 µm were measured on an Autopore IV 9510 187 mercury porosimeter (Micromeritics, Norcross, GA, USA). The equivalent pore diameter was 188 estimated from Washburn's equation: $D_{MIP} = -4\sigma_m \cos\theta_m / P_m$, where D_{MIP} is the pore diameter, 189 $\sigma_{\rm m}$ is the mercury surface tension (0.485 J m⁻²), $\theta_{\rm m}$ the mercury contact angle (assumed 130 190 degs) and P_m the external pressure (Pa). From the obtained pore size distributions, the total 191 pore volume (V_{MIP} , cm³ g⁻¹), total pore area (S_{MIP} , m² g⁻¹) and average pore diameter (D_{MIP} , 192 nm) were calculated. The samples' total porosity (P_c) was estimated according to the formula: 193 $P_c = V_{MIP} \rho_D 100\%$, where V_{MIP} is the total pore volume, and ρ_D is the sample bulk density 194 defined as the ratio of the sample dry mass to its volume (including pores) measured at 0.51 195 psia. 196

Sorption on nitrogen was measured on a 3Flex device (Micromeritics, Norcross, GA, 197 USA). Before the measurement, samples were heated to 105 °C and kept under a vacuum 198 overnight. From the experimental data, the specific surface area (SSA), volume of pores (V_{N2}) 199 and average pore diameter (D_{N2}) were calculated. The Brunauer-Emmett-Teller (BET) and 200 Barrett-Joyner-Halenda (BJH) models were adopted to estimate SSA and pore characteristics, 201 202 respectively. The SSA was estimated in a relative pressure range from 0.05 to 0.35. Pore characteristics (VN2 and DN2) were calculated in the capillary condensation range in 203 mesopores from the desorption curve. The area of the nitrogen molecule was assumed to be 204 0.162 nm² at 77 K. 205

The pseudo-total content of metals (Cd, Cr, Ni, Zn, Pb, Cu, Cr) in the bottom sediment, additives and mixtures was determined after *aqua regia* digestion (HNO₃ (65%) and HCl (30%) acids (3:1 v/v) (suprapure, MERCK, Germany)), using a dedicated microwave system at 180°C and 1200 W for 20 minutes (Mars 6, CEM, NC, USA). Fractions of metals (Cd, Cr, Ni, Zn, Pb, Cu) in the sediment/amendment mixtures were assessed using the BCR sequential extraction procedure (Rauret et al., 1999). Four fractions of metals are distinguished: F1 fraction – acid extractable (CH₃COOH at 0.11 mol/L and pH = 2); F2 fraction – reducible (extractable with NH₂OH.HCl at 0.5 mol/dm³ and pH = 1.5); F3 fraction – oxidisable, (extractable with hot 30% H₂O₂, and then re-extracted with CH₃COONH₄ at 0.5 mol/dm³ and pH = 2); fraction F4 – residual (digested using a hot mixture of HNO₃ and HCl (3:1 v/v)).

The metal content in each fraction was determined using an inductively coupled 217 plasma optical emission spectrophotometer - PerkinElmer ICP-OES Optima 7300 DV 218 219 (PerkinElmer, Inc., Waltham, MA USA) while total metal content was assessed with an iCAP 7400DUO ICP-OES from Thermo Fisher Scientific (MA, USA). The certified reference 220 materials CRM020 Sandy loam 2 trace element (MERCK, Germany) and BCR-701 221 222 (fractions) (LGC Standards, UK) were used for quality control. Recoveries with reference to the certified concentrations were 95% for Zn, 98% for Cu, 105% for Pb, 98% for Cr, 86% for 223 Cd, 113% for Ni (total content) and 93% for Zn, 93% for Cu, 86% for Pb, 119% for Cr, 89% 224 for Cd, 98% for Ni (BCR fractions). 225

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227 **2.3. Ecotoxicological analysis**

Ecotoxicity of the mixtures was assessed through three biotests: Phytotoxkit, Ostracodtoxkit F, and Microtox. In the Phytotoxkit test three plants were used: *Sinapis alba*, *Lepidium sativum*, and *Sorghum saccharatum*. The inhibition of root lengths of each plants were measured after incubation in the dark for 72 hours at 25 °C. In the Ostracodtoxkit F, the crustacean *Heterocypris incongruens* was used as test organism (Ostracodtoxkit F, 2001). After 6 days of incubation of *H. incongruens* with tested samples on multi-well plates at 25 °C in the dark, mortality and inhibition of growth were evaluated. Luminescent bacteria Alivibrio fischeri as test organisms was used in the Microtox. Inhibition of luminescence was
measured before and after 15 minutes of incubating the bacteria with the tested sample.
Microtox M500 Analyzer was used for the test (MicrobicsCorporation, 1992).

All tests were conducted according to the procedures developed by the manufacturer (Microbics Corporation, 1992; Ostracodtoxkit, 2001; Phytotoxkit, 2004). The mixture ecotoxicity was assessed using toxicity classes, based on the percentage effect (PE) of growth inhibition: class I (PE \leq 20%, non-toxic sample); class II (20% < PE \leq 50%, lowly toxic sample); class III (50% < PE < 100%, toxic sample); class IV (PE = 100% for at least one test, very toxic sample), class V (PE = 100% for all tests, very toxic samples) (Persoone et al., 2003).

245 **2.4. Statistical analysis**

The significance of differences between mean values of parameters were analysed using one way ANOVA and post-hoc Tukey's test at a significance level of 0.05. All statistical analyses were conducted using Microsoft Excel 2016 and the Statistica 13 software package.

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251 **3. Results**

252 **3.1. Properties of mixtures**

Selected physico-chemical properties of the mixtures are shown in Table 2. Amendment of mineral and organic materials to acid bottom sediment resulted in a significant increase in pH in all treatments compared to the control. The pH, depending on the treatment, was 3.58 in the bottom sediment and between 4.16 to 12.3 in the mixtures. Mixtures with cellulose waste were alkaline, with a pH as high as 12.3 for the highest dosage (45%). Slightly acidic (DL 5%), neutral (DL 25%) and alkaline (DL 45%) conditions were witnessed in the mixtures of bottom sediment and dolomite. The mixtures of bottom sediment and biochar were acidic to neutral (BC 45%). The additives affected the TOC, which ranged from 6.37 to 18.73%,content of the mixtures differently. TOC increased significantly in mixtures with biochar (all doses) and dolomite (doses of 25 and 45%). The mixtures based on biochar contained from 10 to almost 60% more TOC compared with the control treatment. Dolomite increased the TOC content in mixtures by 2 to 18%. In contrast, the addition of CW reduced the TOC content by 9% on average.

The FTIR spectra of the bottom sediment (Figure 1) shows the contribution of both 266 organic and mineral compounds in the studied material. Applied additives modified the 267 structure of the BS at some conditions. Cellulose waste addition caused a significant increase 268 in the absorption band at ~1425 cm⁻¹ that is assigned to vibrations in aliphatic C-H, aromatic 269 C=C, and C=O (in case of CW – to CH₂ bonds) (Oh et al., 2005; Liu et al., 2013; Boguta et 270 al., 2022), alongside a decrease in the band at ~1090 cm⁻¹, attributed to vibrations of Si–O–Si 271 272 and C-O-C of ethers (Kovač et al., 2005). Decrease in this band at the highest CW content revealed another one at 1039 cm⁻¹ attributed to primary alcoholic OH and CO polysaccharide 273 vibrations (Boguta et al., 2022). Addition of the biochar decreased the absorbance at ~3441 274 cm⁻¹ related to decrease in O-H stretching vibrations of carboxylic acids, phenols, alcohols, 275 and water, N-H bonds, H-bonded OH and intermolecular bonded OH (da Silva et al., 2018). 276 BC additions also contributed to a decrease in the ~1630 cm^{-1} band, attributed to C=O 277 stretching vibrations of deprotonated COO⁻, C-O and C-N stretching in amides, and 278 vibrations of -C=C- on benzene rings (da Silva et al., 2018). BC, similarly to CW, decreased 279 the band at ~1090 cm⁻¹; this drop was however not as marked. Increasing dolomite fractions 280 caused a reduction in bands at ~3441 cm⁻¹, ~1630 cm⁻¹ and at ~1090 cm⁻¹. As in the case of 281 CW, the addition of DL increased the intensity of absorbance at ~ 1425 cm⁻¹. All additives 282 lowered the signal height at ~ 780 cm⁻¹, 530 cm⁻¹ and 470 cm⁻¹, derived mainly from 283 vibrations of Si-O-Si and aromatic C-H structures. This was less noticeable with the addition 284

of BC. The data obtained show that the application of BC, DL and CW additives clearly influenced both the organic and inorganic matrix of the BS resulting mainly in the decrease of the contribution of siloxane, polysaccharide and alcoholic structures.

The potentiometric titration of the bottom sediment revealed the presence of various 288 functional groups generating negative surface charge (Q) at different pH conditions (Figure 289 2). The highest increment in O was observed in the lower pH range, indicating the importance 290 of acidic functional groups such as the carboxyl group (Skic et al., 2016). The increase in Q 291 was less pronounced in neutral pH ranges, before proportionally rising again in the alkaline 292 ranges, which indicated the contribution of e.g. OH groups (Sokołowska et al., 2011). The 293 294 additives influenced the Q value in different ways. Treatments CW 25% and 45% increased this parameter and the strongest effect was observed above pH 6.0. The Q values for both CW 295 5% and BC 5% additives did not differ from the control sample, BS. Interestingly, successive 296 297 increasing concentration of the biochar from 25 to 45% caused a decrease in Q as compared to treatment BS. This reduction concerned the charge generated in the lower pH ranges. The 298 addition of dolomite seemed to decrease the Q value, especially at low pHs ranges, but no 299 clear trend could be observed with this amendment. 300

Distribution function of dissociating constants (f(pKapp)) of the bottom sediment 301 302 (Figure 3) showed that the most abundant population includes the functional groups dissociating at $pK_a \sim 3.5$, as well as these that dissociate under alkaline conditions (broader 303 spectrum of dissociating constants). The clearest influence of amendments was noted when 304 adding cellulose waste at 25%, where a shift of the "acidic" maximum to the pK_a value of 305 ~6.25 was observed, alongside an increase in an "alkaline" maximum, indicating important 306 changes in acidic species for the enriched samples. Interestingly, the increase in biochar in the 307 mixtures caused flattening of the distribution curve of dissociation constants in the entire 308 range of pK_a values, indicating a fairly similar share of functional groups over a wide 309

spectrum of dissociation constants. The addition of dolomite at 5% resulted in a decrease in 311 the signal at pK_a 4.75, while higher amounts of DL caused an additional shift of the main fraction of dissociation constants to pK_a of about 6.0. 312

The differential and cumulative mercury intrusion porosimetry (MIP) curves obtained 313 for mixtures of bottom sediment with cellulose waste, biochar, and dolomite are shown in 314 Figure 4. All cumulative curves are s-shaped with a rapid increase in volume at a pore 315 316 diameter of about 6 µm. Noticeably, biochar increased the pore volume of the analysed mixtures proportionally to amendment rates, while dolomite, in contrast, reduced it 317 significantly. The effect of cellulose waste was equivocal, showing a decrease in the pore 318 319 volume only for low and medium concentrations. The differential pore size distributions show unimodal patterns, with the main peaks observed around 0.02 to 6 µm. In the case of BC 320 mixtures, the peak size increased with amendment concentrations. It is also slightly shifted 321 322 towards pores of a greater diameter. In contrast, a higher concentration of DL diminished peak height. 323

The effects of additives on pore characteristics measured by MIP and N₂ adsorption 324 methods are presented in Table 2. Total porosity (P_c) ranged from about 47.8% to over 55.3%. 325 The highest average value was noticed for treatments BC 25% and 45%, while the lowest was 326 327 for DL 45%. Addition of biochar lowered the pore surface area (S_{MIP}) and bulk density (ρ_D) to respectively 7.2 m²·g⁻¹ and 0.83 g·cm⁻³ at the mixture 45%. In turn, dolomite increased bulk 328 density and diminished the total pore area compared to the control. S_{MIP} was highest, at 21.5 329 m²·g⁻¹, for cellulose waste (CW 45% variant). The value of D_{MIP} ranged from about 473 nm to 330 over 1225 nm and was higher than the control for higher concentrations of biochar. In 331 treatments CW 5% and 45%, D_{MIP} was lower, whereas D_{MIP} was similar to the control for 332 mixtures with dolomite. The N₂ adsorption results revealed that all amendments had a 333 significant impact on the BET surface area (SSA) and cumulative pore volume (V_{N2}). These 334

335 generally increased compared to the bottom sediment, except for the highest dose of dolomite.

336 It is worth noting that the patterns were not always consistent with increasing additive dose.

337 For example, the average pore diameter (D_{N2}) was usually lower in the mixtures than in the

control, but this difference was statistically insignificant in most cases (α =0.05).

339

340 **3.2.** Content and mobility of metals in mixtures

341 The total content of metals in the treatments depended on the metal, type of additive, and its dose (Table 3). It was found that the contents of all metals decreased with the addition 342 of cellulose waste and dolomite. One of the possible explanations for this relationship could 343 344 logically be attributed to a dilution of the metal content during mixing, coincidentally accompanied by alkalinisation due to high amendment pH. Treatments with 45% cellulose 345 waste and dolomite had the lowest metal content, 1.4 to 2.4 times lower than the control. 346 347 Mixtures with biochar had different effects, significantly reducing Cd, Zn, and Pb, but increasing Ni, Cu, and Cr by 1, 9, and 28%, respectively. The BC 45% mixture had the 348 highest content of these latter elements. 349

However, relying solely on the total metal content in the mixtures does not allow to 350 reliably assess metal mobility and risks of release into the environment. Metal fractionation 351 352 and speciation provides a more accurate picture. Information on the most mobile, easily available metal fractions is particularly relevant (Wang et al., 2021). BCR sequential 353 extraction is an essential tool in this context. It is commonly assumed that the following three 354 metal form categories are distinguished based on the results of: (a) easily mobile/bioavailable 355 (F1), (b) potentially mobile/bioavailable (F2 and F3) and (c) biologically unavailable metals 356 (F4) (Devi and Saroha, 2014). Other authors reported that metals in the exchangeable fraction 357 (F1) are more mobile and bioavailable, while the potential mobile fraction of metal fractions 358 F1, F2 and F3 (Σ F1-3), are more bioavailable than the residual (F4) (Baran et al., 2019). 359

Metals fraction F4 are stable, bound within a structure of some primary and secondary minerals, and can therefore not impact organisms. In contrast, metal forms found in fraction F1 are easily interchangeable, water- and acid-soluble, and carbonate-bounded. It happens that the metals in this latter fraction are easily bioavailable, rendering this fraction this most risky to the environment (Liang et al., 2017). As mentioned above, the purpose of the additives introduced into the contaminated bottom sediments was to immobilise metals through the reduction of their mobility.

The F1 fraction differed considerably for the different tested metals under the 367 influence of amendments (Table 3, Figure 5). Cadmium was the most mobile element among 368 369 the analysed metals, as evidenced by its very high share in fraction F1 in the unprocessed sediment (75%). The share of cadmium in the remaining fractions, depending on the 370 treatment, was average or small and ranged from 14 to 37% (F2), from 4 to 45% (F3) as well 371 372 as from 1 to 25% (F4) of the total content. All amendments decreased this fraction, with varying efficiency. Treatments with cellulose waste seemed the most effective, with the F1 373 share impressively dropping as low as 1% for the 45% dose, corresponding to a 60 times 374 lowering compared with the control. Furthermore, CW 25% dosage resulted in a significantly 375 lower Cd fraction in F1 than for 45% of either biochar or dolomite. Interestingly, treatments 376 377 with these two other materials still reduced the % share of Cd in F1 by a factor 1.3 to 1.7. Decreases in F1 were accompanied by increases in the three other fractions, most notably F3 378 and F4. In treatments with 5% share of CW, BC and DL, the reduction in Cd mobility was 379 insignificant. 380

Patterns for Zn were similar. Also for this element, the F1 fraction in the sediment was high. Cellulose waste was again the most effective amending material, impressively lowering the F1 fraction from 71% in the control to 5% for the 45% amendment rate. Additionally,

significant decreases of 1.8 and 1.9 times were achieved with 45% BC and DL, respectively.
Diminished F1 fraction were mostly accompanied by increases in F3 and F4.

Nickel displayed a relatively lower F1 fraction in the unprocessed sediment, which 386 387 was still as high as 26%. All amendments demonstrated significant efficiencies for reducing this fraction, with fairly similar values (2.6 to 3.7 times lowering) for all mixtures at 25 and 388 45%. Both cellulose waste and dolomite even performed significantly well at 5% dosage 389 already (2.2 and 2.6 times lowering, respectively). Decreases in F1 were mostly accompanied 390 by increases in fractions F3 and F4. F2 was even decreased as well (below 1%) for DL 391 treatment at 25 and 45%. Overall, only the BC 5% treatment did not seem to show any 392 immobilizing effect. 393

It is widely considered that Pb, Cu and Cr have a rather low mobility and they occur 394 dominantly in fractions 3 and 4. This was witnessed here as well, these metals being much 395 less present in fraction F1 in the unprocessed sediments (1 to 8%). Fraction F2 was also lower 396 for Cu and Cr than for other metals. The influence of amendments was less marked than for 397 398 other metals. For Cu and Cr, a significant reduction in the F1 fraction could be achieved for all amendment dosages except for cellulose waste 5% (Cr) and biochar 5% (Cu, Cr). This 399 decrease was less marked than for Cd or Zn, and dolomite seemed the most effective material. 400 As for previously discussed metals, this was accompanied by enrichments in fractions F3 and 401 F4, less noticeable than for the others given how predominant these fractions were initially. 402 For Pb, finally, amendments led to various patterns in fraction distributions. In contrast with 403 other metals, no reduction in F1 could be achieved; this fraction remained limited however, 404 ranging from 1 to 3%. Immobilization seemed the most effective under DL treatments, 405 406 whereby 25% and 45% led to a reduction in F2 and F3, accompanied by a higher share in F4. A notable enrichment in fraction F2 was noticed for BC treatments, but this seemed less 407

408 marked at 45% (1.5 times increase) than at 5% (2.6 times increase). Cellulose waste
409 irregularly led to an enrichment in F3, with a high 80% at 45% dosage.

410

411 **3.3.** Assessment of metal immobilisation with indices

The effectiveness of these additives for metal immobilisation was further assessed 412 using three factors (Table 3): Risk Assessment Code (RAC), Individual Contamination Factor 413 414 (ICF) (Liang et al., 2017; Baran et al., 2019; Matabane et al. 2021) and Ecological Risk Factor (ERF) (Tytła and Kostecki 2019). In RAC, the percentage share of the metals associated with 415 fraction F1 is considered relatively to their total content. ICF was calculated by dividing the 416 417 sum in potentially mobile fractions (Σ F1-3) by the residual fraction (F4), while ERF was calculated based on dividing the sum of metal contents in fractions F1 and F2 by their 418 contents in fractions F3 and F4. A common characteristic of all 3 factors is that the lower their 419 420 value, the less mobile and bioavailable a given metal will be. From then, different classes of risks have been defined based on these indices. Value ranges for these classifications are 421 provided under Table 3. 422

From the RAC classification, it appears that cellulose waste decreased metal mobility 423 significantly more than dolomite or biochar, for a variety of elements and doses. Treatments 424 425 with CW at 25% and 45% doses led respectively to a moderate and low risk of releasing Cd. In contrast, other mixtures were associated with a very high (BS, CW 5% dose, BC at all 426 doses, DL 5 and 25% doses) to high (DL 45%) risk regarding the mobility of this metal. A 427 similar trend was seen for Zn, with 25% and 45% CW treatments resulting in moderate and 428 low risks, while others were linked with high (CW 5%, BC 25% and 45% doses, DL 25 and 429 45% doses) or very high (BS, BC 5%, DL 5%) risks. For Ni, a high risk relating to the 430 mobility of this metal was determined only in the 5% BC mixture, and a moderate risk was 431 found in the BS and CW 5% mixture, with a low risk for other treatments. Regardless of the 432

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treatments, Cu was characterised by a low risk. The risk of release of Pb in the majority of the
treatments was low; mixing with 25 and 45% cellulose waste even resulted in no risk.
Chromium was the least mobile element, with no risk observed for all mixtures expect the
original sediment and treatments with CW and BC at 5% dosage (low risk).

The individual contamination factor (ICF) is used to assess the ability of an element to 437 be released from the sediment/soil samples under different environmental conditions (Naji 438 and Sohrabi, 2015). Different physicochemical properties of a sediment or a mixture may 439 cause differences in ICF values. On average, it seemed that amendments led to a decrease in 440 ICF when compared with the control, pointing out a decrease in their mobility. However, DL 441 442 performed seemingly better than the other. In fact, BC treatments were the least effective in reducing ICF, even showcasing increases for Cd at 5% dose. The above relationship was also 443 found for Pb in treatments with 45% CW and BC, and for Cu in treatments with 25% and 444 445 45% cellulose waste (Table 3, Figure 5). Higher ICF values in these treatments indicate a higher tendency to release bioavailable metals to the environment relative to the control. Due 446 to the fact that ICF reflects the risk of metal pollution samples exhibited a very high and 447 considerable contamination for Cd (except CW 25%, DL 45%), Zn (except CW 45%, DL 448 45%), Pb only for CW 45%. Moderate contamination degree was recorded for Ni (except DL 449 450 25%, DL 45%, CW 5%), Cu (except CW 5%), Pb (except DL 25%, DL 45%, CW 45%), and for Cd (CW 25%, DL 45% and Zn (CW 45%, DL 45%). Low contamination rate observed for 451 Cr (all treatments), Ni (CW5%, DL 25%, DL45%), Cu (CW 5%), and Pb (DL 25%, DL 45%). 452 The above observations were partially confirmed by ERF that, similar to ICF, pointed 453 out that the additives and their increasing doses generally reduced the value of this factor 454 relative to the control treatment (Table 3). Lower ERF values indicate the increase in the 455 metal contents in less mobile fractions F3 (associated with organic matter) and F4 (residual 456 fraction), and thus the reduction of the risk of their release to the environment. This 457

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relationship was not demonstrated for Pb in the treatments with BC (all doses), CW and DL 458 (in 5% dose), as well as for Cd in the treatments with 5% share of CW and DL, and for Ni and 459 Cu in the treatments with 5% dose of BC. For Cd (except CW45%) and Zn (all doses of CW), 460 most study treatments showed a high risk of mobility of these metals in the environment. For 461 Cd (except CW45%) and Zn (all doses of CW) most study treatments showed a high risk to 462 the mobile of the environment. The remaining treatments had a moderate or low risk relating 463 464 to the release of the analysed metals (Table 3). Given the average ERE values, the highest effectiveness in decreasing the mobility of the metals (excluding Pb) with respect to the 465 control (BS) was found in BS+CW mixtures, then in BS+DL mixtures, and the lowest 466 467 effectiveness was determined in BS+BC mixtures.

468

469 **3.4. Ecotoxicity of mixtures**

470 The amendments significant reducted ecotoxicity of the sediment/amendment mixtures in most cases. Responses varied importantly according to the amendment, dose, and test 471 organism (Table 4). The highest reduction in toxicity for plants was obtained with dolomite, 472 for which stimulation of S. saccharatum root growth was even observed at 25% and 45% 473 doses. It was noticeable that CW seemed to alleviate toxicity at lower doses (5 and 25%), only 474 475 for the toxicity to increase again for the 45 % rate. The Ostracodtoxkit test revealed most of the mixtures were toxic to *H. incongruens*, with a lower toxicity achieved for treatments CW 476 25%, BC 45% and DL 25 and 45%. Finally, the inhibition of luminescence of A. fischeri 477 showed the widest spectrum in sensitivity, with PE ranging from -30 to 100%. Treatments BC 478 45% and DL 25 and 45% performed best again, even leading to stimulation of the 479 luminescence. Interestingly, DL could already reduce toxicity 2.4 times at 5 % dosage. In 480 contrast, CW barely alleviated toxicity, the 45 % dose being as highly toxic as the control (PE 481 = 100%). 482

These various responses among test organisms can be further visualized in Figure 6. 483 Mixtures based on cellulose waste seemed to trigger more toxicity in A. fischeri than other 484 treatments. Contrastingly, treatments with biochar and dolomite proportionnaly led to more 485 toxicity to *H. incongruens*, and to a certain extent to *S. alba* (for DL in particular). Among 486 plants, monocotyledons S. saccharatum turned out to be the less sensitive species, and S. alba 487 the most. The majority of the treatments were non-toxic or of low toxicity to the former 488 (similarly as for L. sativum), while they were deemed lowly toxic to toxic for the latter. 489 Another studies also showed that on the substratum with different wastes S. alba was the plant 490 most sensitive to the effect of chemical substances in these wastes, and Sorghum saccharatum 491 492 was less sensitive (Baran and Antonkiewicz, 2017; Antonkiewicz et al., 2019). In the studies of Szara et al. (2020) and Gondek et al. (2023) H. incongruens was the most sensitive 493 organism to the substance present in the tested mixtures based on bottom sediment or 494 495 diatomite different waste. H. incongruens is a very attentive organism to the acid pH, and, the oral ingestion is the main route of exposure of this organism to toxic substances (Mariani et 496 al., 2022). Differences in sensitivity of the test organisms were related not only to their 497 species or trophic group, but also to the method of exposure to toxic substances (Szara et al., 498 2020; Heise et al., 2020; Gondek et al., 2023). In the Phytotoxkit and Ostracodtoxkit biotests, 499 500 the organisms are exposed to both soluble and contaminants absorbed on the solid particles of the mixtures, while the exposure of A. fischeri (Microtox) are limited to bioavailable, easy-501 soluble compounds. 502

The 5 tests considered altogether, a lower toxicity class could be achieved in all treatments except for cellulose waste 45%. This latter mixture, like the unprocessed sediment, were deemed highly toxic (class IV), and would pose serious threats in the environment. In contrast, mixtures of bottom sediment with dolomite (in 25% and 45% doses) and biochar (in 45% dose) were assigned to toxicity class II, meaning that these samples present a low

toxicity. The remaining mixtures (DL 5%, BC 5 and 25%, and CW 5 and 25%) were
assigned to class III, which means their toxicity would still represent an environmental threat.

511 **4. Discussion**

512 **4.1. Impacts of the amendments on metal immobilisation**

The metal content in the residual fraction (F4) generally increased with increasing 513 dolomite rates. One possible reason for this is that the low and medium dolomite doses 514 increased the number of binding sites for metals, as evidenced by the increase in SSA and V_{N2} 515 compared to the bottom sediment control. In contrast, the decrease in Pc and V_{MIP} could be 516 517 attributed to the filling of the bottom sediments' skeletal pores with fine particles of high density dolomite (Gruszecka-Kosowska et al., 2017). In accordance with Chen et al. (2007), 518 the specific surface area was the key factor determining Cu(II) adsorption onto the surface of 519 acid-activated dolomite. Salameh et al. (2010) suggested that one of the mechanisms involved 520 in As(V) adsorption was the rough surface of dolomite, which provides contact points for 521 522 chemisorption at low metal concentrations. In line with Emami et al. (2016) and Qui et al. (2015), dolomite surfaces are rough and irregular, show significant porosity, and provide 523 functional groups capable of metallic species complexation. Our studies also showed that the 524 525 highest dose of dolomite caused a marked decrease in SSA and V_{N2} . Diwan et al. (2020) observed a similar effect when studying the adsorptive removal of uranium from an aqueous 526 phase with dolomite. They observed enhanced removal efficiencies up to a specific dolomite 527 dose, followed by a decrease in removal capacity, attributed to a reduction in total surface 528 area due to partial aggregation of adsorbent particles. The decrease in Q values after adding 529 530 the lowest dose of DL (5%) was possibly due to the blocking of O-containing functional groups on the BS by the Mg and Ca ions of the DL (Figure 2C). The highest doses of DL 531 caused an increase in the Q relative to the 5% DL treatment (Table 2), which could be due to 532

the higher contribution of HCO_3^{-1} and CO_3^{-2-1} ions. This increase in Q could also be partly due 533 to the increase in the dissociation of the biochar functional groups with increasing pH of the 534 mixtures. A higher negative charge on the surface of functional groups reduces competition 535 for active sites between metal and protons, making it easier to attract cations, thus enhancing 536 metal sorption. This immobilizing effect at DL 25% and 45% doses was observed for all 537 analysed metals, as evidenced by the increased F4 fraction (Figure 5). Indeed, related studies 538 539 show that the main mechanism for metal retention in systems with dolomite can be precipitation of carbonates (Radziemska, 2018). 540

In certain conditions, a further increase in the pH can adversely affect sorption 541 542 processes due to the weakening of electrostatic attraction forces resulting from the increasing conversion of metal cations to hydrolysed species (Diwan et al., 2020). Additionally, 543 negatively charged forms, such as chromate (VI), have less opportunity to sorb due to 544 545 increased repulsion of negatively charged ions (Khoshraftar et al., 2023). The importance of controlling the pH was also confirmed by Pehlivan et al. (2009) in studies on adsorption of Cu 546 and Pb on dolomite, in which the highest efficiency of cation removal via surface 547 complexation and ion exchange was observed at pH 5, while these processes were reduced 548 above pH 6 due to hydrolysis. 549

550 The addition of cellulose waste significantly increased the surface properties SSA and V_{N2} . The fraction of metals associated with organic matter (F3) also increased, suggesting that 551 metals were immobilized on the outer surface of the cellulosic material, including the walls 552 and/or mouths of small pores (Boguta et al., 2022). Hydroxyl groups on the outer surface of 553 cellulose increases its ability to adsorb and react with various chemical moieties (Godage and 554 Gionfriddo, 2020). The lowest dose of cellulose waste raised the pH of the mixture to 7.84, a 555 value at which dissociation of phenolic OH groups is still limited (pK_a ca. 8.2–8.3; Su et al., 556 2013). An earlier study of the sorption properties of cellulose waste confirmed the relatively 557

low efficiency of the OH groups of cellulose materials in metal binding at lower pH (Jamshed
et al., 2017). This is because more protons compete with metal cations for active sites at lower
pH. Moreover, at lower pH, the number of active sited is reduced because of the overall
reduced netto negative surface charge.

In our cases, the higher additions of cellulose waste increased the pH of the materials 562 to 8.96 and 12.3, promoting the dissociation of organic OH groups. Indeed, the cellulose 563 564 waste amendment, especially at dose of 25%, caused an increase in Q values (Figure 2a). This change was attributed to the presence of OH groups, as evidenced by the shift of the maxima 565 of the distribution function of dissociation constants toward the higher contribution of the 566 567 charge generated in alkaline pH ranges (Figure 3a) (Szatanik-Kloc et al., 2019). The highest values of Q for the cellulose waste treatment 25% treatment corresponded to the highest 568 immobilization of Cd, Pb and Zn in F4 fraction, which suggests that OH groups are involved 569 570 in metal binding. At the pH of this treatment (pH=8.96), the above metals reveal positively charged forms: Cd²⁺, Cd(OH)⁺, Pb²⁺, Pb(OH)⁺, Zn²⁺, ZnOH⁺, which are readily attracted to 571 negative surface charges. Under these conditions, there is also the possibility of precipitating 572 metals as hydroxides, for example, Zn(OH)₂. Such forms could also be a reason for 573 enrichment in stable forms. The immobilization of Cd and Pb for cellulose waste 45% in F4 574 575 was lower as compared to cellulose waste 25%, which is likely due to the presence of a higher number of negatively charged forms of metals, such as $Cd(OH)_3^{-}$ and $Pb(OH)_3^{-}$, at strongly 576 alkaline conditions (pH=12.3) and their repulsion with dissociated OH groups of the cellulose 577 578 waste.

Biochar interacts with metals in various ways, including outer-sphere complexation, inner-sphere complexation, electrostatic interactions, surface precipitation, and ion exchange (Shaheen et al., 2019; Wang et al., 2021). In our studies, the gradual decrease in Q values upon addition of the BC was likely due to the increase in the number of structures poor in

oxygen functional groups, which to a large extent undergo thermal degradation during biochar 583 production at temperatures above 300°C. At the same time, biochars derived from high-584 585 temperature biomass exhibit an increased content of non-volatile cations (Wang et al., 2015). These cations can attach to bottom sediment functional groups, blocking the negative charge. 586 587 The decrease in the value of Q with the simultaneous decrease in the content of metals in the 588 most mobile fraction (F1), except Pb, and the increase in metals in F4 show that metals are 589 bound in more stable forms, and immobilization does not have to be dependent on binding by groups generating a negative charge. 590

591 Considering the well-developed surface area of biochar, which was observed in the mixtures, it is likely that this mechanism depends to some extent on adsorption onto the 592 porous biochar structure. The outer surface of biochar, followed by the interior pores, may 593 cause the cation exchange capacity to shift and enhance element retention (Ahmad et al., 594 2014). This is also supported by other studies on the sorption of metals with the use of 595 biochars (Inyang et al. 2012; Kim et al., 2013; Tomczyk et al., 2020). Simultaneously, the 596 interaction and distribution of heterogeneous biochar particles in the bottom sediment matrix 597 affected the structure of the prepared mixture. Biochar typically has a lower bulk density than 598 599 soil and sediments, so it reduces the mixture density through a mixing or dilution effect (Blanco-Canqui, 2017). The specific surface area of biochar is comparable to or higher than 600 that of clay minerals (Downie et al., 2009), so the use of biochar as an amendment generally 601 resulted in a net increase in SSA and S_{MIP} compared to control bottom sediment. Additionally, 602 the observed increase in total pore volume with a simultaneous decrease in bulk density 603 604 suggests that biochar contained a significant proportion of macropores (Wildman and Derbyshire, 1991; Brewer et al., 2014). This group of pores may affect the transport processes 605 within the bottom sediments (Boguta et al. 2022) and thus improve access to metal adsorption 606 607 centers.

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4.2. Impact of the amendments to ecotoxicity of mixtures

609 Combining different biotests (Phytotoxkit, Ostracodtoxkit, and Microtox) to polluted bottom sediments can be a useful tool for evaluating a range of environmental risks 610 611 (Mierzwa-Hersztek et al., 2018; Heise et al. 2020; Baran et al. 2023). From an ecotoxicological point of view, bottom sediment proved to be the most toxic material, posing 612 a severe environmental threat as demonstrated by its class IV toxicity. This high toxicity may 613 be due to two factors: the acidic reaction of the sediment and its high content in Cd, Pb and 614 Zn. The pH of the bottom sediment (3.58) is below the tolerance range for many organisms. 615 Additionally, pH is one of the main factors determining the solubility and mobility of metals 616 617 in the environment (Liang et al., 2017, Koniarz et al., 2022). The balance of sorption and desorption processes depends on the reaction of protons and metal cations. Metal solubility 618 conditioned by exchangeable sorption processes is low in a range of neutral and alkaline pH, 619 and increases with decrease in pH value (Kim et al., 2015). This increase in solubility in 620 acidic environments is associated with the slow dissolution of iron, aluminum and manganese 621 622 oxides and the release of metals from primary and secondary minerals. The acid pH of the bottom sediment may have increased the mobility and bioavailability of the tested metals to 623 the test organisms, which is why bottom sediment was shown to be the most toxic material. 624

625 Despite a significant reduction in metal mobility, cellulose waste mixtures were highly toxic to test organisms. Studies by other authors also showed high ecotoxicity of 626 substrates and mixtures containing cellulose production waste (Antonkiewicz et al., 2019; 627 Szara et al., 2020). In the studies of Szara et al. (2020), the mixture of bottom sediment and 628 629 cellulose waste (BS + CW) was assigned to toxicity class IV (toxic sample, high acute 630 hazard). Wastes from cellulose production not only contain many valuable components, including organic compounds and macro- and microelements necessary for plants, but are also 631 characterised by a significant content of toxic compounds (Baran and Antonkiewicz, 2017; 632

Antonkiewicz et al., 2019). Wastes from cellulose production not only contain many valuable 633 634 components, including organic compounds and macro- and microelements necessary for plants, but also toxic compounds (Baran and Antonkiewicz, 2017; Antonkiewicz et al., 2019). 635 For example, they may contain chlorinated organic compounds which are used to bleach 636 paper and are highly toxic by causing carcinogenic, clastogenic, mutagenic and endocrine 637 effects (Hag et al., 2022). Another factor contributing to ecotoxicity of cellulose waste is 638 639 bleach residues in this material. Chemicals used to bleach paper include sodium hypochlorite, soda lye, and chloroacetic acid (MCAA), hydrogen peroxide, ozone, and molecular oxygen 640 (Haile et al., 2021). Unfortunately, no specific analysis to detect toxic organic components 641 could be performed during the current study. Additionally, it should be noted that the strong 642 alkaline nature of the cellulose waste resulted in a high pH in mixtures, especially at 45% 643 dosage (pH = 12.3). This is much above endurable values for living organisms, and this 644 645 alkalinity very likely contributed to the ecotoxicity of the mixtures.

In recent years, there has been increasing interest in the use of biochar to immobilise 646 metals in the environment (Wang et al., 2021; Palansooriya et al., 2022). The addition of 647 biochar has been shown to reduce the toxicity of bottom sediments in a dose-dependent 648 manner. Many studies have shown that biochar can stimulate the growth of organisms due to 649 650 its potential content of macro- and micro-elements and the partial bioavailability of the carbon contained in its matrix. In addition, biochar has a porous structure and can be an excellent 651 microenvironment for living organisms (Gondek et al., 2016; Palansooriya et al., 2019). On 652 the other hand, biochar may contain various toxic compounds besides heavy metals, e.g. 653 polycyclic aromatic hydrocarbons (PAHs), volatile organic compounds (VOCs) and dioxin, 654 which may rather lead to ecotoxicity (Godlewska et al., 2021). Toxicity might also occur 655 through specific physicochemical properties such as particle size, pH and/or salinity 656 (Palansooriya et al., 2019; Godlewska et al., 2021). In our case, the pH of the mixtures was 657

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increasingly closer to ideal neutral values, which might be a reason why 25% and 45% doses 658 659 reduced ecotoxicity significantly. Huang et al. (2017) observed that the most mobile fraction (F1) of Cd and Zn decreased when sediment was amended with biochar; however, a dosage 660 rate as low as 50 mg kg⁻¹ was shown to decrease enzymes activity and microbial abundance. 661 It is worth noting that biotests allow not only for the detection of a toxic substance, but also 662 for a full assessment of the interaction of all factors and substances, taking into account 663 synergism or antagonism (Oleszczuk et al., 2013; Heise et al., 2020). Thus, even though no 664 specific pollutant analysis could be carried out in our case, it can be reasonably assumed that 665 BC was not contaminated to an extent that would result in toxicity. 666

667 Dolomite is the most common material used in agriculture for neutralizing soils as a mean to improve plant growth. It has also been assessed for the remediation of areas and 668 materials contaminated with heavy metals (Khoshraftar et al., 2023; Gondek et al. 2023). In 669 670 this study, the addition of dolomite to the contaminated sediments contributed to the highest reduction of the ecotoxicity of the mixtures for the test organisms, especially at 25 and 45% 671 doses. Two test organisms even underwent a stimulating effect on root growth (S. 672 saccharatum) and luminescence (A. fischeri) as compared to the control at 45% amendment 673 rate. In the studies of Gondek et al. (2023), a reduction in the toxicity was also found under 674 675 the influence of dolomite, as they witnessed a class I toxicity level for mixtures of diatomite and dolomite. This is consistent with earlier findings that demonstrated the high potential of 676 dolomite to reduce the availability and toxicity of heavy metals in contaminated soils and 677 water (Sidiri, 2018; Vrînceanu et al., 2019; Khoshraftar et al., 2023). In our case, dolomite 678 helped achieving a near-neutral pH in all mixtures, which might be a main factor towards 679 It may also be that dolomite increased nutrient availability for 680 reducing ecotoxicity. microbes, or positively influenced plant nutrition as a source of magnesium and calcium 681 (Baggs et al., 2010; Shaaban et al., 2017). 682

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Research on living organisms is an important part of research on the toxicity of materials. Such analysis is complementary to chemical analysis and is therefore necessary for the proper assessment of risk related to the release of materials into the environment (Oleszczuk et al., 2013; Szara et al. 2020).

- 687
- 688 **4.3. Implications for remediation**

The effectiveness of the immobilisation process usually depends on the characteristics 689 of the sediment and the materials used (Polrot et al., 2021). The interaction of the 690 immobilisation agents with the metals and with the properties of the sediment is important as 691 692 this will primarily determine the immobilisation efficiency and the short and long term environmental impact. Our studies found that amendments such as biochar, dolomite and 693 cellulose waste can be recommended for the remediation of metal-contaminated sediments. 694 Different doses of the products tested showed good efficacy as immobilising agents 695 (especially for Cd, Zn and Pb) and often resulted in a significant reduction of the initial 696 697 ecotoxicity. The materials significantly neutralised acidity of the bottom sediment (biochar, dolomite, cellulose waste) and biochar in addition contributed significant amounts of organic 698 matter (biochar) to the sediment/amendment mixtures. These materials significantly improved 699 700 the physico-chemical properties of the bottom sediments through an increase in pH improv sorption properties, and accordingly reduced mobility of metals. The industrial by-products 701 702 also reduced the ecotoxicity of the amended bottom sediments. Other advantages can be added to these benefits, such as their quality as soil conditioners, low cost, convenient 703 704 applicability, and economic competitiveness (Lwin et al., 2018). These three materials are 705 processed industrial by-products that are produced in large amounts each year. Using these materials for bottom sediment remediation has the advantage of solving a waste disposal 706 problem, preventing potential environmental pollution in the future (Lwin et al., 2018). 707

Industrial by-products may contain other hazardous substances in high concentrations. 708 709 These can be of concern when the by-products are considered for use in remediation (Lehoux et al., 2020). Currently, Polish legislation lacks regulation about the use of bottom sediments 710 in agriculture. This closest criteria for assessing the permissible content of metals would be 711 existing restrictions on municipal sewage sludge (Cd - 20 mg/kg; Cu - 1000 mg/kg; Ni - 300 712 mg/kg; Pb - 750 mg/kg; Zn - 2500 mg/kg; Cr -500 mg/kg DM (Regulation 2015, 2015)). 713 While most metals in the mixtures comply with these values in our case, Cd (and Zn, in most 714 cases) does not. Valorization would then be restricted to non-agricultural purposes, compost 715 production (Regulation 2015, 2015), or construction purposes. 716

There are not many single technologies that can remediate bottom sediments 717 efficiently (Ma et al., 2019). Different remediation approaches should be adopted for different 718 types of contaminated sediments. Our previous studies have attempted to remediate 719 720 contaminated bottom sediments using physical and biological methods (thermal treatment processes and phytoremediation) (Koniarz et al., 2022; Śliwa-Cebula et al., 2023). Further 721 studies are needed to improve the environmental adaptability of remediation technologies in 722 actual field conditions, and to develop a multistep remediation technology for heavy metal-723 contaminated sediments (Ma et al., 2019). 724

725

726 **5.** Conclusion

The impact of different amendments and their application rate on metal mobility and ecotoxicity in contaminated bottom sediments was assessed through various physicochemical analyses. These revealed a strong dependence of metal immobilization efficiency on on pH, which increased with all amendment additions. Dolomite and cellulose waste, with their high negative surface charge, caused an enhanced metal sorption. For cellulose waste, the presence of OH groups further contributed to its highest metal immobilization, as reflected by the

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highest Q value. Biochar, however, reduced the negative surface charge, suggesting the involvement of other factors like its high specific surface area and pore volume in metals immobilization. Most amendments increased the parameters SSA and V_{N2} compared to bottom sediments, indicating a higher number of binding sites for metals.

Most amendments reduced the ecotoxicity of the sediments. Mixtures with 25% and 45% dolomite and 45% biochar even achieved a low toxocity classification (class II). Cellulose waste, however, showed limited success. All mixtures remained classified as toxic (class III) or highly toxic (class IV), similar to the original sediment. Despite its superior performance in metal immobilization, its inability to mitigate toxicity significantly limits its potential reuse for sediment remediation.

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Figures 990

Figure 1. FTIR spectra for mixtures of bottom sediment (BS) with increasing content of 991 amendments: A) cellulose waste (CW); B) biochar (BC); C) dolomite (DL). 992







- 996 Figure 2. Cumulative negative surface charge (Q) of the bottom sediments (BS) in function of
- the pH and increasing content of additives: A) cellulose waste (CW); B) biochar (BC);
 dolomite (DL). Error bars represent standard deviation (n=30).



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1002 Figure 3. Distribution function of surface dissociation constants of the bottom sediment (BS)
1003 samples as a function of apKs and increasing content of additives: A) cellulose waste (CW);
1004 B) biochar (BC); dolomite (DL). Error bars represent standard deviation (n= 30).



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Figure 4. Cumulative (left) and differential (right) pore size distribution $(0.003-360 \ \mu m)$ determined by mercury intrusion porosimetry for bottom sediment (BS) and mixtures with increasing concentration of cellulose waste (CW), biochar (BC), and dolomite (DL)



- Figure 5. Metal fractionation according to the BCR sequential extraction scheme in bottom sediments
- (BS) and mixtures with cellulose waste (CW), biochar (BC), and dolomite (DL)



Figure 6. Share of the different biotests in the cumulated toxic response for the bottom sediments (BS) and mixtures with cellulose waste (CW), biochar (BC), and dolomite (DL)



1024 Table 1. **pH, total organic carbon (TOC), and selected metal contents of the bottom**

1025 sediment and amendments used in the mixtures

	Para	neter	BS*	CW	BC	DL		
	pH	KCl	3.55	12.50	7.31	8.53		
	TOC	g/kg dm	71.1±1.4	38.4±2.7	375±31	62.8±5.8		
	Cd		54.1±8.5	0.17±0.04	0.50±0.03	0.45±0.15		
	Cr		35.5±16.7	56.5±10.5	87.2±6.9	6.0±0.8		
	Ni	mg/kg dm	30.6±3.2	20.6±5.8	24.4±1.3	3.9±0.1		
	Zn		4409±75	51.6±6.7	219±31	46.6±1.7		
	Pb		279±21	2.6±0.3	28.1±3.9	3.3±0.2		
4007	Cu	1	68.0±16.9	8.9±0.2	43.8±10.5	15.4±0.9		
1027	*BS – bottom se	diment, CW– cellt	ilose waste, BC – bioc	har, DL – dolomite				
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		TOC	Q7	Q10	Pc	ρd	DMIP	Smip	VMIP	SSA	V _{N2}	D _{N2}
Treatments	рНксі	g/kg dm	μm	ol/g	%	g/cm ³	nm	m^2/g^1	cm ³ /g	m²/g	mm ³ /g	nm
BS	3.58 a ¹	76.3 a	622 e	1268 d	53.4 cd	0.97 c	835d	13.2 c	0.55 e	7.9 b	26.6 ab	10.3 bc
CW 5%	7.84 d	72.2 a	617 e	1177 d	52.6 c	1.04 de	541ab	15.6 cd	0.51 cd	11.4 c	26.4 a	9.0 ab
CW 25%	8.96 e	71.5 a	1080 g	2044 f	50.1 b	1.10 f	653 abcd	14.2 cd	0.45 b	15.1 d	37.6 c	9.3 ab
CW 45%	12.3 f	63.7 a	962 f	1600 e	53.7 cd	0.99 cd	473 a	21.5 e	0.54 de	17.6 e	42.8 de	8.8 ab
BC 5%	4.16 ab	85.1 b	528 d	1197 d	54.3 cd	0.96 c	626abc	16.6 d	0.57 e	17.8 e	42.8 de	8.6 ab
BC 25%	6.19 b	136.8 d	350 a	796 ab	55.3 cd	0.90 b	792 cd	16.3 d	0.62 f	14.7 d	31.6 b	7.9 ab
BC 45%	7.04 c	187.3 e	343 a	763 a	55.3 cd	0.83 a	1225 e	7.2 a	0.67 g	17.6 e	38.0 cd	7.8 ab
DL 5%	6.48 b	77.8 a	406 b	914 bc	54.3 cd	0.99 cd	682 bcd	15.7 cd	0.55 e	21.5 f	45.5 e	7.4 a
DL 25%	7.23 cd	86.2 cb	450 bc	1016 c	52.3 c	1.06 ef	661 abcd	13.6 c	0.49 bc	25.5 g	60.2 f	8.3 ab
DL 45%	7.35 d	92.8 c	464 c	1172 d	47.8 a	1.25 g	658 abcd	9.9 b	0.38 a	5.9 a	24.1 a	12.3 c

Table 2. Selected chemical and sorption properties of bottom sediments and mixtures

¹Means followed by different letters indicate significant differences at $\alpha \le 0.05$ according to Tukey's test. BS – bottom sediment; CW – cellulose waste; BC – biochar; DL – dolomite. TOC – total organic carbon; Q7 and Q10 – negative surface charge at pH 7 and 10; P_c - total porosity; ρ_D – bulk density; D_{MIP} – average pore diameter; S_{MIP} – total pore area; V_{MIP} – cumulative volume of pores; SSA – specific surface area; V_{N2} – cumulative volume of pores measured by nitrogen adsorption method; D_{N2} – average pore diameter measured by N₂ method

Treatments	Cd	Zn	Pb	Ni	Cu	Cr
			Total conten	t (mg/kg dm)		
BS	54.1 d ¹	4409 d	279 d	30.6 cd	68.0 bcd	35.5 cd
CW 5%	47.5 cd	3858 c	252 cd	29.5 cd	66.7 bcd	35.2 cd
CW 25%	33.8 b	2782 ab	176 b	21.4 b	57.2 ab	26.4 ab
CW 45%	22.9 a	1935 a	120 a	16.0 a	48.2 a	19.2 a
BC 5%	48.4 cd	3986 cd	267 cd	31.3 cd	75.4 d	41.5 cd
BC 25%	32.1 b	2747 ab	184 b	27.5 cd	71.7 cd	49.0 de
BC 45%	29.6 b	2638 ab	181 b	33.8 d	76.2 d	57.4 e
DL 5%	49.3 cd	3974 cd	261 cd	29.6 cd	66.8 bcd	34.2 cd
DL 25%	43.7 c	3293 bc	235 с	24.7 с	58.4 bc	32.7 cd
DL 45%	30.1 b	2517 a	164 ab	20.0 b	45.0 a	23.6 ab
			RAC (% in to	otal content me	tals bound to F	F1) ²
BS	75.0 d	70.8 c	1.0 a	26.0 b	7.8 c	0.9 b
CW 5%	70.1 d	49.7 bc	3.2 b	11.7 a	2.7 ab	0.7 b
CW 25%	21.3 b	30.2 b	0.8 a	9.7 a	5.0 ab	0.0 a
CW 45%	1.2 a	5.0 a	0.1 a	8.2 a	6.1 b	0.0 a
BC 5%	71.1 cd	54.1 bc	2.8 b	31.6 b	9.6 c	1.1 b
BC 25%	62.4 cd	44.9 b	1.8 ab	10.2 a	3.2 ab	0.0 a
BC 45%	55.6 c	40.1 b	2.0 ab	7.0 a	3.0 ab	0.0 a
DL 5%	67.0 cd	56.3 bc	3.3 b	10.0 a	2.9 ab	0.0 a
DL 25%	58.2 cd	49.2 b	2.2 ab	8.2 a	2.1 ab	0.0 a
DL 45%	42.9 c	36.7 b	2.0 ab	6.9 a	0.8 a	0.0 a
			ICF (∑F1-F3))/F4 ³		
BS	97.0 d	48.0 d	2.36 c	1.30 b	2.35 c	0.59 b
CW 5%	19.0 b	3.89 ab	1.55 b	0.86 a	0.96 a	0.28 ab
CW 25%	2.46 ab	3.36 ab	1.24 b	1.20 ab	2.69 c	0.31 ab
CW 45%	5.23 ab	2.18 a	5.24 d	1.11 ab	2.68 c	0.57 b
BC 5%	98.0 d	3.30 ab	1.62 b	1.25 ab	1.20 ab	0.26 ab
BC 25%	16.1 b	3.44 ab	2.95 c	1.33 ab	1.61 b	0.46 ab
BC 45%	6.69 ab	3.29 ab	2.05 c	1.09 ab	1.48 ab	0.44 ab
DL 5%	49.0 c	9.90 c	1.67 b	1.18 ab	1.64 b	0.35 ab
DL 25%	5.67 ab	4.00 b	0.47 a	0.81 a	1.17 ab	0.26 ab
DL 45%	2.77 a	2.37 a	0.65 a	0.82 a	1.23 b	0.16 a
			ERF (F1+F2)	/(F3+F4) ⁴		
BS	9.89 cd	8.35 c	0.18 b	0.46 c	0.12 b	0.03 b
CW 5%	10.59 cd	2.07 b	0.48 c	0.26 abc	0.04 a	0.03 b
CW 25%	1.37 ab	1.00 ab	0.18 b	0.22 abc	0.06 a	0.01 ab
CW 45%	0.52 a	0.25 a	0.04 a	0.13 ab	0.07 a	0.01ab
BC 5%	6.62 b	2.08 b	0.33 bc	0.58 d	0.14 b	0.02 ab
BC 25%	6.86 b	1.62 ab	0.68 d	0.25 abc	0.05 a	0.02 ab
BC 45%	4.17 ab	1.62 ab	0.50 c	0.24 abc	0.04 a	0.02 ab
DL 5%	11.86 d	3.46 ab	0.45 c	0.25 abc	0.03 a	0.02 ab
DL 25%	3.30 ab	1.44 ab	0.06 a	0.11 ab	0.02 a	0.00 a
DL 45%	1.66 ab	1.06 ab	0.04 a	0.09 a	0.01 a	0.00 a

2 Table 3. Content of metals in mixtures and assessment of their mobility

¹Means followed by different letters indicate significant differences at $\alpha \leq 0.05$ according to Tukey's test. BS – bottom sediment; CW – cellulose waste; BC – biochar; DL – dolomite. , ${}^{2}RAC \le 1\%$ no risk, $1\% < RAC \le 10\%$ low risk, $10\% < RAC \le 30\%$ medium risk, $30\% < RAC \le 50\%$ high risk, 50% < RAC very high risk, ${}^{3}ICF \le 1$ low contamination, $1 < ICF \le 3$ moderate contamination, $3 < ICF \le 6$ $considerable \ contamination, \ ICF > 6 \ very \ high \ contamination, \ ^40 < ERF \le 0.4 \ low \ risk, \ 0.4 < ERF \le 1 \ medium \ risk, \ 1 < ERF \ high \ risk \$

8 Table 4. Ecotoxicity of mixtures based on the response of test organisms and classification of

9 their toxicity

Treatments	SsRI	SaRI	LsRI	HiGI	AfLI	Class of		
Treatments	Percentage effect (PE, %)							
BS	57 d	84 d	80 e	83 cd	100 e	IV		
CW 5%	43 cd	47 b	15 abc	69 bc	84 d	III		
CW 25%	17 ab	40 ab	13 abc	36 a	91 d	III		
CW 45%	33 bc	65 c	41 d	76 bc	100 e	IV		
BC 5%	40 cd	72 cd	71 e	85 cd	98 e	III		
BC 25%	22 b	49 bc	27 bc	89 d	63 c	III		
BC 45%	15 ab	38 ab	14 abc	36 a	-30 a	II		
DL 5%	22 b	32 a	31 c	71 bc	41 b	II		
DL 25%	-32 a	33 a	8 ab	50 a	-16 a	II		
DL 45%	-7 a	27 a	5 a	45 a	-3 a	II		

¹Means followed by different letters indicate significant differences at $\alpha \le 0.05$ according to Tukey's test. BS – bottom 10

11 sediment; CW - cellulose waste; BC - biochar; DL - dolomite. ²SsRI - Sorghum saccharatum, roots growth inhibition, 12 LsRI - Lepidium sativum, root growth inhibition, SaRI - Sinapis alba, roots growth inhibition; HiGI - Heterocypris incongruens, growth inhibition; AfLI – Allivibrio fischeri, luminescence inhibition. ³Toxicity classes: class I (PE ≤ 20%, non-13 14 15 toxic sample); class II (20% < PE \leq 50%, lowly toxic sample); class III (50% < PE < 100%, toxic sample); class IV (PE =

100% for a single test, very toxic sample), class V (PE = 100% for all tests, very toxic samples)

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