| 1 | Embracing plant-plant interactions - retininking predictions of species range sinits |
|--------|---|
| 2 | Pieter Sanczuk ^{1*} , Dries Landuyt ¹ , Emiel De Lombaerde ^{1,2} , Jonathan Lenoir ³ , Eline Lorer ¹ , Miska |
| 3 | Luoto ⁴ , Koenraad Van Meerbeek ^{5,6} , Florian Zellweger ⁷ and Pieter De Frenne ¹ |
| 4 | |
| 5 | ¹ Forest & Nature Lab, Department of Environment, Ghent University, Melle-Gontrode, Belgium |
| 6 | ² Research Institute for Nature and Forest, Brussels, Belgium |
| 7 8 | ³ UMR CNRS 7058 "Ecologie et Dynamique des Systèmes Anthropisés" (EDYSAN), Université de Picardie Jules Verne, Amiens, France |
| 9 | ⁴ Department of Geosciences and Geography, University of Helsinki, Helsinki, Finland |
| 10 | ⁵ Department of Earth and Environmental Sciences, KU Leuven, Leuven, Belgium |
| 11 | ⁶ KU Leuven Plant Institute, KU Leuven, Leuven, Belgium |
| 12 | ⁷ Swiss Federal Research Institute WSL, Birmensdorf, Switzerland |
| 13 | *Corresponding Author: Pieter Sanczuk, Pieter.Sanczuk@UGent.be |
| 14 | Acknowledgments |

Pieter Sanczuk and Pieter De Frenne received funding from the Ghent University grant
BOF23/GOA/019. Dries Landuyt is being funded by a postdoctoral fellowship of the Flemish Research
Foundation FWO (1200321N). Florian Zellweger was supported by the Swiss National Science
Foundation (Grant Number 193645).

19 **Conflict of interest**

20 The authors declare that there is no conflict of interest.

21 Author contribution

All authors contributed to the concept and design of the article; Pieter Sanczuk prepared the manuscript together with Pieter De Frenne and Miska Luoto for the introduction, and Pieter De Frenne, Dries Landuyt, Emiel De Lombaerde and Florian Zellweger for the concept development. Jonathan Lenoir and Koenraad Van Meerbeek performed the SDM review. Pieter Sanczuk performed all analyses of the worked example and prepared the figures. All authors provided input on each section and reviewed the final version of the manuscript.

Data Availability Statement

30 All data and code used in the worked example is available on FigShare (Sanczuk et al., 2024)

31

32 Abstract

Interactions among plants are changing across the globe resulting from a multitude of changes in
 the environment. Obtaining accurate predictions of plant species' range dynamics require us to
 account for plant-plant interactions, but this remains challenging using the existing species
 distribution modelling (SDM) techniques.

- Advanced SDM techniques facilitate the integration of plant species interactions based on species to-species associations. However, for uncharted environmental conditions in which the formerly
 derived species' correlations potentially no longer hold, a more process-based alternative is
 expected to become increasingly relevant.
- We first review the most common SDM techniques that integrate plant-plant interactions, and then
 present the concept for a novel map product: a spatial Plant-plant Interaction Index (PII) depicting
 the link between a focal species' performance and the trait signature of the interacting vegetation.
 The latest developments in remote sensing and the increasing availability of vegetation plot data
 facilitate PII mapping based on vegetation trait-environment relationships.
- 46 4. Synthesis PII mapping holds the potential to advance next-generation biogeographical analyses as
 47 it can serve as a pivotal missing covariate layer necessary for the integration of plant-plant
 48 interactions into SDM applications. This data product adds flexibility to the ecologists' toolbox to
 49 analyze species range shifts and the formation of novel communities as a response to multiple
 50 environmental changes.
- Keywords: plant-plant interactions, biogeography, competition, environmental change, experimental
 macroecology, spatial model, species distribution model, vegetation survey

53 **1. Introduction**

54 Species are rearranging distributions across the globe brought about by changes in the environment 55 (Zellweger et al., 2020). Changing environmental conditions alter colonization and local extinction dynamics of species, resulting in biogeographical shifts at the larger geographic extent (Lenoir et al., 56 57 2020) and the emergence of new assemblages of interacting species that do not yet coexist at present 58 (De Frenne, 2023). Such 'novel communities' will become increasingly apparent in response to 59 uncharted sets of environmental conditions, often with unexpected functional implications to people 60 and nature (Alexander et al., 2015a, 2016; Kerr et al., 2023; Williams & Jackson, 2007). Interactions 61 among plant species, for example, can significantly reduce (e.g., due to competition) or enhance (e.g., 62 due to facilitation) range shifts under climate change (Alexander et al., 2015b; HilleRisLambers et al., 63 2013; Lyu & Alexander, 2022), but are often neglected when projecting future distributions. If we are 64 to adequately anticipate climate-change impacts on biodiversity and ecosystem functioning, we cannot 65 longer ignore the mediating effects of plant-plant interactions on range shifts.

66

67 Accounting for plant-plant interactions into projections of species ranges is complex. It requires us to 68 quantify how changes in biotic interactions, through acting on local population dynamics, impact 69 species' distributions at the continental extent. Species distribution models (SDMs) provide a 70 quantitative analytical framework that allows the prediction of species range shifts due to climate and 71 other environmental changes (Guisan & Thuiller, 2005; Merow et al., 2013), but considering species 72 interactions into SDMs remains difficult (Dormann et al., 2018; Norberg et al., 2019). Advanced 73 modeling techniques, such as direct interaction covariates (Pellissier et al., 2010) and joint SDMs 74 (Ovaskainen & Soininen, 2011), facilitate the integration of biotic interaction effects into projections of 75 species ranges based on species-to-species associations. Unfortunately, these methods often require an 76 *a priori* selection of data on the set of assumed interacting species, while the residual covariance among 77 species is easily confounded (Zurell et al., 2018), and, even more important, there is virtually no method 78 to parameterize these models in the absence of present-day analog communities (i.e., when species do 79 not yet co-occur; but see (Rohr et al., 2010)). Alternatively, biotic interaction effects have been 80 approximated in SDMs using surrogate variables (i.e., variables assumed to correlate with the strength 81 of plant-plant interactions, such as productivity or species richness (Wisz et al., 2013)). Surrogate 82 variables, however, do not necessarily remain constant under a changing environment, making 83 projections beyond the environmental range that was sampled more ambiguous. Furthermore, it is 84 unlikely that surrogate variables will accurately describe interaction strengths within more hidden strata 85 of plant communities such as for forest understorey plants below dense tree canopies and for 86 communities in ravines, on cliffs and on steep mountain slopes (Morueta-Holme et al., 2023).

88 To expand the toolbox of ecologists, we need a complementary alternative that adds flexibility and 89 facilitates a more general integration of plant-plant interactions in the prediction of species range 90 dynamics. Ecological data are increasingly available at the continental and global extent, including 91 species distribution, abundance and trait data, paving the way to step beyond the mere correlative nature 92 of the existing methods. An experimentally informed approach to account for species interactions within 93 SDMs holds the potential to overcome several of the known limitations. Leveraging the strengths of 94 both experimental research and correlative SDMs will likely advance our understanding of plant-plant interaction effects on species' range dynamics. 95

96

97 Here we review the most prominent existing methods that allow the approximation of plant-plant 98 interactions in SDM analyses. Second, we present the concept, development and potential application 99 of a novel gridded map product: a Plant-plant Interaction Index (PII) map depicting the realized 100 estimated impact of plant-plant interactions experienced by a focal plant species. The PII metric is 101 quantified based on experimental research and links a focal species' performance to the trait signature 102 of co-occurring species, and is mapped based on trait-environment relationships. PII maps hold the 103 potential to advance next-generation biogeographical predictions, serving as a pivotal missing covariate 104 layer necessary for the integration of plant-plant interactions effects into modern SDM applications.

105 Table 1 | Schematic representation of the basic principles and the main characteristics of three common species distribution modelling (SDM) techniques, and 106 the proposed plant-plant interaction map as calculated based on field data from interaction-exclusion experiments, that allow integrating plant-plant interaction 107 effects into predictions of the habitat suitability of a focal species (species Y in the diagrams)

| | Direct interaction covariates | Surrogate variables for interactions | Joint SDMs | Plant-plant Interaction Index |
|---|--|--|--|---|
| | Abundance or predicted suitability of species X ₁ Environmental variables | Surrogate variables Species Y Environmental variables | Traits, phylogeny Species community variables | Experiment-informed realized impact of plant interactions Environmental variables |
| Target species | Single species | Single species | Multiple species | Single or multiple species |
| How are species interactions modelled? | Abundance or predicted suitability of interacting species as predictor | Surrogate variables as predictor | Interactions are not incorporated in the model, but can be inferred from the model | Experimentally-informed surrogate (index) variable as predictor |
| Data needed | Abundance/occurrence data of target species or community data | Abundance/occurrence data of target species (or community data) | matrix of species presence/absence or abundance data | Performance data of target species in presence/absence of competitor(s) |
| Can trait data be incorporated in the model? | Yes, community-weighted mean traits if community data are available | Yes, community-weighted mean traits if community data are available | Yes, species-specific trait data help to estimate species niches | Yes, one or multiple traits of target species and interacting species |
| Can phylogenetic data be incorporated in the model? | Possible if community data are available, but not standard | Possible if community data is available, but not standard | Yes | No |
| A priori knowledge about species interactions necessary? | Yes, knowledge about interacting species is necessary | Yes, knowledge about interactions is necessary to select surrogate variables | No | preferentially informed by experiments |
| Applicable to rare species? | Difficult to estimate species niche for rare species | Difficult to estimate species niche for rare species | Rare species borrow information from abundant species | Difficult to estimate species niche for rare species |
| Type of interactions | Asymmetric interactions only | Asymmetric interactions only | Symmetric interaction only | asymmetric interaction only |
| Applicable to large communities? | Yes, but many interaction covariates could lead to multiple testing and overfitting | Yes, interactions are summarized in surrogate variables | Yes, latent variables substitute high-dimensional covariance matrix | Yes, interactions are summarized in surrogate index |
| Applicable to big data? | Yes, but computationally intensive | Yes, but computationally intensive | Joint modelling of species reduces run time | Yes |
| Spatial consistency of interactions assumed? | Yes (not if species-environment associations are included as predictors) | No | No | No |

| Temporal consistency of species | Yes | Yes | No | No, the plant interaction index can |
|---------------------------------|--------------|----------|----------|-------------------------------------|
| interactions assumed? | | | | be projected under environmental |
| | | | | change scenarios |
| Prediction of unobserved | Not possible | Possible | Possible | Possible, based on trait- |
| interactions | | | | environment associations |

110 2. Accounting for plant-plant interactions into projections of species' ranges: a review of 111 existing methods

112 Species distribution models (SDMs) are conventionally employed to analyze the relationship between 113 the distribution of a species - usually represented by presence-only, presence-absence or abundance data 114 - and environmental factors. However, these models do not distinguish between the influence of biotic 115 and abiotic environments (Araújo & Guisan, 2006). Several methods to overcome this issue and 116 approximate species interactions in SDMs exist, ranging from methods directly accounting for the 117 distribution of a selection of co-occurring species as covariates in the model to methods using covariates 118 acting as surrogates for biotic interactions (Table 1). Furthermore, technological progress in numerical 119 ecology has enhanced our ability to adapt statistical models to large community datasets (in so-called joint SDMs), thereby facilitating the consideration of both the environmental preferences of several 120 121 species and their interdependence simultaneously.

122 2.1. Direct interaction covariates

123 Utilizing the abundance of one or several **co-occurring species as predictor variables** in SDMs stands 124 as a primary method to account for biotic interaction effects on range predictions of a focal species. 125 Various interaction types have been explored through this approach, such as the consideration of 126 interspecific competition among plant species by adding plot-level predictors representing frequency or 127 count data (Leathwick & Austin, 2001; Pellissier et al., 2010), basal area (Rouget et al., 2001), or 128 proportion data (Meier et al., 2010) of co-occurring species. However, this method is sensitive to false 129 absences of the co-occurring species used as a predictor in the model which introduces bias to the model 130 predictions. An alternative method involves the use of model outputs from the SDM of the interacting 131 species as a covariate layer into SDM predictions of the focal species, effectively leveraging predicted 132 suitability (Leathwick & Austin, 2001). This method has advantages over the former approaches as it 133 minimizes bias attributed to false absences in the co-occurring species. Yet, inaccurate predictions may 134 in turn introduce other errors to the model.

135

136 While the use of empirical data (e.g., plot-level abundances) or predicted data (e.g., modelled habitat 137 suitability) of co-occurring species as predictors presents a straightforward technique, it also bears 138 certain limitations inherent to its simplicity. First, these methods model the unidirectional effect of the 139 co-occurring species on the focal species, and thus fail to capture the bidirectional nature of most biotic 140 interactions. For instance, the fitness benefits arising from mutualistic interactions can extend the ranges 141 of both involved species (Afkhami et al. 2014). Species' co-occurrence may be attributed to indirect 142 interactions, wherein both species respond to an external species, such as a common predator or 143 herbivore (Dormann et al. 2018). A shared response to an unmeasured environmental variable could 144 also result in a positive correlation between co-occurring species, even if species do not directly interact 145 (Poggiato et al. 2021). Issues may further arise when the predictor species and the focal species share a 146 similar environmental niche, causing multicollinearity among predictor species and environmental 147 covariables, ultimately blurring the determination of potential occurrences for the focal species (Zurell et al., 2018). Additionally, the suitable environmental range for the focal species might extend beyond 148 149 that of the predictor species, causing SDMs to misinterpret empty cells as unsuitable environmental 150 conditions rather than missing predictor species (Kissling et al., 2012). Finally, in communities with 151 numerous species, the issue of multiple testing and overfitting arises, which can be mitigated by 152 considering only the most abundant species as predictors (le Roux et al., 2014).

153

154 Another approach involves aggregating pairwise species interactions into **co-occurrence-based indices** 155 at the community level, serving as proxies for interactions in SDMs (Boulangeat et al., 2012). When 156 accounting for biotic interactions by including direct interaction covariates, knowledge of ecological 157 links between species pairs, such as for obligate parasite plant species (common in e.g., 158 Orobanchaceae), is crucial and should be supported by evidence. If the interacting species are unknown, 159 inference from data - ideally considering geographic and environmental variation - can be achieved 160 through (Bayesian) network analyses (Harris, 2016; Morueta-Holme et al., 2016; Pellissier et al., 2013; 161 Staniczenko et al., 2017). Insights derived from network analyses or prior ecological knowledge can 162 also help simplify the parameterization of biotic interactions within SDMs (Kissling et al., 2012).

163

164 Biotic modifiers (also referred to as ecosystem engineers, niche constructors, keystone species or 165 foundation species) significantly affect the growing conditions of the focal species, and consequently, 166 influence the presence of other interacting species reliant on the same resources and conditions (Linder 167 et al., 2012). Remarkable examples of biotic modifiers are earthworms enhancing litter decomposition 168 and soil turnover, trees possessing flammable biomass that can alter fire regimes (Bonanomi et al., 169 2008), nitrogen-fixing species that enhance nutrient conditions (Esther et al., 2008), and tree species 170 that have the capacity to modulate forest-floor microclimates (De Frenne et al., 2021; Haesen, Lenoir, 171 et al., 2023). Modeling the indirect interactions or mediating effects of these biotic modifiers on the 172 distribution of the focal species finds natural application in structural equation modeling (SEM) or path 173 analysis (Da Re et al., 2023). Nonetheless, within SDMs, environmental modulation can also be 174 captured through the inclusion of the abundances or predicted suitability of the biotic modifiers (Linder 175 et al., 2012) without necessarily involving a SEM framework. However, SEM or path analysis appear 176 more powerful in this regard because they enable testing of both the direct and indirect effects of 177 environmental factors on a focal species' occurrence. It does so by examining one or multiple species 178 acting as biotic modifiers, which may provide a more accurate reflection of the natural complexity of 179 the growing conditions (such as microclimatic conditions) experienced by the focal species.

181 2.2. Surrogate variables for plant interactions

182

In ecological systems, comprehensively deciphering the intricate web of interactions between multiple 183 184 species is a great challenge. To overcome this complexity, surrogate variables have been included in 185 SDMs that approximate gradients of biotic interactions within the landscape (Wisz et al., 2013). 186 These methods necessitate a foundation of *a priori* ecological knowledge concerning the nature of 187 relevant biotic interactions, such as competition for resources like light in the context of plant-plant 188 interactions. Researchers have utilized species richness derived from macroecological models (Rahbek 189 et al., 2007) or empirical models (Sabatini et al., 2022) as a surrogate indicator for the maximum 190 potential co-occurrence of species within landscape pixels. Moreover, proxies like productivity, tree 191 cover or vegetation height, often estimated through remote sensing, have been employed to gauge 192 competition intensity of the more visible layers of the plant community (Maestre et al., 2010; Nieto-193 Lugilde et al., 2015; Weber et al., 2016). Such surrogates, however, are not often available to describe 194 plant-plant interactions within the more hidden layers, such as for understorey vegetation in forest 195 ecosystems.

196 Community-weighted means of trait values also serve as valuable surrogate variables to infer various 197 ecological interactions within ecosystems (Morales-Castilla et al., 2015). Species traits could, for 198 instance, be used to suggest competitive exclusion among ecologically similar species (Davies et al., 199 2007). Biotic interactions are thus able to shape the functional composition of complete communities 200 (Gross et al., 2009). In instances where direct trait data might be unavailable, phylogenetic information 201 emerges as a useful proxy (Morales-Castilla et al., 2015), given that closely related species tend to 202 exhibit niche conservatism, thereby competing for analogous resources. Community phylogenetics 203 could thus be employed to gauge plant-plant interactions with more competition associated with 204 increasing phylogenetic relatedness (Valiente-Banuet & Verdú, 2008).

205

206 2.3. Joint species distribution models (jSDMs)

207

208 Multi-species or joint species distribution models (jSDMs) employ a hierarchical framework to 209 simultaneously fit multiple species-specific models, wherein the regression coefficients originate 210 from a common distribution (Ovaskainen & Soininen, 2011). This hierarchical structure enables the 211 accommodation of species with limited data by 'borrowing statistical information' from more abundant 212 species to estimate coefficients for rarer species (Hui et al., 2013; Ovaskainen & Soininen, 2011). 213 Species co-occurrence data also facilitate the inference of biotic interactions. Species that co-occur more 214 frequently than expected suggest facilitation, while occurrences less frequent than expected imply niche 215 partitioning or negative interactions like competition. Evidence of species interactions could thus be 216 found in the residuals of a jSDM after controlling for abiotic factors (Pollock et al., 2014). The inference of species associations from the residuals, rather than through predictor variables circumvents issues related to multicollinearity (Pollock et al., 2014), yet, overlooks the asymmetric nature of most interactions given the symmetrical parameterization of covariance matrices (Dormann et al., 2018).

220 Importantly, the mere co-occurrence of species should not unilaterally indicate ecological interactions 221 (Blanchet et al., 2020; Townsend Peterson et al., 2011). The residual species association patterns in 222 jSDMs may arise from various sources, including missing environmental covariates or unaccounted 223 biotic interactions (Dormann et al., 2018). As with direct interaction covariates, issues may arise with 224 indirect interactions and shared responses to unmeasured environmental variables, as these can affect 225 the residual correlation between species. Additionally, zero residual correlation between species should 226 not be conclusively interpreted as evidence of an absence of interaction, as this might stem from 227 insufficient data to estimate these interactions (Popovic et al., 2019). Furthermore, even if the residual 228 correlation matrix would reflect species interactions, it likely encompasses the realized impact of direct 229 negative (e.g., competition), direct positive (e.g., facilitation), and other more indirect (e.g., due to 230 shared predators) associations between species (Poggiato et al. 2021). Some of these pitfalls also apply 231 to other methods outlined above.

232

233 Joint SDMs delineate species distributions exclusively within the environmental space, while other 234 factors including interaction effects are included within the residual covariance matrix and thus might 235 not improve the assessment of the fundamental niche (Poggiato et al., 2021). The prevailing consensus 236 now is that residual correlation is not necessarily indicative for species' interaction (Dormann et al., 237 2018). The interpretation of the residual covariance matrix should consider the context of unaccounted 238 predictor variables, biotic interactions and model inaccuracies (Zurell et al., 2018). Joint SDMs may 239 thus serve as a useful tool for hypothesis generation - rather than definitive detectors of interactions -240 where substantial residual correlation between species warrants further investigation (Dormann et al., 241 2018; Pollock et al., 2014). Nevertheless, jSDMs have the proven ability to increase the predictive 242 accuracy of SDMs by incorporating the information inferred from species-to-species associations 243 (Norberg et al., 2019).

244

245 Early jSDMs relied on the multivariate probit model, inferring species-to-species associations 246 through a full-rank covariance matrix (Pollock et al., 2014). However, this approach faces limitations 247 in handling species-rich datasets as the number of parameters to estimate increases quadratically with 248 the number of species (Warton et al., 2015). To address this issue, most current methodologies substitute 249 the species covariance matrix with a reduced set of latent variables (Warton et al., 2015). The 250 reparameterization of large covariance matrices with latent variables enhances the computational 251 efficiency of jSDMs. Species responding similarly or differently to these latent variables exhibit 252 positive or negative associations (Warton et al., 2015). The factor loadings of the latent variables can 253 thus be translated into a species-species covariance matrix. The gain in computational efficiency stems from the considerably fewer latent factors (and thus, number of parameters to estimate) in comparison to the number of species in the data set (Ovaskainen et al., 2016). Further methodological developments in jSDMs have facilitated the incorporation of species' traits and phylogenies in the estimation of species niches (Abrego et al., 2017; Pollock et al., 2012) and enabled species associations to covary with measured environmental covariates (Tikhonov et al., 2017).

259

260 Joint SDMs, unlike models employing direct interaction covariates, do not explicitly model species 261 interactions and, as a result, do not necessitate a priori knowledge of these interactions. However, 262 Ohlmann et al. (2023) presented a statistical model that allows to integrate the topological structure of 263 a predefined metanetwork into the analysis of species distributions. In contrast to conventional jSDMs, 264 this model explicitly incorporates known interactions, offering a practical implementation of embedding 265 network ecology within jSDMs (Thuiller et al., 2023). This model condenses all local pairwise interactions into a unified metric, which can subsequently be mapped and correlated with spatial layers, 266 267 enabling the exploration of the effects of spatial environmental variation on biotic interactions 268 (Ohlmann et al., 2023).

269

270 2.4. Limitations of the existing SDM methods hindering the accurate integration of biotic interactions271

272 Several existing SDM techniques require assumptions that hinder the accurate integration of biotic 273 interaction effects. Most of these assumptions originate from insufficient understanding of the 274 mechanisms at play, which nonetheless can be deduced from experimental macroecological approaches. 275 For example, most SDM methods assume constant species-to-species associations in space. In natural 276 communities, however, the strength and direction of plant-plant interactions is likely to vary under 277 different environmental conditions (Morales-Castilla et al., 2015; Olsen et al., 2016). Similarly, several 278 methods also assume stationarity in interaction strengths across time, which is probably unlikely. 279 Interaction strengths within forest plant communities, for instance, have been shown to vary under 280 changing climate and forest density scenarios (Sanczuk et al., 2023). This concern has motivated the 281 modelling of temporal variation in species distribution via dynamic SDMs where environmental 282 linkages or residual spatial variation is allowed to vary over time (DeVISSER et al., 2010; Merow et 283 al., 2011), which is possible through an increased understanding of the operating processes (e.g., from 284 demography-environment relations). An experimentally-informed spatial prediction of the strength and 285 direction of plant-plant interactions is likely to advance our understanding on species-to-species 286 associations by integrating new field evidence, hereby leveraging the strengths of both experimental 287 research and correlative SDMs. Experimental macroecological approaches have already been 288 successful in disentangling plant-plant interactions along large environmental gradients (Maestre et al., 289 2005; Sanczuk et al., 2022; Shepard et al., 2021), and can become particularly valuable when combined

with manipulative treatments as they allow projections towards uncharted abiotic (De Frenne, 2023) and biotic conditions (Williams & Jackson, 2007).

- 292

293 3. Introduction of a novel concept: a plant-plant interaction index map

294

295 3.1. Rationale of the concept

296 Here we introduce the concept and potential application of a novel gridded map product: a plant-plant 297 interaction map depicting the realized strength of plant interactions experienced by a focal species or 298 functionally similar group of species. Fundamental to the plant-plant interaction map is the 299 quantification of the Plant-plant Interaction Index (from here referred to as PII), a numerical index 300 proportional to the impact of plant interactions on a focal species' performance, which can be assessed 301 in terms of differences in fitness, demographic rates, cover or presence/absence in experimental 302 vegetation plots with and without interactions with co-occurring plant species. The quantification of 303 plant-plant interaction effects based on performance differences is not new (Armas et al., 2004). The 304 PII of plant communities can be obtained preferably from manipulative experiments, but more 305 opportunistic data-driven approximations may be considered too (Fig. 1). Since the magnitude and 306 direction (e.g., positive under competition, negative under facilitation) of plant-plant interactions are 307 driven by specific plant functional traits of the co-occurring species (Freckleton & Watkinson, 2001; 308 Kunstler et al., 2012, 2016), we assume that the PII can be predicted for any given plant community 309 based on community weighted mean (CWM) values of a selection of functional traits (e.g., plant height, 310 specific leaf area, shoot and/or root biomass, rooting depth, etc.), trait syndromes (e.g., Grime C-scores 311 (Hunt et al., 2004) or colonization capacities (Verheyen et al., 2003)), or more general community 312 characteristics (e.g., total vegetation cover or leaf area index) (Salguero-Gómez et al., 2018). Following 313 ecological theory, CWM trait values or communities characteristic are driven by environmental 314 conditions at fine and coarse spatial grains (Govaert et al., 2023; Kemppinen et al., 2021; Moles et al., 315 2009, 2014; Sanczuk et al., 2021), such that community trait signatures, and the resulting PII for a focal 316 species can be estimated based on the environmental covariates to arrive at a gridded data layer, i.e., a 317 PII map.

318 The PII map provides a more process-based alternative to account for biotic interactions into SDM 319 analyses compared to - for instance - surrogate approaches, and is expected to become increasingly 320 relevant for the prediction of species' distributions under novel biotic and abiotic conditions (De Frenne, 321 2023; Williams & Jackson, 2007). In particular for novel environmental conditions that entail biotic 322 communities that have no present-day analog, and in which the formerly derived correlations between 323 plant interaction effects and the assumed environmental surrogates potentially no longer hold (Kerr et 324 al., 2023).



326 Figure 1 | Concept summary and flowchart of data to calculate a Plant-plant Interaction Index 327 (PII) map. Input data: based on experimental data from interaction-exclusion experiments replicated in 328 different environments (with the removal of all other species; or the removal/absence of only one 329 interacting species if a focal pairwise interaction is considered), the PII is quantified as the natural log-330 transformed (ln) response ratio of performance differences (performance in control versus performance 331 in exclusion treatment). The PII represents a numerical index proportional to the impact of plant-plant 332 interactions on a focal species' performance. Vegetation plot data and gridded environmental covariates are required for trait mapping. Modeling: Statistical models relate the PII derived from interaction-333 334 exclusion experiments to the Community-Weighted Mean (CWM) trait values of the co-occurring 335 species and the environment in the experimental plots. Note that more general community 336 characteristics (e.g., total cover or leaf area index (LAI)) may be considered too. Predictive models such as random forest or boosted regression trees are used to develop CWM trait maps based on the trait-337 338 environment relations based on the vegetation plot data set. Note that the CWM trait model and PII 339 model relies on independent environmental data. Projecting statistical model: The coefficient estimates 340 extracted from the statistical model and the gridded CWM trait- and environmental data allow to project 341 the PII across the study area. Map products: a spatially-explicit prediction of PII that estimates the 342 realized strength of plant-plant interactions experienced by a focal species. In case interaction-exclusion 343 experiments are impossible, the PII map may be approximated by one or multiple CWM trait maps of 344 the vegetation for a selection of traits assumed to determine interactions strength within the focal 345 stratum of the community.

346 3.2. Steps to develop the plant-plant interaction map

347 The development of the PII map based on environmental data and the trait signatures of plant 348 communities requires a four-step process: (i) quantification of the PII from experimental data based on 349 differences in plant performance following variation in plant-plant interactions, (ii) parametrization of 350 a statistical plant interaction model by linking the PII the environmental data and CWM trait data for 351 traits determining plant interaction strengths within the plant community in the experimental sites; (iii) 352 parameterization of a predictive model using vegetation survey data to map CWM trait values across 353 the study area based on environmental data, and (iv) projecting the plant-plant interaction model across 354 the study area using gridded CWM trait and environmental data layers.

355 3.2.1. Quantifying the PII, a proxy for plant-plant interaction strength

The quantification of the PII based on differences in plant performance following variation in plant-356 357 plant interactions ideally requires manipulative experiments (Box 1) (Armas et al., 2004). Spatially 358 replicated manipulative experiments (cf., experimental macroecology) are highly suited to unravel the 359 relations between community trait characteristics and the focal species' performance in great detail, and allows us to disentangle biotic effects from effects brought about by other environmental conditions 360 361 such as temperature, precipitation or soil characteristics (Dunne et al., 2004; Nemer et al., 2021; Shepard 362 et al., 2021). In these experiments, the focal species is transplanted or sown into paired plots with and 363 without co-occurring plant species (by e.g., clipping all above-ground plant biomass; e.g., Sanczuk et 364 al., 2022). It is not the identity of the interacting species itself, but rather the trait information of these 365 species that is used for predicting the PII. The PII may also depict plant-plant interactions between only 366 one focal species pair, in which case the PII is predicted based on key characteristics of the focal 367 interacting species while also accounting for interaction effects with other resident species (such as 368 demonstrated in the worked example, Box 2). Comparing the plant performances in the paired 369 experimental plots allows us to calculate the PII as the natural-log transformed response ratio between 370 both treatments (Hedges et al., 1999) (see also Box 1), which can be statistically linked to the trait-371 signature of the interacting plant community (or one single interacting species) and the environmental 372 covariates measured at the experimental sites. Using the response ratio (i.e., the PII) calculated from 373 paired plots as a dependent variable allows us to isolate the effects brought about by the unique 374 contribution of individual traits of the co-occurring species from other environmental conditions on the 375 focal species' performance. Especially when spatially replicated into different environmental conditions 376 along multi-scale gradients, interaction-exclusion transplant experiments become highly valuable to 377 gain detailed understanding on the relative importance of biotic interactions versus other environmental 378 conditions (e.g., macro- and microclimatic conditions, soil moisture or nutrient contents) (Olsen et al., 379 2016; Sanczuk et al., 2023; Töpper et al., 2018). Plant-herbivore interactions may also be modelled 380 using this framework, for instance, by calculating the PII metric from paired herbivore-exclusion

treatments applied on plant communities (cf., experimental designs like the ECOSHRUB research network; <u>https://www.ntnu.edu/biology/research/ecoshrub/</u>). Interaction-exclusion treatments are likely the most feasible for smaller or short-lived species. In case experiments are impossible, more opportunistic data-driven approaches may be considered (section 3.2.4).

385 The performance of the focal species can be assessed based on one or multiple performance attributes 386 at a single point in time (such as fitness, plant height, cover or fitness (De Pauw et al., 2022; Pérez-387 Harguindeguy et al., 2016)) or based on population-level demographic rates (e.g., vital rates such as the 388 survival and growth rate, or population growth rate) inferred from time series data (Fernández-389 Fernández et al., 2022; Merow et al., 2014) (Box 1). These approaches are technically challenging, but 390 may greatly benefit from recent developments in near-surface remote sensing. For example, plant trait-391 and performance-measurements, such as species-specific abundances, plant height or volume estimates, 392 can be obtained from photogrammetry, which is increasingly used in experimental ecology (Iglhaut et 393 al., 2019; Luscier et al., 2006). Photogrammetry overcomes common limitations inherent to field 394 observations (e.g., lack of consistency and objectivity) and can operate autonomously and continuously 395 at remote locations, which facilitates studying plant interactions through time.

Manipulative transplant experiments are ubiquitous for herbaceous species due to their short generation times and prompt responses to environmental variation, but become often impractical to run for many species together and even impossible to conduct for certain taxa. For tree species that have long generation times, the *in-situ* quantification of PIIs is generally impossible based on experiments, and alternative surrogate approaches (section 2.2) may be preferred (cf., Morales-Castilla et al., 2015).

401 3.2.2. Parameterizing a statistical Plant-plant Interaction Index model

The PII values of a focal species inferred from spatially replicated interaction-exclusion experiments can be related to the trait-signature of the co-occurring species and experimental environmental covariate data using multivariate regression models (with CWM traits and environmental covariates as predictors; hereafter referred to as PII models). The PII can also be considered for multiple focal species simultaneously, in which case also the trait values of (or, trait dissimilarity between) these focal species can be included in the PII model. PII models provide the information to project (map) the PII across the study area based on gridded environmental and CWM trait data (described in *section 3.2.4*).

The plant characteristics of the co-occurring species can be described in terms of CWM functional traits, but also other summary statistics of the traits' distributions (e.g., maximum) or community characteristics such as community below- or aboveground biomass, total cover or the leaf area index (LAI) may be considered. Differences in trait values between the focal species and the interacting vegetation (rather than absolute trait values of the interacting vegetation) can also be relevant. The 414 selection of relevant traits and/or community characteristics as predictor variables in the PII model will 415 depend on the nature of the biotic interactions under investigation. If, for example, competition for light 416 is considered the dominant type of interaction determining the performance of the focal species within 417 the community, community characteristics that relate to shading (e.g., CWM plant height in 418 combination with community LAI) should be focused upon for modelling. If belowground competition 419 is dominant, other community characteristics including, for example, belowground plant biomass and 420 CWM root traits (e.g., specific root length, rooting depth) will be more relevant. If no prior information 421 on the types of plant-plant interaction is available, one can consider more general community 422 characteristics that relate to plant competition in general, such as the CWM colonization capacity index 423 (Verheven et al., 2003) or CWM Grime-C (competition) score (Grime, 1977).

PII model development will depend on the data types considered: in experiments, trait data of the co-424 425 occurring species in the community will mostly be available from *in situ* measurements. Direct plant 426 trait measurements are especially relevant when (i) traits are very plastic and adapt to the environment 427 and (ii) in case the required trait data are not available in existing databases. In case the *in situ* trait 428 measurements are not available, species-specific average trait values can be extracted from globally-429 available trait databases such as LEDA (Kleyer et al., 2008) or TRY (Kattge et al., 2020). Based on collected trait data and focal species' performance data, a PII model can be fitted using a variety of 430 431 candidate models, going from simple linear regressions to more complex non-linear models such as 432 additive models or polynomial regression models. Models that explicitly test for interaction effects 433 between the CWM trait and environmental conditions, are more likely to describe the natural 434 complexity of the system (Shepard et al., 2021; Töpper et al., 2018).

435 Box 1 | Optimal experimental design to quantify the Plant-plant interactions index in the field

436 A well-chosen design of spatial interaction-exclusion experiments is key to develop a PII-map with 437 spatial detail (i.e., grain size) and extent relevant for biogeographical predictions. Interaction-exclusion 438 experiments (**Fig. 2**) become particularly valuable when replicated at fine spatial grains (for example, 439 100 m or smaller if micro-gradients are deemed to be important) and across several locations covering 440 a large spatial extent (for example, >1,000 km) in order to understand, and predict, the effects along 441 multi-scale environmental gradients (see **Box 2** for an example).



Figure 2 | Example of an interaction-exclusion experiment replicated in an understorey community of two different forest sites in Belgium. Control plots are plots where the focal species (*Ornithogalum pyrenaicum*) has been sown within the resident vegetation. Interaction-exclusion treatments were performed by clipping and removing all above-ground biomass. Wooden sticks indicated germinated seedlings. See (Sanczuk et al., 2022) for details.

448 Several performance attributes can be considered to quantify PIIs. Single sample estimates of 449 performances (such as derived from plant height, volume estimates or an integrated metric across 450 multiple traits (Verheyen & Hermy, 2004)) are evidently easier. However, such performance metrics 451 are more prone to bias introduced by stochastic environmental processes and individual outliers. 452 Population-level performance estimates inferred from annual transition data such as survival, fecundity 453 or population growth rates appear more robust and better accommodate environmental stochasticity 454 across the entire study period (Sanczuk et al., 2022).

A focal species likely experiences different types of interactions, including ones that can both negatively 455 456 and positively impact its performance. For example, certain trait signatures of the co-occurring species might have facilitative effects brought about by e.g., the protection against drought or herbivory by high 457 458 above-ground volume (Fig. 3a) (Rothstein & Zak, 2001), while simultaneously impacting the focal 459 species negatively due to, for example root competition (Fig. 3b) or high leaf area index (Fig. 3c). 460 Although the PII estimate is determined based on the overall realized impact of all different aspects of the co-occurring vegetation, statistical modeling allows us to disentangle individual PII – CWM trait 461 462 relations such that the overall effect of each investigated CWM trait of the co-occurring species can be 463 estimated.





Figure 3 | **Schematic illustration of hypothetical plant interaction models and formula of the Plant-plant Interaction Index (PII).** Illustration of a hypothetical relation between the PII and community weighted mean (CWM) trait values (e.g., CWM plant volume (a)), CWM plant syndromes (e.g., CWM Grime-C (competition) score (b)) or general community characteristic (e.g., Leaf Area Index (c)) under positive (e.g., due to facilitation) and negative (e.g., due to competition) plant-plant interactions. A PII of zero indicates that plant-plant interaction had no effects on the focal species performance.

473 3.2.3. Predicting and mapping CWM traits based on environmental covariates

474 After modelling the PII based on the trait signature of plant communities and the environment in the 475 experimental sites, the PII can be calculated for virtually any georeferenced assemblage of plant species 476 that have been described in terms of species composition, and for which environmental data and trait 477 characteristics for all co-occurring species are measured or can be extracted from databases. In order to 478 develop a PII map, gridded data layers of each predictor in the PII model (section 3.2.2) are required. 479 While spatial environmental data layers are available with an increasing level of detail (both in terms 480 of quality and grain size), CWM trait maps generally don't exist. Most recent developments in 481 predictive modelling, however, enable mapping of the CWM trait values as a function of environmental 482 covariates with increasing accuracy. Machine learning models such as random forest models or boosted 483 regression trees can be trained based on vegetation survey databases (e.g., following the protocol of 484 (Haesen et al., 2021)) and relate CWM trait values to environmental covariates of each plot in the database (extracted from gridded environmental data layers (Haesen et al., 2021; Hansen et al., 2010; 485 486 Karger et al., 2017), soilGrids.org or derived from more advanced high-resolution remote sensing 487 techniques (Ewald et al., 2018)). Using this model, CWM trait values for the set of traits considered in 488 the PII model can be mapped for each grid cell in the study area at the relevant grain size – resulting in 489 maps of CWM trait values. CWM trait model prediction can be validated independently based on 490 experimental site data (section 3.2.1).

491 Networks of vegetation survey plots are possibly the most important source of community-level plant 492 data, as they have been performed for various layers within the communities and generally contain data 493 on at least species identity and (relative) abundance. At the continental to global extent, numerous 494 vegetation databases (e.g., sPlotOpen (Sabatini et al., 2021), forestREplot survey 495 (https://forestreplot.ugent.be/), GLORIA (https://www.gloria.ac.at/home), EVA (Chytrý et al., 2016) or 496 AVA (https://www.geobotany.uaf.edu/ava/)) are available that contain exactly this type of information, 497 often for an enormous amount of vegetation plots spanning large environmental gradients at coarse (i.e., 498 along macroclimate gradients of temperature or precipitation) and finer spatial grains (e.g., along 499 gradients of microclimate or tree cover density).

500 Trait mapping is facilitated by the growing level of detail and availability of spatial environmental data 501 to describe the growing conditions of plant species. Latest developments in remote sensing, in 502 particular, increasingly enable the direct inference of trait values at the community level (CWM traits 503 data), or indirectly through describing the biotic growing conditions of the focal plant community, 504 because many remote sensing technologies are now able to capture different aspects of key plant 505 community traits (Chen et al., 2022; Cho et al., 2012). Prominent examples of such traits include plant 506 height, volume, leaf area, canopy cover and biomass, some of which have already been mapped in great 507 detail at continental and global scales by linking satellite imagery to 3-D Light Detection and Ranging 508 (LiDAR) data (Lang et al., 2023; Liu et al., 2023). LiDAR data is especially useful to map 3-D 509 morphological traits that are indicative of interaction strengths, e.g., forest understorey density estimates 510 or vertical profiles of leaf area and light availability (Kükenbrink et al., 2021). These attributes can be 511 complemented with recently developed layers describing leaf physiological and functional trait 512 information derived from hyperspectral imaging, such as photosynthetic pigments, nutrient content and 513 defensive compounds (Asner et al., 2015; Ewald et al., 2018; Wang et al., 2020). Furthermore, remote 514 sensing tools are also providing new insights into below-ground interactions, e.g., by using magnetic 515 resonance imaging (MRI) or ground penetrating radar, however these close-range techniques are yet to 516 be upscaled (Chen et al., 2022).

517 Biotic covariates can also be combined with abiotic covariates. For example, advances in microclimate 518 modelling and mapping improve the estimation of species climatic niches and (re-)distributions and are 519 thus expected to improve PII- and CWM trait-mapping (Haesen, Lenoir, et al., 2023; Maclean & Early, 520 2023). Microclimate maps are increasingly available at national, continental and even global scales 521 (Haesen, Lembrechts, et al., 2023; Lembrechts et al., 2022; Zellweger et al., 2023) and can be 522 complemented with coarser grained macroclimate data, as well as datasets describing soil properties 523 (Hengl et al., 2017; Karger et al., 2017). Mapping the PII and CWM trait values is thus increasingly 524 possible across a range of scales.

525 3.2.4. Calculating the PII-map

526 Combining the knowledge of the PII model (regression coefficients) and the spatial derivates of each 527 predictor variable in the PII model allows us to calculate an experimentally informed PII map. The 528 resulting spatial extent and grain size of the PII map inherently depends on the spatial design of the 529 interaction-exclusion experiment (section 3.2.1). Note that, although fine-grained large-extent map 530 predictions based on plant responses measured in experimental macroecological settings have already 531 been performed with success (Merow et al., 2017; Sanczuk et al., 2023), the accuracy of the PII- and 532 CWM trait-predictions far beyond the (biotic and abiotic) environmental range that is sampled can be 533 less accurate due to extrapolation uncertainty. In particular for novel environmental conditions that 534 entail biotic communities that have no present-day analog, the accuracy of PII model predictions 535 remains to be validated since formerly derived PIIs – trait- and environmental-relations potentially no 536 longer hold.

537 In the case experiments are impossible for the focal species, one may explore the use of CWM trait 538 maps directly as a covariate layer into SDM (cf. section 2.2; surrogate variables of interactions). 539 Although CWM trait maps don't capture direct information (from field evidence) on the link between 540 the focal species' performances and the trait signature of the resident vegetation, integrating these maps 541 for certain traits selected based on prior knowledge may still improve the accuracy of SDM predictions

542 (Morales-Castilla et al., 2015).

543 Box 2 | Worked example: PII-mapping using plant community data from a transplant experiment

544 To illustrate a potential workflow, we here develop a PII-map for the common European forest herb 545 Geum urbanum based on the realized impacts of the competitive generalist herb Urtica dioica. Urtica 546 is a strongly competitive herb species that can produce large volumes of biomass on nutrient-rich soils 547 and when light is not limited, such as in forest edges. We focus on the individual effects brought about 548 by Urtica on the performance of Geum while also accounting for potential interaction effects with other 549 resident herb species. To do so, we estimate the PII based on data from a fine-grained large-extent plant 550 community transplant experiment (Sanczuk et al., 2023). We then link the PII estimates to the 551 environment of the experiment and the proportional cover of the competitive generalist Urtica (here, as 552 a single plant community trait).

Step 1 - Quantifying the PII, a proxy for plant-plant interaction strength: The PII is quantified as the log-ratio of *Geum*'s biomass volume (calculated as cover × average plant height in the plot) in paired plots (i.e., in the same environment next to each other) with and without *Urtica*, using data from a spatially replicated community transplant experiment (see Sanczuk et al., 2023 and Fig. 4)

Step 2 - Parameterizing a statistical Plant-plant Interaction Index model: The PII is linked to the
percentage cover of *Urtica* and the environmental variation at coarse (growing-season temperature and
precipitation) and fine spatial grains (tree cover density) within the experiment using a linear model. A

model selection based on AICc (Akaike information criterion corrected for small sample size) is applied
to identify the most parsimonious model structure and avoid overfitting.

562 Step 3 - Predicting and mapping CWM traits based on environmental covariates: We predict the 563 percentage cover of Urtica across the study area based on an independent vegetation plot data set 564 (sPlotOpen; Sabatini et al., 2021), and gridded environmental covariates. The percentage cover of 565 Urtica across the vegetation plots is linked to three gridded environmental predictor variables (for 566 illustration we used the same variables as in step 2, but these don't necessarily have to be the same), 567 using the predictive Random Forest algorithm. The trained model was projected across the study area 568 to obtain the 'community trait map'.

Step 4 - Calculating the PII-map: Finally, a PII measure is estimated for each grid cell in the study area
using the optimal model structure and coefficient estimates of the linear model (data from step 2) and
the gridded environmental (temperature, precipitation and tree cover density) and trait variable
(predicted cover of *Urtica*, data from step 3).

573 The PII map product may be integrated in SDMs to account for biotic interaction effects brought about 574 by Urtica on the habitat suitability of Geum. This PII map will be especially relevant to obtain high-575 resolution predictions of the habitat suitability in habitat types where Urtica is expected to be a determinant driver of the presence of Geum – such as in forest edges. Knowing the link between Geum 576 577 performance and the percentage cover of Urtica, we can, for instance, assess the possible impacts of 578 changes in tree cover density on the percentage cover of *Urtica* and how this affects the PII, to ultimately 579 analyze to what extent these biotic changes propagate up to biogeographical range shifts of *Geum*. This 580 way, PII mapping is a powerful and flexible tool that effectively leverages the strengths of both 581 experimental research and correlative SDMs.



582

Figure 4 | Methods summary and data to calculate a Plant-plant Interaction Index (PII) map used 583 in the worked example. We mapped the PII for the forest herb *Geum urbanum* based on the realized 584 impact of the competitive generalist herb Urtica dioica. We thus focus on the individual effects brought 585 about by Urtica on the performance of Geum while accounting for the effects of ambient plant 586 587 interaction with other resident herb species. We estimate the PII based on data from a fine-grained largeextent plant community transplant experiment (right grey panel, a-c) (Sanczuk et al., 2023). The PII is 588 589 quantified as the log-ratio of *Geum*'s biomass in plant communities with and without *Urtica* (right grey 590 panel, d-e). The PII is linked to the percentage cover of Urtica and the environmental variation at coarse 591 (temperature and precipitation) and fine spatial grains (tree cover density) within the experiment using a linear model. A model selection on the full model is applied based on AICc (Akaike information 592 593 criterion corrected for small sample size). We map the proportional cover of the Urtica across the study 594 area based on an independent vegetation plot data set (sPlotOpen), and gridded environmental 595 covariates (left grev panel) using a predictive Random Forest algorithm, to obtain a 'trait map' (i.e., predicted cover (%) of Urtica). Trait map predictions are made at 25 m resolution for the extent of 596 597 temperate Europe. Red squares indicate the raster prediction of landscape windows illustrated at the 598 right. The PII map product is estimated for 1,000,000 random forested locations in the study area and 599 of each forested grid cell within the three landscape windows by projecting the linear model across the (abiotic and biotic) environment. Note that, as expected, forest edges and regions with warmer 600 macroclimate generally have a higher estimated PII. 601

602 3.5. Challenges for plant-plant interaction mapping

603 Plant-plant interaction mapping based on the link between species' performances and the trait-landscape 604 of the co-occurring plant species poses several challenges. First, the accuracy of the map predictions 605 inherently depends on the predictive performance of the PII model. It remains to be evaluated based on 606 real-world data to what extent PII models allow us to accurately predict plant-plant interactions 607 strengths, and which experimental designs, plant community and remote sensing data sources are most 608 promising. Second, model predictions beyond the environmental range that was sampled might be less 609 accurate as a result of extrapolation uncertainty. Third, unaccounted biotic interactions such as plant-610 animal dynamics may substantially impact plant communities and these effects do not necessarily 611 remain constant along an environmental gradient (see e.g., variable effects of herbivory along a nitrogen 612 gradient (Segar et al., 2022)). Last, a focal species likely experiences different modes of interactions, 613 with likely divergent effects on the species' performances (e.g., both facilitative and competitive 614 interactions; Fig. 3). The PII is not able to depict these different aspects of the biotic environment, and 615 its final estimate will be determined based on the overall, realized impact of the resident vegetation.

616 **4. Conclusions and outlook**

617 Changing environmental conditions lead to novel assemblages of species that may start interacting 618 within an uncharted portion of the environmental space. Accounting for plant-plant interactions using 619 existing SDM techniques remains challenging. Advanced SDMs facilitate the integration of biotic 620 interaction effects based on species-to-species associations. However, when novel environmental 621 conditions arise in which the formerly derived species' correlations potentially no longer hold, a more 622 process-based approach is warranted.

623 By linking species performances to the trait characteristics of interacting species, biotic interactions 624 maps can be obtained for virtually any focal natural community, including maps for less visible 625 vegetation layers as well as for plant-herbivore interactions or interactions within animal communities. 626 Upscaling manipulative experiments at macroecological settings to generate broad-scale plant-plant 627 interaction maps hold promise for an improved integration of biotic interaction effects by leveraging 628 the strengths of experimental research and correlative SDMs. In terms of perspectives, PII mapping 629 may also be deployed to aid the prediction of impacts of alterations in community trait values on the 630 focal species' performance, even if certain combinations of species do not yet co-occur. For example, 631 knowing the link between a focal species performance and the trait signature of the co-occurring 632 species, one can assess the possible impacts of a successful invasive species (with known trait values) 633 on the community PII, and analyzes to what extent these biotic alterations propagate up to 634 biogeographical range shifts of the focal species. Hence, PII mapping adds an additional layer of flexibility to the ecologists' toolbox. 635

636 Data Availability Statement

637 All data and code of used in the worked example will be made available on FigShare638 (<u>https://figshare.com/</u>).

639 **References**

- Abrego, N., Norberg, A., & Ovaskainen, O. (2017). Measuring and predicting the influence of traits
 on the assembly processes of wood-inhabiting fungi. *Journal of Ecology*, *105*(4), 1070–1081.
- Alexander, J. M., Diez, J. M., Hart, S. P., & Levine, J. M. (2016). When Climate Reshuffles
 Competitors: A Call for Experimental Macroecology. *Trends in Ecology & Evolution*, *31*(11),
 831–841.
- Alexander, J. M., Diez, J. M., & Levine, J. M. (2015a). Novel competitors shape species' responses to
 climate change. *Nature*, 525(7570), 515–518.
- Alexander, J. M., Diez, J. M., & Levine, J. M. (2015b). Novel competitors shape species' responses to
 climate change. *Nature*, 525(7570), 515–518.
- Araújo, M. B., & Guisan, A. (2006). Five (or so) challenges for species distribution modelling.
 Journal of Biogeography, 33(10), 1677–1688.
- Armas, C., Ordiales, R., & Pugnaire, F. I. (2004). MEASURING PLANT INTERACTIONS: A NEW
 COMPARATIVE INDEX. *Ecology*, 85(10), 2682–2686.
- Asner, G. P., Martin, R. E., Anderson, C. B., & Knapp, D. E. (2015). Quantifying forest canopy traits:
 Imaging spectroscopy versus field survey. *Remote Sensing of the Environment*, *158*, 15–27.
- Blanchet, F. G., Cazelles, K., & Gravel, D. (2020). Co-occurrence is not evidence of ecological
 interactions. *Ecology Letters*, 23(7), 1050–1063.
- Bonanomi, G., Rietkerk, M., Dekker, S. C., & Mazzoleni, S. (2008). Islands of fertility induce cooccurring negative and positive plant-soil feedbacks promoting coexistence. *Plant Ecology*,
 197(2), 207–218.
- Boulangeat, I., Gravel, D., & Thuiller, W. (2012). Accounting for dispersal and biotic interactions to
 disentangle the drivers of species distributions and their abundances. *Ecology Letters*, 15(6),
 584–593.
- Chen, B. J. W., Teng, S. N., Zheng, G., Cui, L., Li, S.-P., Staal, A., Eitel, J. U. H., Crowther, T. W.,
 Berdugo, M., Mo, L., Ma, H., Bialic-Murphy, L., Zohner, C. M., Maynard, D. S., Averill, C.,
 Zhang, J., He, Q., Evers, J. B., Anten, N. P. R., ... Xu, C. (2022). Inferring plant–plant
 interactions using remote sensing. *Journal of Ecology*, *110*(10), 2268–2287.
- Cho, M. A., Mathieu, R., Asner, G. P., Naidoo, L., van Aardt, J., Ramoelo, A., Debba, P., Wessels,
 K., Main, R., Smit, I. P. J., & Erasmus, B. (2012). Mapping tree species composition in South
 African savannas using an integrated airborne spectral and LiDAR system. *Remote Sensing of Environment*, 125, 214–226.
- Chytrý, M., Hennekens, S. M., Jiménez-Alfaro, B., Knollová, I., Dengler, J., Jansen, F., Landucci, F.,
 Schaminée, J. H. J., Aćić, S., Agrillo, E., Ambarlı, D., Angelini, P., Apostolova, I., Attorre, F.,
 Berg, C., Bergmeier, E., Biurrun, I., Botta-Dukát, Z., Brisse, H., ... Yamalov, S. (2016).
 European Vegetation Archive (EVA): an integrated database of European vegetation plots. *Applied Vegetation Science*, 19(1), 173–180.
- Da Re, D., Tordoni, E., Lenoir, J., Rubin, S., & Vanwambeke, S. (2023). Towards causal relationships
 for modelling species distribution. In *EcoEvoRxiv*. https://doi.org/10.32942/x2188q
- Davies, T. J., Meiri, S., Barraclough, T. G., & Gittleman, J. L. (2007). Species co-existence and
 character divergence across carnivores. *Ecology Letters*, 10(2), 146–152.
- De Frenne, P. (2023). Novel light regimes in European forests. *Nature Ecology & Evolution*.
 https://doi.org/10.1038/s41559-023-02242-2
- De Frenne, P., Lenoir, J., Luoto, M., Scheffers, B. R., Zellweger, F., Aalto, J., Ashcroft, M. B.,
 Christiansen, D. M., Decocq, G., De Pauw, K., Govaert, S., Greiser, C., Gril, E., Hampe, A.,
 Jucker, T., Klinges, D. H., Koelemeijer, I. A., Lembrechts, J. J., Marrec, R., ... Hylander, K.
 (2021). Forest microclimates and climate change: Importance, drivers and future research
 agenda. *Global Change Biology*, *27*(11), 2279–2297.
- be Pauw, K., Sanczuk, P., Meeussen, C., Depauw, L., De Lombaerde, E., Govaert, S., Vanneste, T.,
 Brunet, J., Cousins, S. A. O., Gasperini, C., Hedwall, P.-O., Iacopetti, G., Lenoir, J., Plue, J.,
- 689 Selvi, F., Spicher, F., Uria-Diez, J., Verheyen, K., Vangansbeke, P., & De Frenne, P. (2022).
- Forest understorey communities respond strongly to light in interaction with forest structure, but
 not to microclimate warming. *The New Phytologist*, 233(1), 219–235.

- 692 DeVISSER, M. H., Messina, J. P., Moore, N. J., Lusch, D. P., & Maitima, J. (2010). A dynamic
 693 species distribution model of Glossina subgenus Morsitans: The identification of tsetse reservoirs
 694 and refugia. *Ecosphere*, 1(1), art6.
- Dormann, C. F., Bobrowski, M., Dehling, D. M., Harris, D. J., Hartig, F., Lischke, H., Moretti, M. D.,
 Pagel, J., Pinkert, S., Schleuning, M., Schmidt, S. I., Sheppard, C. S., Steinbauer, M. J., Zeuss,
 D., & Kraan, C. (2018). Biotic interactions in species distribution modelling: 10 questions to
 guide interpretation and avoid false conclusions. *Global Ecology and Biogeography*, 27(9),
 1004–1016.
- Dunne, J. A., Saleska, S. R., Fischer, M. L., & Harte, J. (2004). Integrating experimental and gradient
 methods in ecological climate change research. *Ecology*, 85(4), 904–916.
- Esther, A., Groeneveld, J., Enright, N. J., Miller, B. P., Lamont, B. B., Perry, G. L. W., Schurr, F. M.,
 & Jeltsch, F. (2008). Assessing the importance of seed immigration on coexistence of plant
 functional types in a species-rich ecosystem. *Ecological Modelling*, *213*(3), 402–416.
- Ewald, M., Aerts, R., Lenoir, J., Fassnacht, F. E., Nicolas, M., Skowronek, S., Piat, J., Honnay, O.,
 Garzón-López, C. X., Feilhauer, H., Van De Kerchove, R., Somers, B., Hattab, T., Rocchini, D.,
 & Schmidtlein, S. (2018). LiDAR derived forest structure data improves predictions of canopy N
 and P concentrations from imaging spectroscopy. *Remote Sensing of Environment*, *211*, 13–25.
- Fernández-Fernández, P., Sanczuk, P., Vanneste, T., Brunet, J., Ehrlén, J., Hedwall, P.-O., Hylander,
 K., Van Den Berge, S., Verheyen, K., & De Frenne, P. (2022). Different effects of warming
 treatments in forests versus hedgerows on the understorey plant Geum urbanum. *Plant Biology*,
 24(5), 734–744.
- Freckleton, R. P., & Watkinson, A. R. (2001). Predicting competition coefficients for plant mixtures:
 reciprocity, transitivity and correlations with life-history traits. *Ecology Letters*, 4(4), 348–357.
- Govaert, S., Meeussen, C., Vanneste, T., Bollmann, K., Brunet, J., Calders, K., Cousins, S. A. O., De
 Pauw, K., Diekmann, M., Graae, B. J., Hedwall, P.-O., Iacopetti, G., Lenoir, J., Lindmo, S.,
 Orczewska, A., Ponette, Q., Plue, J., Sanczuk, P., Selvi, F., ... De Frenne, P. (2023). Trait–
 micro-environment relationships of forest herb communities across Europe. *Global Ecology and Biogeography*, 33(2), 286-302.
- Grime, J. P. (1977). Evidence for the Existence of Three Primary Strategies in Plants and Its
 Relevance to Ecological and Evolutionary Theory. *The American Naturalist*.
 https://doi.org/10.1086/283244
- Gross, N., Kunstler, G., Liancourt, P., De Bello, F., Suding, K. N., & Lavorel, S. (2009). Linking
 individual response to biotic interactions with community structure: a trait-based framework.
 Functional Ecology, 23(6), 1167–1178.
- Guisan, A., & Thuiller, W. (2005). Predicting species distribution: offering more than simple habitat
 models. *Ecology Letters*, 8(9), 993–1009.
- Haesen, S., Lembrechts, J. J., De Frenne, P., Lenoir, J., Aalto, J., Ashcroft, M. B., Kopecký, M.,
 Luoto, M., Maclean, I., Nijs, I., Niittynen, P., van den Hoogen, J., Arriga, N., Brůna, J.,
 Buchmann, N., Čiliak, M., Collalti, A., De Lombaerde, E., Descombes, P., ... Van Meerbeek, K.
- 731 (2021). ForestTemp Sub-canopy microclimate temperatures of European forests. *Global* 732 *Change Biology*, 27(23), 6307–6319.
- Haesen, S., Lembrechts, J. J., De Frenne, P., Lenoir, J., Aalto, J., Ashcroft, M. B., Kopecký, M.,
 Luoto, M., Maclean, I., Nijs, I., Niittynen, P., van den Hoogen, J., Arriga, N., Brůna, J.,
 Buchmann, N., Čiliak, M., Collalti, A., De Lombaerde, E., Descombes, P., ... Van Meerbeek, K.
 (2023). ForestClim-Bioclimatic variables for microclimate temperatures of European forests. *Global Change Biology*, 29(11), 2886–2892.
- Haesen, S., Lenoir, J., Gril, E., De Frenne, P., Lembrechts, J. J., Kopecký, M., Macek, M., Man, M.,
 Wild, J., & Van Meerbeek, K. (2023). Microclimate reveals the true thermal niche of forest plant
 species. *Ecology Letters*, 26(12), 2043-2055.
- Hansen, M. C., Stehman, S. V., & Potapov, P. V. (2010). Quantification of global gross forest cover
 loss. *Proceedings of the National Academy of Sciences of the United States of America*, 107(19),
 8650–8655.
- Harris, D. J. (2016). Inferring species interactions from co-occurrence data with Markov networks.
 Ecology, 97(12), 3308–3314.
- Hedges, L. V., Gurevitch, J., & Curtis, P. S. (1999). The meta-analysis of response ratios in

- 747 experimental ecology. *Ecology*, *80*(4), 1150–1156.
- Hengl, T., Mendes de Jesus, J., Heuvelink, G. B. M., Ruiperez Gonzalez, M., Kilibarda, M., Blagotić,
 A., Shangguan, W., Wright, M. N., Geng, X., Bauer-Marschallinger, B., Guevara, M. A., Vargas,
 R., MacMillan, R. A., Batjes, N. H., Leenaars, J. G. B., Ribeiro, E., Wheeler, I., Mantel, S., &
 Kempen, B. (2017). SoilGrids250m: Global gridded soil information based on machine learning. *PloS One*, *12*(2), e0169748.
- HilleRisLambers, J., Harsch, M. A., Ettinger, A. K., Ford, K. R., & Theobald, E. J. (2013). How will
 biotic interactions influence climate change-induced range shifts? *Annals of the New York Academy of Sciences*, 1297, 112–125.
- Hui, F. K. C., Warton, D. I., Foster, S. D., & Dunstan, P. K. (2013). To mix or not to mix: comparing
 the predictive performance of mixture models vs. separate species distribution models. *Ecology*,
 94(9), 1913–1919.
- Hunt, R., Hodgson, J. G., Thompson, K., Bungener, P., Dunnett, N. P., & Askew, A. P. (2004). A new
 practical tool for deriving a functional signature for herbaceous vegetation. *Applied Vegetation Science*, 7(2), 163–170.
- Iglhaut, J., Cabo, C., Puliti, S., Piermattei, L., O'Connor, J., & Rosette, J. (2019). Structure from
 motion photogrammetry in forestry: A review. *Current Forestry Reports*, 5(3), 155–168.
- Karger, D. N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R. W., Zimmermann, N. E.,
 Linder, H. P., & Kessler, M. (2017). Climatologies at high resolution for the earth's land surface
 areas. *Scientific Data*, *4*, 170122.
- Kattge, J., Bönisch, G., Díaz, S., Lavorel, S., Prentice, I. C., Leadley, P., Tautenhahn, S., Werner, G.
 D. A., Aakala, T., Abedi, M., Acosta, A. T. R., Adamidis, G. C., Adamson, K., Aiba, M., Albert,
 C. H., Alcántara, J. M., Alcázar C, C., Aleixo, I., Ali, H., ... Wirth, C. (2020). TRY plant trait
 database enhanced coverage and open access. *Global Change Biology*, 26(1), 119–188.
- Kemppinen, J., Niittynen, P., le Roux, P. C., Momberg, M., Happonen, K., Aalto, J., Rautakoski, H.,
 Enquist, B. J., Vandvik, V., Halbritter, A. H., Maitner, B., & Luoto, M. (2021). Consistent traitenvironment relationships within and across tundra plant communities. *Nature Ecology & Evolution*, 5(4), 458–467.
- Kerr, M. R., Ordonez, A., Riede, F., & Svenning, J.C. (2023). A biogeographic–macroecological
 perspective on the rising novelty of the biosphere in the Anthropocene. *Journal of Biogeography*,
 51(4), 575-587.
- Kissling, W. D., Dormann, C. F., Groeneveld, J., Hickler, T., Kühn, I., McInerny, G. J., Montoya, J.
 M., Römermann, C., Schiffers, K., Schurr, F. M., Singer, A., Svenning, J.-C., Zimmermann, N.
 E., & O'Hara, R. B. (2012). Towards novel approaches to modelling biotic interactions in
 multispecies assemblages at large spatial extents. *Journal of Biogeography*, *39*(12), 2163–2178.
- Kleyer, M., Bekker, R. M., Knevel, I. C., Bakker, J. P., Thompson, K., Sonnenschein, M., Poschlod,
 P., van Groenendael, J. M., Klimeš, L., Klimešová, J., Klotz, S., Rusch, G. M., Hermy, M.,
 Adriaens, D., Boedeltje, G., Bossuyt, B., Dannemann, A., Endels, P., Götzenberger, L., ... Peco,
 B. (2008). The LEDA Traitbase: a database of life-history traits of the Northwest European flora.
- 786 *Journal of Ecology*, *96*(6), 1266–1274.
- Kükenbrink, D., Schneider, F. D., Schmid, B., Gastellu-Etchegorry, J.-P., Schaepman, M. E., &
 Morsdorf, F. (2021). Modelling of three-dimensional, diurnal light extinction in two contrasting
 forests. *Agricultural and Forest Meteorology*, *296*(108230), 108230.
- Kunstler, G., Falster, D., Coomes, D. A., Hui, F., Kooyman, R. M., Laughlin, D. C., Poorter, L.,
 Vanderwel, M., Vieilledent, G., Wright, S. J., Aiba, M., Baraloto, C., Caspersen, J., Cornelissen,
 J. H. C., Gourlet-Fleury, S., Hanewinkel, M., Herault, B., Kattge, J., Kurokawa, H., ... Westoby,
 M. (2016). Plant functional traits have globally consistent effects on competition. *Nature*, *529*(7585), 204–207.
- Kunstler, G., Lavergne, S., Courbaud, B., Thuiller, W., Vieilledent, G., Zimmermann, N. E., Kattge,
 J., & Coomes, D. A. (2012). Competitive interactions between forest trees are driven by species'
 trait hierarchy, not phylogenetic or functional similarity: implications for forest community
 assembly. *Ecology Letters*, 15(8), 831–840.
- Lang, N., Jetz, W., Schindler, K., & Wegner, J. D. (2023). A high-resolution canopy height model of
 the Earth. *Nature Ecology & Evolution*, 7(11), 1778–1789.
- 801 Leathwick, J. R., & Austin, M. P. (2001). Competitive interactions between tree species in New

- 802 Zealand's old-growth indigenous forests. *Ecology*, 82(9), 2560–2573.
- Lembrechts, J. J., van den Hoogen, J., Aalto, J., Ashcroft, M. B., De Frenne, P., Kemppinen, J.,
 Kopecký, M., Luoto, M., Maclean, I. M. D., Crowther, T. W., Bailey, J. J., Haesen, S., Klinges,
 D. H., Niittynen, P., Scheffers, B. R., Van Meerbeek, K., Aartsma, P., Abdalaze, O., Abedi, M.,
 ... Lenoir, J. (2022). Global maps of soil temperature. *Global Change Biology*, 28(9), 3110–
 3144.
- Lenoir, J., Bertrand, R., Comte, L., Bourgeaud, L., Hattab, T., Murienne, J., & Grenouillet, G. (2020).
 Species better track climate warming in the oceans than on land. *Nature Ecology & Evolution*,
 4(8), 1044–1059.
- le Roux, P. C., Pellissier, L., Wisz, M. S., & Luoto, M. (2014). Incorporating dominant species as
 proxies for biotic interactions strengthens plant community models. *Journal of Ecology*, *102*(3),
 767–775.
- Linder, H. P., Bykova, O., Dyke, J., Etienne, R. S., Hickler, T., Kühn, I., Marion, G., Ohlemüller, R.,
 Schymanski, S. J., & Singer, A. (2012). Biotic modifiers, environmental modulation and species
 distribution models. *Journal of Biogeography*, *39*(12), 2179–2190.
- Liu, S., Brandt, M., Nord-Larsen, T., Chave, J., Reiner, F., Lang, N., Tong, X., Ciais, P., Igel, C.,
 Pascual, A., Guerra-Hernandez, J., Li, S., Mugabowindekwe, M., Saatchi, S., Yue, Y., Chen, Z.,
 & Fensholt, R. (2023). The overlooked contribution of trees outside forests to tree cover and
 woody biomass across Europe. *Science Advances*, 9(37), eadh4097.
- Luscier, J. D., Thompson, W. L., Wilson, J. M., Gorham, B. E., & Dragut, L. D. (2006). Using digital
 photographs and object-based image analysis to estimate percent ground cover in vegetation *Frontiers in Ecology and the Environment*, 4(8), 408–413.
- Lyu, S., & Alexander, J. M. (2022). Competition contributes to both warm and cool range edges.
 Nature Communications, *13*(1), 2502.
- Maclean, I. M. D., & Early, R. (2023). Macroclimate data overestimate range shifts of plants in
 response to climate change. *Nature Climate Change*, *13*(5), 484–490.
- Maestre, F. T., Bowker, M. A., Escolar, C., Puche, M. D., Soliveres, S., Maltez-Mouro, S., GarcíaPalacios, P., Castillo-Monroy, A. P., Martínez, I., & Escudero, A. (2010). Do biotic interactions
 modulate ecosystem functioning along stress gradients? Insights from semi-arid plant and
 biological soil crust communities. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 365(1549), 2057–2070.
- Maestre, F. T., Valladares, F., & Reynolds, J. F. (2005). Is the change of plant–plant interactions with
 abiotic stress predictable? A meta-analysis of field results in arid environments. *Journal of Ecology*, 93(4), 748–757.
- Meier, E. S., Kienast, F., Pearman, P. B., Svenning, J.-C., Thuiller, W., Araújo, M. B., Guisan, A., &
 Zimmermann, N. E. (2010). Biotic and abiotic variables show little redundancy in explaining tree
 species distributions. *Ecography*, *33*(6), 1038–1048.
- Merow, C., Bois, S. T., Allen, J. M., Xie, Y., & Silander, J. A., Jr. (2017). Climate change both
 facilitates and inhibits invasive plant ranges in New England. *Proceedings of the National Academy of Sciences of the United States of America*, 114(16), E3276–E3284.
- Merow, C., Lafleur, N., Silander, J. A., Jr, Wilson, A. M., & Rubega, M. (2011). Developing dynamic
 mechanistic species distribution models: predicting bird-mediated spread of invasive plants
 across northeastern North America. *The American Naturalist*, *178*(1), 30–43.
- Merow, C., Latimer, A. M., Wilson, A. M., McMahon, S. M., Rebelo, A. G., & Silander, J. A., Jr.
 (2014). On using integral projection models to generate demographically driven predictions of
 species' distributions: development and validation using sparse data. *Ecography*, *37*(12), 1167–
 1183.
- Merow, C., Smith, M. J., & Silander, J. A., Jr. (2013). A practical guide to MaxEnt for modeling
 species' distributions: what it does, and why inputs and settings matter. *Ecography*, *36*(10),
 1058–1069.
- Moles, A. T., Perkins, S. E., Laffan, S. W., Flores-Moreno, H., Awasthy, M., Tindall, M. L., Sack, L.,
 Pitman, A., Kattge, J., Aarssen, L. W., Anand, M., Bahn, M., Blonder, B., Cavender-Bares, J.,
 Cornelissen, J. H. C., Cornwell, W. K., Díaz, S., Dickie, J. B., Freschet, G. T., ... Bonser, S. P.
 (2014). Which is a better predictor of plant traits: temperature or precipitation? *Journal of*
- 856 *Vegetation Science*, 25(5), 1167–1180.

- Moles, A. T., Warton, D. I., Warman, L., Swenson, N. G., Laffan, S. W., Zanne, A. E., Pitman, A.,
 Hemmings, F. A., & Leishman, M. R. (2009). Global patterns in plant height. *Journal of Ecology*, 97(5), 923–932.
- Morales-Castilla, I., Matias, M. G., Gravel, D., & Araújo, M. B. (2015). Inferring biotic interactions
 from proxies. *Trends in Ecology & Evolution*, 30(6), 347–356.
- Morueta-Holme, N., Blonder, B., Sandel, B., McGill, B. J., Peet, R. K., Ott, J. E., Violle, C., Enquist,
 B. J., Jørgensen, P. M., & Svenning, J.-C. (2016). A network approach for inferring species
 associations from co-occurrence data. *Ecography*, *39*(12), 1139–1150.
- Morueta-Holme, N., Iversen, L. L., Corcoran, D., Rahbek, C., & Normand, S. (2023). Unlocking
 ground-based imagery for habitat mapping. *Trends in Ecology & Evolution*.
 https://doi.org/10.1016/i.tree.2023.11.005
- Nemer, D., Liancourt, P., Delerue, F., Randé, H., & Michalet, R. (2021). Species stress tolerance and
 community competitive effects drive differences in species composition between calcareous and
 siliceous plant communities. *Journal of Ecology*, *109*(12), 4132–4142.
- Nieto-Lugilde, D., Lenoir, J., Abdulhak, S., Aeschimann, D., Dullinger, S., Gégout, J.-C., Guisan, A.,
 Pauli, H., Renaud, J., Theurillat, J.-P., Thuiller, W., Van Es, J., Vittoz, P., Willner, W.,
 Wohlgemuth, T., Zimmermann, N. E., & Svenning, J.-C. (2015). Tree cover at fine and coarse
 spatial grains interacts with shade tolerance to shape plant species distributions across the Alps. *Ecography*, 38(6), 578–589.
- Norberg, A., Abrego, N., Blanchet, F. G., Adler, F. R., Anderson, B. J., Anttila, J., Araújo, M. B.,
 Dallas, T., Dunson, D., Elith, J., Foster, S. D., Fox, R., Franklin, J., Godsoe, W., Guisan, A.,
 O'Hara, B., Hill, N. A., Holt, R. D., Hui, F. K. C., ... Ovaskainen, O. (2019). A comprehensive
 evaluation of predictive performance of 33 species distribution models at species and community
 levels. *Ecological Monographs*, 89(3), e01370.
- Ohlmann, M., Matias, C., Poggiato, G., Dray, S., Thuiller, W., & Miele, V. (2023). Quantifying the
 overall effect of biotic interactions on species distributions along environmental gradients.
 Ecological Modelling, 483, 110424.
- Olsen, S. L., Töpper, J. P., Skarpaas, O., Vandvik, V., & Klanderud, K. (2016). From facilitation to
 competition: temperature-driven shift in dominant plant interactions affects population dynamics
 in seminatural grasslands. *Global Change Biology*, 22(5), 1915–1926.
- 887 Ovaskainen, O., & Soininen, J. (2011). Making more out of sparse data: hierarchical modeling of
 888 species communities. *Ecology*, 92(2), 289–295.
- Pellissier, L., Anne Bråthen, K., Pottier, J., Randin, C. F., Vittoz, P., Dubuis, A., Yoccoz, N. G., Alm,
 T., Zimmermann, N. E., & Guisan, A. (2010). Species distribution models reveal apparent
 competitive and facilitative effects of a dominant species on the distribution of tundra plants. *Ecography*, 33(6), 1004–1014.
- Pellissier, L., Rohr, R. P., Ndiribe, C., Pradervand, J.-N., Salamin, N., Guisan, A., & Wisz, M. (2013).
 Combining food web and species distribution models for improved community projections. *Ecology and Evolution*, *3*(13), 4572–4583.
- Pérez-Harguindeguy, N., Díaz, S., Garnier, E., Lavorel, S., Poorter, H., Jaureguiberry, P., Bret-Harte,
 M. S., Cornwell, W. K., Craine, J. M., Gurvich, D. E., Urcelay, C., Veneklaas, E. J., Reich, P. B.,
 Poorter, L., Wright, I. J., Ray, P., Enrico, L., Pausas, J. G., de Vos, A. C., ... Cornelissen, J. H.
 C. (2016). Corrigendum to: New handbook for standardised measurement of plant functional
- 900 traits worldwide. *Australian Journal of Botany*, 64(8), 715–716.
- Poggiato, G., Münkemüller, T., Bystrova, D., Arbel, J., Clark, J. S., & Thuiller, W. (2021). On the
 Interpretations of Joint Modeling in Community Ecology. *Trends in Ecology & Evolution*, 36(5),
 391–401.
- Pollock, L. J., Morris, W. K., & Vesk, P. A. (2012). The role of functional traits in species
 distributions revealed through a hierarchical model. *Ecography*, 35(8), 716–725.
- 906 Pollock, L. J., Tingley, R., Morris, W. K., Golding, N., O'Hara, R. B., Parris, K. M., Vesk, P. A., &
- 907McCarthy, M. A. (2014). Understanding co-occurrence by modelling species simultaneously908with a Joint Species Distribution Model (JSDM). Methods in Ecology and Evolution / British909Ecological Society, 5(5), 397–406.
- Popovic, G. C., Warton, D. I., Thomson, F. J., Hui, F. K. C., & Moles, A. T. (2019). Untangling direct
 species associations from indirect mediator species effects with graphical models. *Methods in*

- 912 *Ecology and Evolution*, *10*(9), 1571–1583.
- Rahbek, C., Gotelli, N. J., Colwell, R. K., Entsminger, G. L., Rangel, T. F. L. V. B., & Graves, G. R.
 (2007). Predicting continental-scale patterns of bird species richness with spatially explicit
 models. *PNAS*, 274(1607), 165–174.
- Rohr, R. P., Scherer, H., Kehrli, P., Mazza, C., & Bersier, L.-F. (2010). Modeling food webs:
 exploring unexplained structure using latent traits. *The American Naturalist*, *176*(2), 170–177.
- Rothstein, D. E., & Zak, D. R. (2001). Photosynthetic adaptation and acclimation to exploit seasonal
 periods of direct irradiance in three temperate, deciduous-forest herbs. *Functional Ecology*,
 15(6), 722–731.
- Sabatini, F. M., Jiménez-Alfaro, B., Jandt, U., Chytrý, M., Field, R., Kessler, M., Lenoir, J., Schrodt,
 F., Wiser, S. K., Arfin Khan, M. A. S., Attorre, F., Cayuela, L., De Sanctis, M., Dengler, J.,
 Haider, S., Hatim, M. Z., Indreica, A., Jansen, F., Pauchard, A., ... Bruelheide, H. (2022). Global
 patterns of vascular plant alpha diversity. *Nature Communications*, *13*(1), 4683.
- Sabatini, F. M., Lenoir, J., Hattab, T., Arnst, E. A., Chytrý, M., Dengler, J., De Ruffray, P.,
 Hennekens, S. M., Jandt, U., Jansen, F., Jiménez-Alfaro, B., Kattge, J., Levesley, A., Pillar, V.
 D., Purschke, O., Sandel, B., Sultana, F., Aavik, T., Aćić, S., ... Bates, A. (2021). sPlotOpen –
 An environmentally balanced, open-access, global dataset of vegetation plots. *Global Ecology and Biogeography*, *30*(9), 1740–1764.
- Salguero-Gómez, R., Violle, C., Gimenez, O., & Childs, D. (2018). Delivering the promises of trait based approaches to the needs of demographic approaches, and vice versa. *Functional Ecology*,
 32(6), 1424–1435.
- Sanczuk, P., De Lombaerde, E., Haesen, S., Van Meerbeek, K., Luoto, M., Van der Veken, B., Van
 Beek, E., Hermy, M., Verheyen, K., Vangansbeke, P., & De Frenne, P. (2022). Competition
 mediates understorey species range shifts under climate change. *Journal of Ecology*, *110*(8),
 1813–1825.
- Sanczuk, P., De Pauw, K., De Lombaerde, E., Luoto, M., Meeussen, C., Govaert, S., Vanneste, T.,
 Depauw, L., Brunet, J., Cousins, S. A. O., Gasperini, C., Hedwall, P.-O., Iacopetti, G., Lenoir, J.,
 Plue, J., Selvi, F., Spicher, F., Uria-Diez, J., Verheyen, K., ... De Frenne, P. (2023).
- 940 Microclimate and forest density drive plant population dynamics under climate change. *Nature*941 *Climate Change*, *13*(8), 840–847.
- Sanczuk, P., Govaert, S., Meeussen, C., De Pauw, K., Vanneste, T., Depauw, L., Moreira, X.,
 Schoelynck, J., De Boevre, M., De Saeger, S., Bollmann, K., Brunet, J., Cousins, S. A. O., Plue,
 J., Diekmann, M., Graae, B. J., Hedwall, P.-O., Iacopetti, G., Lenoir, J., ... De Frenne, P. (2021).
 Small scale environmental variation modulates plant defence syndromes of understorey plants in
- 946 deciduous forests of Europe. *Global Ecology and Biogeography*, *30*(1), 205–219.
- Sanczuk, P., Landuyt, D., De Lombaerde ,E., Lenoir, J., Lorer, E., Luoto, M., Van Meerbeek ,K.,
 Zellweger, F., & De Frenne, P. (2024). Embracing plant-plant interactions rethinking the
 prediction of species range shifts [Dataset]. *FigShare*, doi: 10.6084/m9.figshare.26943757
- Segar, J., Pereira, H. M., Baeten, L., Bernhardt-Römermann, M., De Frenne, P., Fernández, N.,
 Gilliam, F. S., Lenoir, J., Ortmann-Ajkai, A., Verheyen, K., Waller, D., Teleki, B., Brunet, J.,
- Gilliam, F. S., Lenoir, J., Ortmann-Ajkai, A., Verneyen, K., Waller, D., Teleki, B., Brunet, J.,
 Chudomelová, M., Decocq, G., Dirnböck, T., Hédl, R., Heinken, T., Jaroszewicz, B., ... Staude,
 I. R. (2022). Divergent roles of herbivory in eutrophying forests. *Nature Communications*, 13(1),
 7837.
- Shepard, I. D., Wissinger, S. A., & Greig, H. S. (2021). Elevation alters outcome of competition
 between resident and range-shifting species. *Global Change Biology*, 27(2), 270–281.
- Staniczenko, P. P. A., Sivasubramaniam, P., Suttle, K. B., & Pearson, R. G. (2017). Linking
 macroecology and community ecology: refining predictions of species distributions using biotic
 interaction networks. *Ecology Letters*, 20(6), 693–707.
- Thuiller, W., Calderón-Sanou, I., Chalmandrier, L., Gaüzère, P., O'Connor, L. M. J., Ohlmann, M.,
 Poggiato, G., & Münkemüller, T. (2023). Navigating the integration of biotic interactions in
 biogeography. *Journal of Biogeography*, *51*(4), 550-559.
- Töpper, J. P., Meineri, E., Olsen, S. L., Rydgren, K., Skarpaas, O., & Vandvik, V. (2018). The devil is
 in the detail: Nonadditive and context-dependent plant population responses to increasing
 temperature and precipitation. *Global Change Biology*, 24(10), 4657–4666.
- 966 Townsend Peterson, A., Soberón, J., Pearson, R. G., Anderson, R. P., Martínez-Meyer, E., Nakamura,

- 967 M., & Araújo, M. B. (2011). *Ecological Niches and Geographic Distributions (MPB-49)*.
 968 Princeton University Press.
- 969 Ovaskainen, B. O., Roy, D. B., Fox, R., & Anderson, B. J. (2016). Uncovering hidden spatial
 970 structure in species communities with spatially explicit joint species distribution models.
 971 *Methods in Ecology and Evolution*, 7(4), 428–436.
- Valiente-Banuet, A., & Verdú, M. (2008). Temporal shifts from facilitation to competition occur
 between closely related taxa. *Journal of Ecology*, 96(3), 489–494.
- Verheyen, K., & Hermy, M. (2004). Recruitment and growth of herb-layer species with different
 colonizing capacities in ancient and recent forests. *Journal of Vegetation Science*, 15(1), 125–
 134.
- Verheyen, K., Honnay, O., Motzkin, G., Hermy, M., & Foster, D. R. (2003). Response of forest plant
 species to land-use change: a life-history trait-based approach. *Journal of Ecology*, *91*(4), 563–
 577.
- Wang, Z., Chlus, A., Geygan, R., Ye, Z., Zheng, T., Singh, A., Couture, J. J., Cavender-Bares, J.,
 Kruger, E. L., & Townsend, P. A. (2020). Foliar functional traits from imaging spectroscopy
 across biomes in eastern North America. *The New Phytologist*, 228(2), 494–511.
- Warton, D. I., Blanchet, F. G., O'Hara, R. B., Ovaskainen, O., Taskinen, S., Walker, S. C., & Hui, F.
 K. C. (2015). So Many Variables: Joint Modeling in Community Ecology. *Trends in Ecology & Evolution*, 30(12), 766–779.
- Weber, B., Büdel, B., & Belnap, J. (2016). *Biological Soil Crusts: An Organizing Principle in Drylands.* Springer.
- Williams, J. W., & Jackson, S. T. (2007). Novel climates, no-analog communities, and ecological
 surprises. *Frontiers in Ecology and the Environment*, 5(9), 475–482.
- Wisz, M. S., Pottier, J., Kissling, W. D., Pellissier, L., Lenoir, J., Damgaard, C. F., Dormann, C. F.,
 Forchhammer, M. C., Grytnes, J.-A., Guisan, A., Heikkinen, R. K., Høye, T. T., Kühn, I., Luoto,
 M., Maiorano, L., Nilsson, M.-C., Normand, S., Öckinger, E., Schmidt, N. M., ... Svenning, J.C. (2013). The role of biotic interactions in shaping distributions and realised assemblages of
 species: implications for species distribution modelling. *Biological Reviews of the Cambridge Philosophical Society*, 88(1), 15–30.
- Zellweger, F., De Frenne, P., Lenoir, J., Vangansbeke, P., Verheyen, K., Bernhardt-Römermann, M.,
 Baeten, L., Hédl, R., Berki, I., Brunet, J., Van Calster, H., Chudomelová, M., Decocq, G.,
 Dirnböck, T., Durak, T., Heinken, T., Jaroszewicz, B., Kopecký, M., Máliš, F., ... Coomes, D.
 (2020). Forest microclimate dynamics drive plant responses to warming. *Science*, *368*(6492),
 772–775.
- Zellweger, F., Sulmoni, E., Malle, J. T., Baltensweiler, A., Jonas, T., Zimmermann, N. E., Ginzler, C.,
 Karger, D. N., De Frenne, P., Frey, D., & Webster, C. (2023). Microclimate mapping using novel
 radiative transfer modelling. *EGU Biogeosciences*, 21(2), 605-623.
- Zurell, D., Pollock, L. J., & Thuiller, W. (2018). Do joint species distribution models reliably detect
 interspecific interactions from co-occurrence data in homogenous environments? *Ecography*,
 41(11), 1812–1819.
- Chen, I., Hill, J. K., Ohlemüller, R., Roy, D. B., & Thomas, C. D. (2011). Rapid Range Shifts of
 Species of Climate Warming. *Science*, *333*, 1024–1027.
- Suding, K.N., Goldberg, D.E. & Hartman, K.M. (2003) Relationships among species traits: separating
 levels of response and identifying linkages to abundance. Ecology, 84(1), 1–16.
- Tikhonov, G., Abrego, N., Dunson, D., & Ovaskainen, O. (2017). Using joint species distribution
 models for evaluating how species-to-species associations depend on the environmental context.
- 1013 *Methods in Ecology and Evolution*, 8(4), 443-452.