

## RESEARCH ARTICLE

# Fasciola spp. in Southeast Asia: A systematic review

Vinh Hoang Quang<sup>1,2,3,4</sup>, Bruno Levecke<sup>2</sup>, Dung Do Trung<sup>1</sup>, Brecht Devleesschauwer<sup>2,5</sup>, Binh Vu Thi Lam<sup>1</sup>, Kathy Goossens<sup>3</sup>, Katja Polman<sup>6</sup>, Steven Callens<sup>4</sup>, Pierre Dorny<sup>3</sup>, Veronique Dermauw<sup>3\*</sup>

**1** Department of Parasitology, National Institute of Malaria, Parasitology and Entomology, Hanoi, Vietnam, **2** Department of Translational Physiology, Infectiology and Public Health, Ghent University, Merelbeke, Belgium, **3** Department of Biomedical Sciences, Institute of Tropical Medicine, Antwerp, Belgium, **4** Department of Internal Medicine and Infectious Diseases, Ghent University Hospital, Ghent, Belgium, **5** Department of Epidemiology and Public Health, Sciensano, Brussels, Belgium, **6** Department of Public Health, Institute of Tropical Medicine, Antwerp, Belgium

\* [vdermauw@itg.be](mailto:vdermauw@itg.be)



## Abstract

### OPEN ACCESS

**Citation:** Hoang Quang V, Levecke B, Do Trung D, Devleesschauwer B, Vu Thi Lam B, Goossens K, et al. (2024) *Fasciola* spp. in Southeast Asia: A systematic review. PLoS Negl Trop Dis 18(1): e0011904. <https://doi.org/10.1371/journal.pntd.0011904>

**Editor:** Krystyna Cwiklinski, University of Liverpool, UNITED KINGDOM

**Received:** September 10, 2023

**Accepted:** January 7, 2024

**Published:** January 17, 2024

**Copyright:** © 2024 Hoang Quang et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** All data are available in the manuscript.

**Funding:** This study was conducted within the context of the EmFaVie project, which is funded by the Flemish Interuniversities Council - University Development Co-operation (VLIR-UOS, <https://www.vliruos.be/en>; grant number VN2020SIN317A103; granted to BL & DDT) and the FasciCoM project, which is jointly funded by the Research Foundation - Flanders (FWO, <https://www.fwo.be/en>; grant number G0E2921N;

## Background

Fasciolosis is an emerging public health threat in a number of regions worldwide. To date, we lack an overview of both its occurrence and distribution in Southeast Asia across all actors involved in the life cycle, which impedes the development of disease control measures. Therefore, our objective was to collect recent information on the distribution and the prevalence of *Fasciola* spp. and the associated risk factors for infection in humans, animals, snails and plant carriers in Southeast Asia.

## Methodology

Bibliographic and grey literature databases as well as reference lists of important review articles were searched for relevant records published between January 1<sup>st</sup>, 2000, and June 30<sup>th</sup>, 2022. The systematic review was conducted according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines for reporting systematic reviews. A total of 3,887 records were retrieved, of which 100 were included in the final analysis.

## Principal findings

The studies focused mainly on one host species (96.0%), with *Fasciola* spp. infection in animals being the most studied (72.0%), followed by humans (21.0%). Based on the used inclusion and exclusion criteria, reports were retrieved describing the presence of *Fasciola* spp. infection in seven out of 11 countries in Southeast Asia. Depending on the diagnostic tool applied, the prevalence of *Fasciola* spp. infection ranged between 0.3% and 66.7% in humans, between 0% and 97.8% in animals, and between 0% and 66.2% in snails. There were no studies reporting the presence of metacercariae on plant carriers.

granted to BL) and the Vietnamese National Foundation for Science and Technology Development (NAFOSTED, <https://nafosted.gov.vn/en/>; grant number FWO.108.2020.01; granted to DDT). The funders did not have any role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

## Conclusions/Significance

Our study reconfirms that *Fasciola* spp. infections are widespread and highly prevalent in Southeast Asia, but it remains difficult to accurately assess the true occurrence of *Fasciola* spp. in absence of well-designed surveys covering all hosts. As next steps we propose to assess the occurrence of the infection across all actors involved in the transmission, to identify associated risk factors and to estimate the burden of the disease to support national and international decision makers.

## Author summary

Fasciolosis is a foodborne disease that is caused by a parasitic worm. The disease affects both public and animal health worldwide. The transmission of *Fasciola* is complex and involves humans, animals and plants. The adult worms are observed in both humans and ruminants, while larval stages develop in freshwater snails. The infective larval stage attaches to aquatic plants. In recent decades, human fasciolosis has been considered an emerging disease in many regions of the world, including Southeast Asia. Although various studies surveyed *Fasciola* infection, an overview of existing knowledge on the distribution and prevalence of *Fasciola* across the different hosts is missing for this region. Yet, this is important to develop appropriate disease control measures. In this study, we systematically searched for literature on *Fasciola* spp. infection in Southeast Asia. Our study reconfirms that *Fasciola* is widespread in this region, but that it remains difficult to accurately assess the occurrence of parasite in the absence of well-designed surveys. A firm response to control fasciolosis requires both transdisciplinary and international partnerships. As next steps we propose to assess the occurrence of *Fasciola* spp. across all actors involved in the transmission, to identify associated risk factors and to estimate the disease burden to support national and international decision makers.

## Introduction

Fasciolosis is a parasitic disease caused by the zoonotic trematodes *Fasciola hepatica* and *F. gigantica* [1]. The transmission cycle of *Fasciola* spp. includes a final host (e.g., cattle, sheep, goats, buffaloes and humans), harboring the adult worms in the biliary ducts of the liver, an intermediate host (*Lymnaea* spp.) in which the larval stages develop and multiply, and a carrier (e.g. water plants) to which the metacercariae (i.e. the infective stage) are attached.

Fasciolosis poses a major threat to both animal and public health. In livestock, it is estimated that fasciolosis causes an annual global loss of 3.2 billion US dollars [2]. In humans, fasciolosis has recently emerged as a public health problem. Indeed, it is estimated that 2.4 million individuals are infected worldwide, with 180 million at risk in more than 70 nations [3], Bolivia, Peru, Egypt, Iran and Vietnam being the most affected countries [4]. To reduce the growing global disease burden, the World Health Organization (WHO) promotes large-scale administration of triclabendazole to individuals at risk in endemic countries (e.g., Bolivia, Ecuador and Peru, the Nile Delta in Egypt, and central Vietnam) [4]. However, due to the complexity of the transmission cycle of *Fasciola*, deworming programs aimed exclusively at humans may not be the most cost-effective strategy for long-term management of fasciolosis. Furthermore, there is an increasing number of reports of the ineffectiveness of triclabendazole

against fasciolosis due to the development of resistance of the parasite to this drug, both in humans [5] and livestock, once again highlighting the urgent need for a transdisciplinary approach.

Indeed, a One Health strategy involving a wide range of specialties and sectors would be necessary [6]. Today, such a multisectoral strategy is often ineffective due to a lack of up-to-date information on the disease. For example, a case in point are the Southeast Asian countries (e.g. Thailand, Vietnam, Malaysia and Indonesia), where rice and water plants are important crops, and buffaloes and cattle are often kept alongside these fields [7]. In addition, the tradition of eating raw vegetables and using livestock manure to fertilize crops is still present in this region. In Southeast Asia, human fascioliasis is considered an emerging disease [8,9], and the economic impact of bovine fasciolosis is considerable, with annual losses estimated to range between 314 and 1884 million US dollars [7].

Although various studies surveyed *Fasciola* spp. infection in Southeast Asian countries, an overview of existing knowledge on the distribution and prevalence of *Fasciola* across the different hosts is missing, which impedes the development of a sustainable and integrated control strategy. The present study systematically searched literature on both (i) the geographical distribution and (ii) the prevalence of *Fasciola* infections in humans, mammals, snails, and water plants across Southeast Asia. In addition, we (iii) identified putative risk factors contributing to disease transmission across the different actors of the life cycle.

## Methods

This systematic review has been registered with the International Prospective Register of Systematic Reviews (PROSPERO), reference number: CRD42021261104. The detailed review procedures have been described elsewhere [10]. Briefly, records were retrieved from the bibliographic databases Cumulative Index to Nursing and Allied Health Literature (CINAHL), Excerpta Medica Database (EMBASE), PubMed, Scopus, and Web of Science (all databases), applying the following search phrase: (*Fasciola* OR fascioliasis OR fasciolosis OR *F. hepatica* OR *F. gigantica* OR liver fluke) AND (Southeast Asia OR Brunei OR Cambodia OR Indonesia OR Laos OR Malaysia OR Myanmar OR Philippines OR Singapore OR Thailand OR Timor-Leste OR Vietnam). Furthermore, grey literature was sought from the following sources: Asian Digital Library (<http://www.theadl.com>), the Index Medicus for South-East Asian Region (<https://www.globalindexmedicus.net/biblioteca/imsear/>), and the WHO Institutional Repository for Information Sharing (IRIS) (<http://apps.who.int/iris/>). Finally, reference lists of important review articles were screened for relevant records.

After merging the lists of the retrieved records, duplicates were deleted. In a first step, the relevance of the records was determined based on title and abstract screening. Subsequently, full-text articles were retrieved for selected records and examined for eligibility. Two members of the review team (VHQ and VD) evaluated the records independently for eligibility using a list of inclusion and exclusion criteria. Records were excluded based on the following criteria: (i) language not English, (ii) topic outside the research question (i.e. not covering the geographical distribution, the prevalence of and risk factors for *Fasciola* spp. in humans, animals or plant carriers), (iii) data from outside the study region (i.e. data not from Brunei, Cambodia, Indonesia, Laos, Malaysia, Myanmar, Philippines, Singapore, Thailand, Timor-Leste or Vietnam), (iv) data published beyond the study period (January 1<sup>st</sup>, 2000 and June 30<sup>th</sup>, 2022), (v) no full-text available or (vi) duplicate record. In case the independent eligibility evaluations of the two members of the review team were not in agreement, the records were jointly checked and thoroughly discussed until a consensus on the final evaluation for each article was reached.

Data gathering forms were developed and tested to ensure relevant columns were included for the data we wished to extract. The data were retrieved by one of the review team members (VHQ), and a second team member then double-checked the data extraction (VD). In case the data extraction had to be edited by the second team member, a discussion was held until agreement was attained. From each retained record, author, reference, and publication year were extracted. For population studies, all information was extracted to allow for a description of study design and sampled population, as well as for the estimation of the prevalence and associated 95% confidence interval (95%CI) (e.g., sample size and number of positives). Additionally, for cross-sectional studies and cohort studies investigating risk factors, numerators and denominators required to calculate odds ratios (OR) and associated 95%CI were extracted. For case reports and case series, general descriptors were extracted (e.g., age, gender, diagnostic tests applied). Generally, the review findings were reported in line with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines for reporting systematic reviews [11] (see Fig 1).

The statistical data analyses were run separately for each host type (i.e., humans, mammals, snails, plant carriers). Both for the population studies and case reports and series, a descriptive statistical analysis was conducted. For cross-sectional and cohort studies, the prevalence of *Fasciola* spp. was additionally calculated, as well as the associated Wilson score 95%CI. Odds ratios for the risk factors were calculated as well as associated Wald 95%CI. The significance was set at  $p < 0.05$ . The statistical data analysis was performed in R 4.2.1 [12].

Finally, for each of the included population studies, the risk of bias was assessed by the (modified) Newcastle–Ottawa scale, which entails scoring for six different criteria [13,14]. A score for information on non-respondents was only given where relevant (i.e. in human studies, in the general population), while a score for the ascertainment of exposure and comparability was only given when risk factors were investigated in the study. The total score was calculated by summing all the criteria where the specific study had received a positive score, and dividing by the number of criteria for which a score was given. A study was given the high-quality label in case it reached a score of 100%.

## Results

A total of 3,887 records were retrieved from both the scientific databases and the grey literature sources (Fig 1). Following the removal of duplicates, the titles and abstracts of 2,205 records were reviewed. Subsequently, we retrieved the full text of 251 records and evaluated the eligibility. At this point, 151 records were excluded: 67 were omitted because the study topic was outside the scope of the systematic review; 48 records only reported data on other liver flukes, such as *Clonorchis sinensis* or *Opisthorchis viverrini*, or other parasites, such as soil-transmitted helminths; 10 records provided duplicate results, 10 records were not in English, eight records reported data from outside the study region, and for eight records a full text was not available. Finally, 100 records were included in the data analysis.

Based on the used inclusion and exclusion criteria, records were retrieved reporting the presence of *Fasciola* spp. in seven out of the 11 countries that are part of Southeast Asia: Thailand ( $n = 20$ ), Vietnam ( $n = 19$ ), Indonesia ( $n = 19$ ), Malaysia ( $n = 13$ ), the Philippines ( $n = 13$ ), Cambodia ( $n = 10$ ), and Laos ( $n = 6$ ). For the remaining four countries (Brunei, Myanmar, Singapore, and Timor-Leste) no records were retrieved. The studies focused mainly on one host type (96.0%), with *Fasciola* spp. infection in animals being the most studied (72.0%), followed by humans (21.0%). In only a minority of the records, the presence of *Fasciola* spp. was assessed in snails (10.0%). One study reported *Fasciola* spp. on plant carriers, although not of the infectious life stage (1.0%). One study assessed *Fasciola* spp. infection in

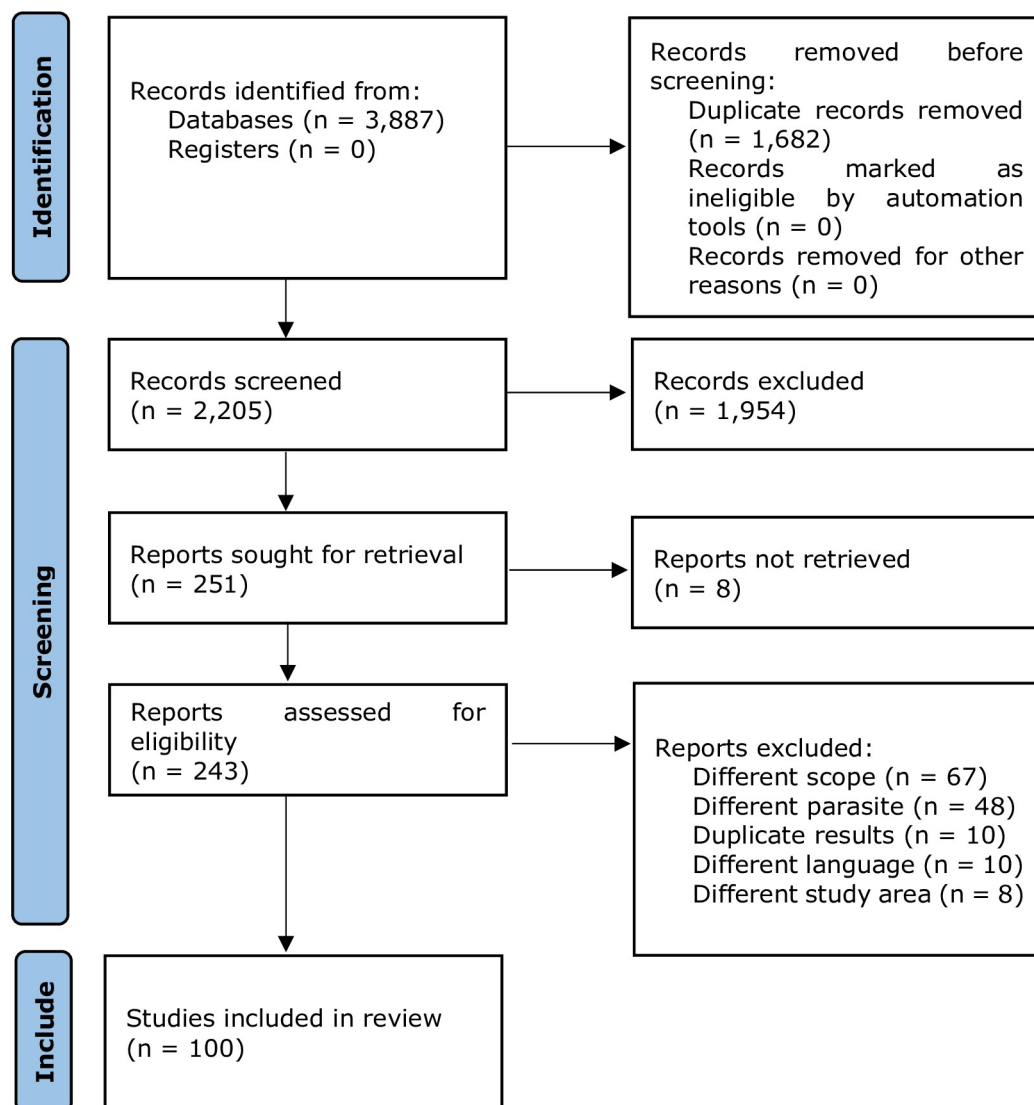


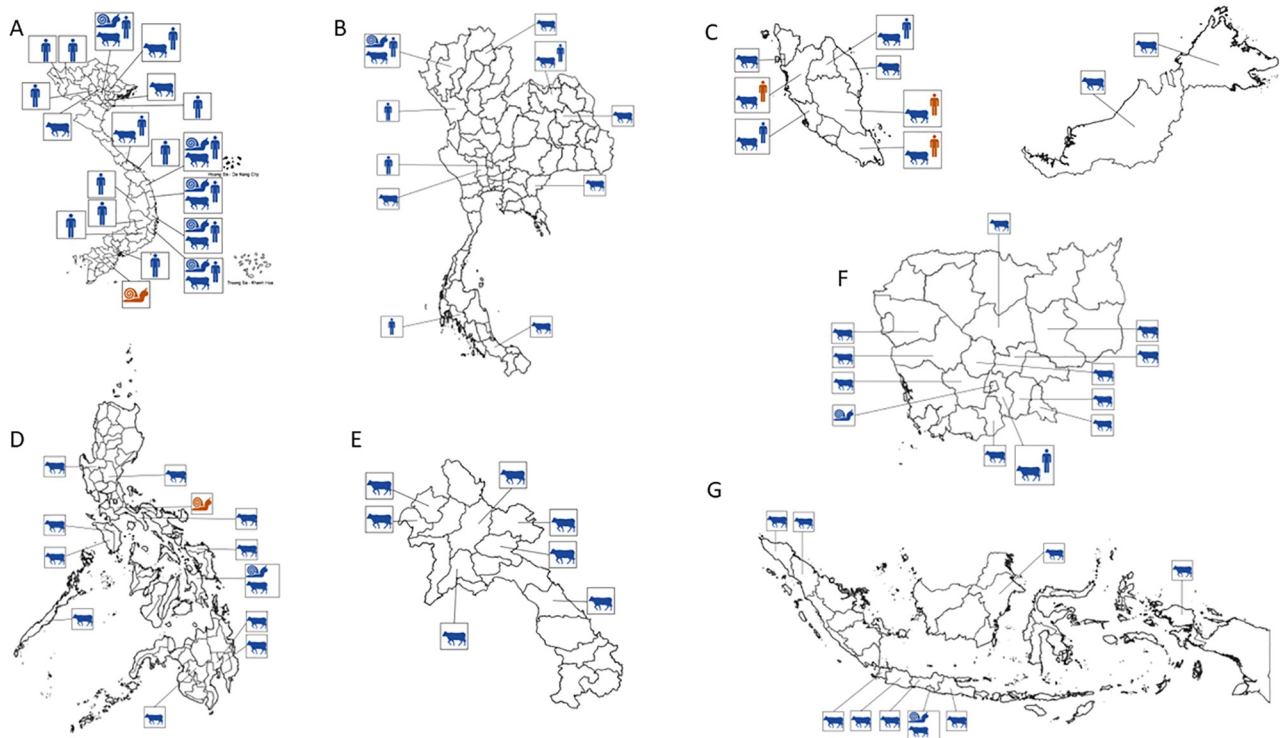
Fig 1. Flow diagram of the database searches according to PRISMA 2020 guidelines [11].

<https://doi.org/10.1371/journal.pntd.0011904.g001>

both cattle and snails [15], one in buffalo and snails [16], another screened both humans and snails [17], and one reported *Fasciola* spp. infection in humans and cattle [18]. The majority of the studies were cross-sectional studies (87.0%). The remaining 13 records were case reports. Fig 2 illustrates the geographical distribution of *Fasciola* spp. infection across the different Southeast Asian countries. In the following paragraphs we will summarize the findings in more details for each of host (humans, animals, and snails) and study type, separately.

### *Fasciola* spp. infection in humans

A total of 10 population surveys, representing 14 studies, investigated the presence of human *Fasciola* spp. infection in Vietnam (n = 7), Cambodia (n = 4), Malaysia (n = 2), and Thailand (n = 1) (Table 1). The studies involved different groups: school children (n = 5), hospital patients (n = 5), the general population (n = 2), and healthy adult subpopulations (n = 2).



**Fig 2. The geographical distribution of *Fasciola* spp. infection across the different Southeast Asian countries.** Each of the 7 panels illustrates the distribution of *Fasciola* spp. infection in humans, animals and snails in Vietnam (Panel A), Thailand (Panel B), Malaysia (Panel C), the Philippines (Panel D), Laos (Panel E), Cambodia (Panel F) and Indonesia (Panel G). A blue silhouette indicates the presence of *Fasciola* spp., while a red silhouette indicates absence, for areas where no icons are shown, as well as for Brunei, Myanmar, Singapore, and Timor-Leste, no reports were found. Shapefiles were republished from the DIVA-GIS database (<https://www.diva-gis.org/gdata>) under a CC BY 4.0 license, with permission from Global Administrative Areas (GADM), original copyright 2018.

<https://doi.org/10.1371/journal.pntd.0011904.g002>

Antibody-Enzyme-Linked Immunosorbent Assay (Ab-ELISA) ( $n = 6$ ) and copro-microscopy ( $n = 5$ ) were the most deployed assays to diagnose *Fasciola* spp. infection. Other diagnostic methods used were copro-antigen-ELISA (copro-Ag-ELISA) ( $n = 1$ ) and copro-Polymerase Chain Reaction (copro-PCR) ( $n = 1$ ) [18]. One study did not mention the diagnostic method [19]. In the six studies that used the Ab-ELISA to detect antibodies against *Fasciola* spp., the prevalence ranged between 0.8 and 66.7%. In the five studies that used copro-microscopy, the prevalence ranged between 0.3 and 46.5%, while in the study that used a copro-PCR, the proportion of samples containing *Fasciola* DNA equaled 0.5% (Table 1).

Two population surveys identified risk factors associated with human *Fasciola* spp. infection, these studies focused in patients and healthy adult populations in Vietnam and Malaysia (Table 2). In both surveys Ab-ELISA was deployed to diagnose *Fasciola* spp. infection. Being female, belonging to an older age group (at least 18 years in the study performed in Malaysia [22], above 60 years in the one in Vietnam [25]), and working with animals for a long period were identified as risk factors.

In total, 872 fasciolosis cases were reported across 11 records in Thailand ( $n = 6$ ), Vietnam ( $n = 4$ ), and Malaysia ( $n = 1$ ). There were 11 individual cases reported in six records (Table 3). On average these cases were 27.1 years old and seven of the 11 cases were female. Nine individuals reported abdominal discomfort (with five reporting abdominal pain and three reporting pain at the right upper quadrant), six patients reported fever, five cases lost weight, and four

Table 1. Human population surveys on *Fasciola* spp. infection in Southeast Asia.

| Country                            | Population studied                                                        | Study Period         | No. of people | No. positive | Prevalence (%) | 95%CI     | Test used        | Species                                        | Reference |
|------------------------------------|---------------------------------------------------------------------------|----------------------|---------------|--------------|----------------|-----------|------------------|------------------------------------------------|-----------|
| <b>General population</b>          |                                                                           |                      |               |              |                |           |                  |                                                |           |
| Malaysia                           | Villagers                                                                 | Nov 2007 to Jul 2009 | 716           | 2            | 0.3            | 0.03–1.01 | Copro-microscopy | <i>Fasciolopsis/Fasciola</i>                   | [20]      |
| Thailand                           | Population older than 15 years                                            | May and Oct 2012     | 221           | 1            | 0.5            | 0.01–2.50 | Copro-microscopy | <i>Fasciolopsis buski/Fasciola/Echinostoma</i> | [21]      |
| <b>Adult healthy subpopulation</b> |                                                                           |                      |               |              |                |           |                  |                                                |           |
| Malaysia                           | Farm workers and dwellers                                                 | Dec 2017 to Dec 2018 | 90            | 60           | 66.7           | 56.4–75.5 | Ab-ELISA         | <i>Fasciola</i> spp.                           | [22]      |
| Vietnam                            | Adult commune members                                                     | Mar and May 2013     | 1,612         | 125          | 7.8            | 6.55–9.16 | Ab-ELISA         | <i>F. gigantica</i> <sup>a</sup>               | [17]      |
| <b>Child healthy subpopulation</b> |                                                                           |                      |               |              |                |           |                  |                                                |           |
| Cambodia                           | Primary school children                                                   | May and June 2011    | 228           | 106          | 46.5           | 40.1–53.0 | Copro-microscopy | Large trematode eggs                           | [18]      |
| Cambodia                           | Primary school children                                                   | May and June 2011    | 221           | 1            | 0.5            | 0.01–2.50 | Copro-PCR        | <i>Fasciola</i> spp.                           | [18]      |
| Cambodia                           | Primary school children                                                   | May and June 2011    | 228           | 6            | 2.6            | 1.2–5.6   | Copro-Ag-ELISA   | <i>Fasciola</i> spp.                           | [18]      |
| Cambodia                           | Primary school children                                                   | May and June 2011    | 240           | 2            | 0.8            | 0.1–3.0   | Ab-ELISA         | <i>Fasciola</i> spp.                           | [18]      |
| Vietnam                            | Fifth grade children (14–15 years old)                                    | Aug 2003 to Feb 2004 | 217           | 2            | 0.9            | 0.11–3.29 | Copro-microscopy | Fasciolidae                                    | [23]      |
| <b>Patient subpopulation</b>       |                                                                           |                      |               |              |                |           |                  |                                                |           |
| Vietnam                            | Hepatic disease patients                                                  | 2006–2007            | 143           | 37           | 25.9           | 19.4–33.6 | Ab-ELISA         | <i>Fasciola</i> spp.                           | [24]      |
| Vietnam                            | Hepatic disease patients                                                  | 2006–2007            | 143           | 3            | 2.1            | 0.4–6.0   | Copro-microscopy | <i>Fasciola</i> spp.                           | [24]      |
| Vietnam                            | Liver tumor patients                                                      | 2006–2010            | 283           | 98           | 34.6           | 29.3–40.3 | N/A              | <i>Fasciola</i> spp.                           | [19]      |
| Vietnam                            | Patients with serum samples submitted for <i>Fasciola</i> spp. antibodies | 2012                 | 10,084        | 590          | 5.8            | 5.41–6.33 | Ab-ELISA         | <i>Fasciola</i> spp.                           | [25]      |
| Vietnam                            | Patients visiting the hospital for helminthiasis                          | 2018                 | 1,120         | 125          | 11.2           | 9.45–13.1 | Ab-ELISA         | <i>Fasciola</i> spp.                           | [26]      |

<sup>a</sup>As reported in the study, without reporting the method for species identification.

<https://doi.org/10.1371/journal.pntd.0011904.t001>

cases showed signs and symptoms such as, anorexia, chills, dark red serpentine track under the skin, and allergy. The remaining 861 fasciolosis cases were documented in five case reports series. In a record conducted in the south of Vietnam [27], 500 cases of fasciolosis were reported, with the majority of the patients (85%) being between 21 and 50 years old, and two-

Table 2. Factors related with *Fasciola* spp. infection investigated in human population surveys in Southeast Asia.

| Country  | Variable                      | Comparison               | Odds ratio (95%CI) | Reference         |
|----------|-------------------------------|--------------------------|--------------------|-------------------|
| Malaysia | Age                           | ≥18 years vs. <18 years  | 3.2 (1.1–9.8)      | [22] <sup>a</sup> |
| Malaysia | Duration working with animals | ≥ 5 years vs. <5 years   | 2.6 (1.1–6.4)      | [22]              |
| Vietnam  | Gender                        | Male vs. female          | 0.8 (0.7–1.0)      | [25]              |
| Vietnam  | Age                           | ≤60 years vs. > 60 years | 0.7 (0.5–0.9)      | [25]              |

<sup>a</sup>Other factors investigated, but not significantly associated with infection: gender, education, monthly household income, drinking water treatment, consumption of raw vegetables, consumption of washed fruits and vegetables, consumption of fully cooked food.

<https://doi.org/10.1371/journal.pntd.0011904.t002>

Table 3. Human fasciolosis case reports in Southeast Asia.

| Country  | Year of diagnosis | Sex    | Age       | Signs and symptoms                                                                                       | Species                          | Tests used                                                                 | Reference |
|----------|-------------------|--------|-----------|----------------------------------------------------------------------------------------------------------|----------------------------------|----------------------------------------------------------------------------|-----------|
| Malaysia | N/A               | Male   | 56 years  | Right hypochondrial pain for one month, chills and rigors                                                | <i>Fasciola</i> spp.             | Histopathology after mastectomy                                            | [31]      |
| Thailand | 1985              | Female | 47 years  | Multiple space-occupying lesions in the liver                                                            | <i>F. hepatica</i> <sup>b</sup>  | Adult worm collection                                                      | [32]      |
|          | 1999              | Female | 40 years  | Abdominal mass for three months                                                                          | <i>F. hepatica</i> <sup>b</sup>  | Adult worm collection                                                      | [32]      |
|          | N/A               | Female | 67 years  | Anorexia and weight loss for one month                                                                   | <i>F. gigantica</i> <sup>b</sup> | Adult worm collection                                                      | [33]      |
|          | N/A               | Female | 36 years  | Right-upper-quadrant abdominal pain with low-grade fever during the previous three months                | <i>F. gigantica</i> <sup>c</sup> | Computed Tomography (CT)-scan, adult worm collection, and molecular assays | [34]      |
| Vietnam  | 2000              | Female | 40 years  | Burning pain at the right upper quadrant and a dark red serpentine track under the skin at the same site | <i>Fasciola</i> spp.             | Adult worm collection                                                      | [35]      |
|          | N/A               | Female | 10 months | Fever (39 °C), upper abdominal pain, weight loss (0.5 kg), allergy                                       | <i>F. gigantica</i> <sup>c</sup> | Ultrasound, stool (+), Ab-ELISA (+), and molecular assays                  | [36]      |
|          | N/A               | Male   | 1 year    | Fever (39–40 °C), upper abdominal pain, weight loss (0.7 kg)                                             | <i>F. gigantica</i> <sup>c</sup> | Ultrasound, stool (+), Ab-ELISA (+), and molecular assays                  | [36]      |
|          | N/A               | Female | 3 years   | Abdominal pain, chest pain, fever (38.5°C) as first symptoms. Weight loss (1.5 kg)                       | <i>F. gigantica</i> <sup>a</sup> | Ultrasound, stool (-), and Ab-ELISA (+)                                    | [36]      |
|          | N/A               | Male   | 3 years   | Abdominal pain, cough, chest pain, fever (39 °C)                                                         | <i>F. gigantica</i> <sup>a</sup> | Ultrasound, stool (-), and Ab-ELISA (+)                                    | [36]      |
|          | N/A               | Male   | 4 years   | Fever (39 °C) followed by abdominal pain appearing after two weeks and weight loss of around 2 kg/month  | <i>F. gigantica</i> <sup>b</sup> | Ultrasound, stool (-), and Ab-ELISA (+)                                    | [36]      |

<sup>a</sup>As reported in the study, without reporting the method for species identification;

<sup>b</sup>Based on morphometrics;

<sup>c</sup>Based on molecular analyses;

+: positive test result; -: negative test result

<https://doi.org/10.1371/journal.pntd.0011904.t003>

third being female. Another record described fasciolosis in 145 people diagnosed in a tertiary referral hospital in northern Vietnam [28]. Most of these cases (68.3%) were between 30 and 59 years old, and about half (51.0%) were male. The most frequently reported symptoms were upper quadrant (61.4%) and epigastric (35.2%) pain. In Thailand, fasciolosis cases were reported in three records, with the most common signs and symptoms being, abdominal pain (74.9%), especially in the right upper quadrant, abdominal distension (31.4%), weight loss (29.1%) in one record [29], while in another record [30] abdominal pain was present in all 12 patients, fever in three, and jaundice in one. Eleven patients had eosinophilia [30].

### *Fasciola* spp. infection in animals

A total of 70 population surveys were retrieved, describing 104 studies on *Fasciola* spp. infection in animals. These studies were conducted in seven countries, in Indonesia (n = 21), Malaysia (n = 18), the Philippines (n = 18), Thailand (n = 17), Cambodia (n = 11), Laos (n = 10), and Vietnam (n = 9). The most studied animal species were cattle (n = 67), buffaloes (n = 18), and both animal species surveyed in six studies. Other species examined were goats (n = 6), pigs (n = 2), deer (n = 1), monkeys (n = 1), leopard cats (n = 1), elephants (n = 1), and sheep and goats (n = 1). Copro-microscopy was the most commonly used diagnostic method, a total of 75 surveys used this diagnostic tool to demonstrate *Fasciola* spp. eggs in feces. Other

Table 4. Animal population surveys on *Fasciola* spp. infection in Southeast Asia using copro-microscopic examination.

| Country       | Study Period          | No. of animals  | No. positive | Prevalence (%)         | 95%CI     | Species                          | Reference |
|---------------|-----------------------|-----------------|--------------|------------------------|-----------|----------------------------------|-----------|
| <b>Cattle</b> |                       |                 |              |                        |           |                                  |           |
| Cambodia      | Dec 1998 to Nov 1999  | 575             | 142          | 24.7                   | 21.3–28.3 | <i>Fasciola</i> spp.             | [37]      |
|               | 1999 to 2000          | N/A             | N/A          | N/A <sup>a</sup>       | N/A       | <i>F. gigantica</i> <sup>b</sup> | [38]      |
|               | Apr 2008              | 540             | N/A          | 9.5                    | N/A       | <i>F. gigantica</i> <sup>b</sup> | [39]      |
|               | July 2008 to May 2009 | 2,391           | N/A          | 5.0–20.0               | N/A       | <i>Fasciola</i> spp.             | [40]      |
|               | Sep 2008              | 540             | N/A          | 21.5                   | N/A       | <i>F. gigantica</i> <sup>b</sup> | [39]      |
|               | May to June 2011      | 205             | 182          | 88.8 <sup>c</sup>      | N/A       | <i>Fasciola</i> spp.             | [18]      |
|               | Dec 2014 to June 2015 | 172             | 28           | 16.3                   | 11.5–22.5 | <i>Fasciola</i> spp.             | [41]      |
| Indonesia     | Nov 2011 to Jan 2012  | 394             | 49           | 12.5                   | 9.54–16.1 | <i>F. gigantica</i> <sup>b</sup> | [42]      |
|               | Feb to March 2016     | 369             | 129          | 35.0                   | 30.3–40.0 | <i>Fasciola</i> spp.             | [43]      |
|               | Feb 2017              | 109             | 36           | 33.0                   | 24.9–42.3 | <i>F. gigantica</i> <sup>b</sup> | [44]      |
|               | Apr to Oct 2018       | 100             | 48           | 48.0                   | 38.5–57.7 | <i>Fasciola</i> spp.             | [45]      |
|               | Jun to Sep 2018       | 314             | 113          | 36.0                   | 30.9–41.4 | <i>Fasciola</i> spp.             | [46]      |
|               | 2020                  | 400             | 66           | 16.5                   | 13.2–20.5 | <i>Fasciola</i> spp.             | [47]      |
|               | N/A                   | 134             | 2            | 1.5                    | 0.18–5.29 | <i>Fasciola</i> spp.             | [48]      |
|               | N/A                   | 49              | 11           | 22.4                   | 13.0–35.9 | <i>Fasciola</i> spp.             | [49]      |
|               | N/A                   | 100             | 12           | 12.0                   | 7.0–19.8  | <i>Fasciola</i> spp.             | [50]      |
|               | N/A                   | 30              | 7            | 23.3                   | 11.8–40.9 | <i>Fasciola</i> spp.             | [51]      |
|               | N/A                   | 103             | 58           | 56.3                   | 46.7–65.5 | <i>F. gigantica</i> <sup>b</sup> | [52]      |
|               | N/A                   | 35              | 13           | 37.1                   | 23.2–53.7 | <i>Fasciola</i> spp.             | [53]      |
| Laos          | Sep 2010 to Feb 2011  | 172             | 20           | 11.6                   | 7.6–17.3  | <i>F. gigantica</i> <sup>b</sup> | [54]      |
|               | May and June 2015     | 21 <sup>d</sup> | 3            | 14.3                   | 3.0–36.3  | <i>Fasciola</i> spp.             | [55]      |
|               | 2016                  | 241             | 95           | 39.4                   | 33.5–45.7 | <i>F. gigantica</i> <sup>b</sup> | [56]      |
| Malaysia      | Jan 2007 to Dec 2017  | 1,988           | 35           | 1.8                    | 1.3–2.4   | <i>Fasciola</i> spp.             | [57]      |
|               | 2008–2018             | 541             | 170          | 31.4                   | 27.7–35.5 | <i>Fasciola</i> spp.             | [58]      |
|               | Mar 2015 to Jan 2016  | 219             | 111          | 50.7                   | 44.1–57.2 | <i>Fasciola</i> spp.             | [59]      |
|               | Dec 2017 to June 2018 | 308             | 45           | 14.6                   | 11.1–19.0 | <i>Fasciola</i> spp.             | [60]      |
|               | N/A                   | 33              | 6            | 18.2                   | 8.6–34.4  | <i>Fasciola</i> spp.             | [61]      |
| Philippines   | July to Sep 2011      | 45              | 44           | 97.8                   | 88.4–99.6 | <i>F. gigantica</i> <sup>b</sup> | [62]      |
|               | 2017                  | 15              | 5            | 33.3                   | 15.2–58.3 | <i>Fasciola</i> spp.             | [63]      |
|               | N/A                   | 250             | N/A          | 27.1–49.0 <sup>e</sup> | N/A       | <i>F. gigantica</i> <sup>b</sup> | [64]      |
|               | N/A                   | 80              | 13           | 16.3                   | 9.7–25.8  | <i>Fasciola</i> spp.             | [65]      |
| Thailand      | Mar to Sep 2007       | 1,599           | 59           | 3.7                    | 2.87–4.73 | <i>Fasciola</i> spp.             | [66]      |
|               | May 2013              | 322             | 34           | 10.6                   | 7.7–14.4  | <i>Fasciola</i> spp.             | [67]      |
|               | 2016 to 2019          | 1,501           | 35           | 2.3                    | 1.7–3.22  | <i>Fasciola</i> spp.             | [68]      |
|               | Nov 2017 to July 2018 | 38              | 3            | 7.9                    | 1.6–21.4  | <i>Fasciola</i> spp.             | [67]      |
|               | Jan and June 2019     | 311             | 26           | 8.4                    | 5.8–12.0  | <i>Fasciola</i> spp.             | [69]      |
|               | Jan 2019              | 239             | 19           | 8.0                    | 5.1–12.1  | <i>Fasciola</i> spp.             | [70]      |
|               | Aug to Nov 2020       | 333             | 67           | 20.1                   | 16.2–24.8 | <i>Fasciola</i> spp.             | [67]      |
|               | Oct and Nov 2020      | 46              | 16           | 34.8                   | 22.7–49.2 | <i>Fasciola</i> spp.             | [71]      |
|               | N/A                   | 272             | 28           | 10.3                   | 7.2–14.5  | <i>Fasciola</i> spp.             | [72]      |
| Vietnam       | Jan 1999 to Jan 2000  | 74              | N/A          | 22                     | 9.8–22.6  | <i>Fasciola</i> spp.             | [73]      |
|               | June 2002 to Mar 2003 | 157–204         | N/A          | 0–31.0                 | N/A       | <i>Fasciola</i> spp.             | [74]      |
|               | June to Sep 2006      | 266             | N/A          | 28.0–39.0              | N/A       | <i>Fasciola</i> spp.             | [75]      |
|               | Apr 2008 to May 2009  | 1,075           | 487          | 45.3                   | 42.3–48.3 | <i>Fasciola</i> spp.             | [15]      |
|               | June to Sept 2008     | 825             | 453          | 54.9                   | 51.5–58.3 | <i>Fasciola</i> spp.             | [76]      |
|               | Nov 2010              | 153             | 89           | 58.2                   | 50.2–65.7 | <i>Fasciola</i> spp.             | [77]      |
|               | Apr and Oct 2014      | 572             | 134          | 23.4                   | 20.1–27.1 | <i>Fasciola</i> spp.             | [78]      |

(Continued)

Table 4. (Continued)

| Country                   | Study Period             | No. of animals   | No. positive | Prevalence (%) | 95%CI     | Species                          | Reference |
|---------------------------|--------------------------|------------------|--------------|----------------|-----------|----------------------------------|-----------|
| <b>Buffalo</b>            |                          |                  |              |                |           |                                  |           |
| Indonesia                 | Mar 2017 to May 2018     | 580              | 93           | 16.0           | 13.3–19.2 | <i>F. gigantica</i> <sup>f</sup> | [79]      |
| Laos                      | May to June 2015         | 61 <sup>d</sup>  | 1            | 1.6            | 0.4–8.8   | <i>Fasciola</i> spp.             | [55]      |
| Malaysia                  | 2008–2018                | 219              | 19           | 8.7            | 5.6–13.2  | <i>Fasciola</i> spp.             | [58]      |
| Philippines               | Apr 2006                 | 15               | 3            | 20.0           | 4.3–48.1  | <i>Fasciola</i> spp.             | [80]      |
|                           | July to Sep 2011         | 105              | 100          | 95.2           | 89.3–97.9 | <i>F. gigantica</i> <sup>g</sup> | [62]      |
|                           | Dec to June 2015 to 2016 | 335              | 213          | 63.6           | 58.3–68.6 | <i>Fasciola</i> spp.             | [16]      |
|                           | Sep 2019                 | 91               | 49           | 53.8           | 43.7–63.7 | <i>Fasciola</i> spp.             | [81]      |
|                           | N/A                      | 32               | N/A          | 37.5–50.0      | N/A       | <i>F. gigantica</i> <sup>b</sup> | [64]      |
|                           | N/A                      | 48               | 12           | 25.0           | 14.9–38.8 | <i>Fasciola</i> spp.             | [82]      |
|                           | N/A                      | 80               | 15           | 18.8           | 11.7–28.7 | <i>Fasciola</i> spp.             | [65]      |
| Thailand                  | May 2013                 | 180              | 37           | 20.6           | 15.3–27.0 | <i>Fasciola</i> spp.             | [67]      |
|                           | N/A                      | 83               | 25           | 30.1           | 21.3–40.7 | <i>Fasciola</i> spp.             | [72]      |
|                           | N/A                      | 1,120            | N/A          | 20.0–61.0      | N/A       | <i>Fasciola</i> spp.             | [83]      |
| <b>Cattle and buffalo</b> |                          |                  |              |                |           |                                  |           |
| Cambodia                  | Dec 2001 to Jan 2002     | 1,406            | 160          | 11.4           | 9.8–13.1  | <i>F. gigantica</i> <sup>b</sup> | [84]      |
|                           | 2004                     | 801              | 103          | 12.9           | 10.7–15.4 | <i>Fasciola</i> spp.             | [85]      |
| Laos                      | 2010 to 2011             | 1,262            | 217          | 17.2           | 15.2–19.4 | <i>F. gigantica</i> <sup>b</sup> | [86]      |
|                           | Jan and Feb 2011         | 306 <sup>d</sup> | 81           | 26.5           | 21.8–31.7 | <i>F. gigantica</i> <sup>b</sup> | [87]      |
| <b>Goat</b>               |                          |                  |              |                |           |                                  |           |
| Indonesia                 | N/A                      | 32               | 11           | 34.4           | 20.4–51.7 | <i>Fasciola</i> spp.             | [51]      |
| Laos                      | May to June 2015         | 18 <sup>d</sup>  | 1            | 5.6            | 0.1–27.3  | <i>Fasciola</i> spp.             | [55]      |
| Malaysia                  | 2008 to 2018             | 221              | 5            | 2.3            | 1.0–5.2   | <i>Fasciola</i> spp.             | [58]      |
| Philippines               | N/A                      | 85               | 9            | 10.6           | 5.7–18.9  | <i>Fasciola</i> spp.             | [65]      |
| <b>Sheep and goat</b>     |                          |                  |              |                |           |                                  |           |
| Malaysia                  | Mar to Dec 2015          | 267              | 0            | 0              | 0–1.4     | <i>Fasciola</i> spp.             | [88]      |
| <b>Pig</b>                |                          |                  |              |                |           |                                  |           |
| Indonesia                 | N/A                      | 4                | 1            | 25.0           | 0.6–80.6  | <i>Fasciola</i> spp.             | [51]      |
| Philippines               | Sep to Oct 2013          | 36 <sup>b</sup>  | 1            | 2.8            | 0.07–14.5 | <i>Fasciola</i> spp.             | [89]      |
| <b>Deer</b>               |                          |                  |              |                |           |                                  |           |
| Indonesia                 | N/A                      | 75               | 20           | 26.7           | 18.0–37.6 | <i>Fasciola</i> spp.             | [90]      |
| <b>Asian elephant</b>     |                          |                  |              |                |           |                                  |           |
| Malaysia                  | N/A                      | 104              | 73           | 70.2           | 60.8–78.1 | <i>Fasciola</i> spp.             | [91]      |
| <b>Proboscis monkey</b>   |                          |                  |              |                |           |                                  |           |
| Philippines               | 2011 to 2013             | 65               | 1            | 1.5            | 0.04–8.3  | <i>Fasciola</i> spp.             | [92]      |
| <b>Leopard cat</b>        |                          |                  |              |                |           |                                  |           |
| Philippines               | May 2013 to Feb 2014     | 3                | 1            | 33.3           | 0.8–90.6  | Fasciolidae                      | [93]      |

<sup>a</sup>Reporting the village one-month incidence only, varying between 3.4 and 87.5%;<sup>b</sup>As reported in the study, without reporting the method for species identification;<sup>c</sup>Prevalence based on the combined use of copro-microscopic examination and copro-Ag-ELISA<sup>d</sup>Prevalence represents the proportion of composite samples containing eggs;<sup>e</sup>Prevalence based on the combined use of copro-microscopic examination and worm collection;<sup>f</sup>Based on morphometrics;<sup>g</sup>Based on molecular analyses;<https://doi.org/10.1371/journal.pntd.0011904.t004>

Table 5. Animal population surveys on *Fasciola* spp. infection in Southeast Asia using other methods than copro-microscopic examination.

| Country                   | Study Period          | Diagnostic method | No. of animals | No. positive | Prevalence (%) | 95% CI    | Species                                                        | Reference |
|---------------------------|-----------------------|-------------------|----------------|--------------|----------------|-----------|----------------------------------------------------------------|-----------|
| <b>Cattle</b>             |                       |                   |                |              |                |           |                                                                |           |
| Cambodia                  | May to June 2011      | Liver examination | 191            | 35           | 18.3           | 13.5–24.4 | <i>Fasciola</i> spp.                                           | [18]      |
|                           | July 2017             | Liver examination | 294            | 21           | 7.1            | 4.72–10.7 | <i>F. gigantica</i> <sup>a</sup>                               | [94]      |
| Indonesia                 | Feb to Mar 2019       | Liver examination | 100            | 39           | 39.0           | 30.0–48.8 | <i>Fasciola</i> spp.                                           | [95]      |
|                           | N/A                   | Copro-Ag-ELISA    | 150            | 87           | 58.0           | 50.0–65.6 | <i>F. gigantica</i> <sup>c</sup>                               | [96]      |
|                           | N/A                   | Liver examination | 150            | 92           | 61.3           | 53.3–68.8 | <i>F. gigantica</i> <sup>c</sup>                               | [96]      |
|                           | N/A                   | Liver examination | 100            | 17           | 17.0           | 10.9–25.5 | <i>Fasciola</i> spp.                                           | [50]      |
|                           | N/A                   | Liver examination | 157            | 34           | 21.7           | 15.9–28.8 | <i>F. gigantica</i> <sup>b</sup>                               | [97]      |
| Laos                      | Sep 2010 to Feb 2012  | Ab-ELISA          | 172            | 163          | 94.8           | 90.4–97.2 | <i>F. gigantica</i> <sup>c</sup>                               | [54]      |
|                           | July 2018 to Feb 2019 | Copro-PCR         | 153            | 91           | 59.5           | 51.6–66.9 | <i>Fasciola</i> spp. hybrids; <i>F. gigantica</i> <sup>a</sup> | [98]      |
| Malaysia                  | 2008–2018             | Liver examination | 1,128          | 19           | 1.7            | 1.1–2.6   | <i>Fasciola</i> spp.                                           | [58]      |
|                           | Feb to Aug 2013       | Liver examination | 67             | 5            | 7.5            | 3.2–16.3  | <i>F. gigantica</i> <sup>b</sup>                               | [99]      |
|                           | Mar 2015 to Jan 2017  | Ab-ELISA          | 85             | 70           | 82.4           | 72.9–89.0 | <i>Fasciola</i> spp.                                           | [59]      |
|                           | Dec 2017 to June 2018 | Ab-ELISA          | 308            | 115          | 37.3           | 32.1–42.9 | <i>Fasciola</i> spp.                                           | [60]      |
| Philippines               | July to Sep 2011      | Copro-PCR         | 45             | 42           | 93.3           | 82.1–97.7 | <i>F. gigantica</i> <sup>a</sup>                               | [62]      |
|                           | 2017                  | Liver examination | 15             | 5            | 33.3           | 4.3–48.1  | <i>Fasciola</i> spp.                                           | [63]      |
| Thailand                  | Oct 2010 to Sep 2012  | Liver examination | 51             | 27           | 52.9           | 39.5–65.9 | <i>F. gigantica</i> <sup>a</sup>                               | [100]     |
|                           | Oct and Nov 2021      | Copro-PCR         | 46             | 31           | 67.4           | 53.0–79.1 | <i>Fasciola</i> spp.                                           | [71]      |
|                           | N/A                   | Ab-ELISA          | 277            | 95           | 34.3           | 29.0–40.1 | <i>Fasciola</i> spp.                                           | [72]      |
| Vietnam                   | Jun to Sep 2008       | Ab-ELISA          | 400            | 289          | 72.3           | 67.7–76.4 | <i>Fasciola</i> spp.                                           | [76]      |
|                           | Aor 2008 to May 2010  | Ab-ELISA          | 235            | 205          | 87.2           | 82.4–90.9 | <i>Fasciola</i> spp.                                           | [15]      |
| <b>Buffalo</b>            |                       |                   |                |              |                |           |                                                                |           |
| Malaysia                  | 2008–2018             | Liver examination | 245            | 2            | 0.8            | 0.1–2.9   | <i>Fasciola</i> spp.                                           | [58]      |
| Malaysia                  | Feb to Aug 2013       | Liver examination | 13             | 1            | 7.7            | 1.4–33.3  | <i>F. gigantica</i> <sup>b</sup>                               | [99]      |
| Philippines               | July to Sep 2011      | Copro-PCR         | 105            | 101          | 96.2           | 90.6–98.5 | <i>F. gigantica</i> <sup>a</sup>                               | [62]      |
| Thailand                  | Oct 2010 to Sep 2012  | Liver examination | 55             | 37           | 67.3           | 54.1–78.2 | <i>F. gigantica</i> <sup>a</sup>                               | [100]     |
|                           | N/A                   | Ab-ELISA          | 95             | 75           | 78.9           | 69.7–85.9 | <i>Fasciola</i> spp.                                           | [72]      |
| <b>Cattle and buffalo</b> |                       |                   |                |              |                |           |                                                                |           |
| Laos                      | Mar to June 2011      | Liver examination | 123            | 42           | 34.1           | 26.4–42.9 | <i>F. gigantica</i> <sup>c</sup>                               | [87]      |
| Malaysia                  | 2018                  | Liver examination | 7,786          | 25           | 0.3            | 0.2–0.5   | <i>F. gigantica</i> <sup>a</sup>                               | [101]     |
| <b>Goat</b>               |                       |                   |                |              |                |           |                                                                |           |
| Malaysia                  | 2008–2018             | Liver examination | 924            | 0            | 0              | 0–0.4     | <i>Fasciola</i> spp.                                           | [58]      |
|                           | Mar to Dec 2015       | Ab-ELISA          | 86             | 76           | 88.4           | 79.9–93.6 | <i>Fasciola</i> spp.                                           | [88]      |

<sup>a</sup>Based on molecular analyses;<sup>b</sup>based on morphometrics;<sup>c</sup>As reported in the study, without reporting the method for species identification;<https://doi.org/10.1371/journal.pntd.0011904.t005>

diagnostic methods deployed were liver inspection at slaughter ( $n = 16$ ), Ab-ELISA ( $n = 8$ ), copro-PCR ( $n = 4$ ) and copro-Ag-ELISA ( $n = 1$ ). The prevalence across the different animal species and diagnostic methods are reported in Table 4 (copro-microscopy) and Table 5 (liver inspection, Ab-ELISA, copro-PCR and copro-Ag-ELISA).

Using copro-microscopy, *Fasciola* spp. infection rates varied from 0 to 97.8% in cattle and buffaloes, from 0 to 34.4% in goats, and from 2.8 to 25.0% in pigs. Based on liver inspection, the infection rates in cattle and buffalo ranged from 0.3 to 67.3%, while the prevalence ranged from 34.3 to 94.8% when the Ab-ELISA was used. When using copro-PCR, the prevalence

Table 6. Factors related with *Fasciola* spp. infection investigated in animal population surveys in Southeast Asia.

| Country                  | Variable                              | Comparison                   | Odds ratio (95%CI)           | Reference |
|--------------------------|---------------------------------------|------------------------------|------------------------------|-----------|
| <b>Cattle</b>            |                                       |                              |                              |           |
| Cambodia                 | Cut and carry grass from natural lake | No vs. yes                   | 0.06 (0.01–0.5) <sup>a</sup> | [41]      |
|                          | Flooding                              | Yes vs. no                   | 20.7 (2.7–156.3)             | [41]      |
|                          | Age                                   | ≥ 3 years vs. < 3 years      | 3.5 (1.3–9.2)                | [41]      |
|                          | Sex                                   | Female vs. male              | 8.1 (1.1–61.9)               | [41]      |
| Indonesia                | Sex                                   | Male vs. female              | 3.6 (1.6–8.3)                | [52]      |
|                          | Deworming program                     | No vs. yes                   | 2.3 (1.2–4.2)                | [47]      |
|                          | Manure processed                      | No vs. yes                   | 3.1 (1.6–5.9)                | [47]      |
|                          | Relative humidity                     | ≥ 70% vs. < 70%              | 2.0 (1.1–3.5)                | [47]      |
|                          | Participation in extension program    | Never vs. ever               | 2.6 (1.5–4.5)                | [47]      |
| Thailand                 | Age                                   | > 7 years vs. ≤ 7 years      | 1.9 (1.1–3.4)                | [72]      |
|                          | Sex                                   | Female vs. male              | 3.1 (1.5–6.2)                | [72]      |
|                          | Age                                   | > 4 years vs. 2–4 years      | 3.5 (1.4–9.0)                | [69]      |
|                          | Age                                   | > 4 years vs. < 2 years      | 3.2 (1.1–8.9)                | [69]      |
| Vietnam                  | Age                                   | ≥ 2 years vs. < 2 years      | 3.6 (2.7–4.9)                | [76]      |
|                          | Age                                   | ≥ 1 years vs. < 1 year       | 7.4 (4.6–12.1)               | [76]      |
|                          | Season                                | Rainy vs. dry                | 1.7 (1.3–2.2)                | [15]      |
|                          | Age                                   | > 2 years vs. ≤ 2 years      | 1.9 (1.5–2.5)                | [15]      |
|                          | Treatment                             | Yes vs. no                   | 0.09 (0.06–0.1)              | [78]      |
| <b>Buffalo</b>           |                                       |                              |                              |           |
| Indonesia<br>Philippines | Age                                   | > 32 months vs. 18–32 months | 2.9 (1.2–6.8)                | [79]      |
|                          | Age                                   | Per year increase            | 1.1 (1.1–1.2) <sup>a</sup>   | [16]      |
|                          | Wallowing pond                        | Yes vs. no                   | 0.1 (0.08–0.2) <sup>a</sup>  | [16]      |
|                          | Irrigation water                      | Yes vs. no                   | 10.4 (1.9–81.7) <sup>a</sup> | [16]      |

<sup>a</sup>Odds ratio (95%CI) from multivariable model<https://doi.org/10.1371/journal.pntd.0011904.t006>

ranged from 59.5 to 96.2%. Only one study applied copro-Ag-ELISA was to detect *Fasciola* spp. antigens in feces, with a reported prevalence of 58.0%.

Nine surveys reported data for factors associated with *Fasciola* spp. infection in animals, in Cambodia, Indonesia, the Philippines, Thailand and Vietnam (Table 6). The studies investigated *Fasciola* spp. infections in cattle or buffaloes, and all but two studies were based on copro-microscopic examination. Increasing age was a risk factor for cattle and buffalo in seven studies. Sex was found to be a risk factor in three studies, two of which determined that female animals had a higher risk of being infected. Furthermore, other risk factors for infection in cattle were flooding [41], the absence of deworming programs, not processing (i.e.) manure before further use, the presence of a high relative humidity, owners not participating in extension programs [47], and the rainy season [15], whereas in buffalo, the use of irrigation water for drinking was a risk factor [16]. Avoiding the use of cut and carry grass from a natural lake [41], treating animals [78] and access to wallowing ponds [16] were considered as protective factors in cattle or buffalo.

The number of case reports in animals was limited. One record described a fasciolosis case in a two-year-old male buffalo in Malaysia. In this case, *F. gigantica* adult worms were collected in the bile duct of the liver, and *Fasciola* eggs were found in the feces with a sedimentation test [102]. Another report described adult *Fasciola* in the liver of a deer in the Philippines [103].

Table 7. Snail population surveys on *Fasciola* spp. infection in Southeast Asia.

| Country     | Study Period          | Population studied                            | No of snails | No positive | Prevalence (%) | 95%CI     | Test used  | Species                           | Reference |
|-------------|-----------------------|-----------------------------------------------|--------------|-------------|----------------|-----------|------------|-----------------------------------|-----------|
| Cambodia    | Mar 2007 to May 2008  | <i>L. auricularia rubiginosa</i> <sup>a</sup> | 79           | N/A         | 1.3            | N/A       | Shedding   | <i>Fasciola</i> spp. <sup>b</sup> | [106]     |
| Indonesia   | Apr 1993              | <i>L. rubiginosa</i>                          | 6,225        | 10          | 0.2            | 0.1–0.3   | Crushing   | <i>F. gigantica</i> <sup>c</sup>  | [105]     |
|             | N/A                   | Lymnaeid snails                               | 320          | 12          | 3.8            | 2.2–6.4   | Dissection | <i>F. gigantica</i> <sup>d</sup>  | [110]     |
| Philippines | Dec 2015 to June 2016 | Lymnaeid snails                               | 748          | 495         | 66.2           | 62.7–69.5 | Crushing   | <i>Fasciola</i> spp.              | [16]      |
|             | N/A                   | Lymnaeid snails                               | 750          | 0           | 0              | 0–0.5     | Shedding   | <i>Fasciola</i> spp.              | [111]     |
| Thailand    | Apr 2008 to Feb 2012  | <i>Bithynia siamensis</i> <sup>e</sup>        | 365          | 2           | 0.6            | 0.1–2.0   | Crushing   | <i>Fasciola</i> spp. <sup>b</sup> | [108]     |
|             | Apr 2008 to Feb 2012  | <i>L. auricularia rubiginosa</i>              | 60           | 0           | 0              | 0.0–6.0   | Crushing   | <i>Fasciola</i> spp. <sup>b</sup> | [108]     |
| Vietnam     | Apr to May 2013       | <i>Lymnaea</i> spp.                           | 2,669        | 14          | 0.5            | 0.3–0.9   | Crushing   | <i>Fasciola</i> spp.              | [17]      |
|             | N/A                   | <i>L. rubiginosa</i>                          | 10,159       | 178         | 1.8            | 1.5–2.0   | Crushing   | <i>F. gigantica</i> <sup>c</sup>  | [107]     |
|             | N/A                   | <i>L. viridis</i>                             | 3,269        | 31          | 0.9            | 0.7–1.3   | Crushing   | <i>Fasciola</i> spp.              | [15]      |
|             | N/A                   | <i>L. swinhoei</i>                            | 1,128        | 7           | 0.6            | 0.3–1.3   | Crushing   | <i>Fasciola</i> spp.              | [15]      |
|             | N/A                   | <i>Radix rubiginosa</i>                       | 1,000        | 0           | 0              | 0–0.4     | Crushing   | <i>F. gigantica</i> <sup>c</sup>  | [104]     |
|             | N/A                   | <i>Radix auricularia</i>                      | 6,324        | 0           | 0              | 0–0.06    | Crushing   | <i>F. gigantica</i> <sup>c</sup>  | [104]     |
|             | N/A                   | <i>Austropeplea viridis</i>                   | 17,167       | 124         | 0.7            | 0.6–0.9   | Crushing   | <i>F. gigantica</i> <sup>c</sup>  | [104]     |

<sup>a</sup>Seven other species were all negative;

<sup>b</sup>Based on the presence of gymnocephalous cercariae, a cercarial type to which *Fasciola* spp. belong (among others);

<sup>c</sup>As reported in the study, without reporting the method for species identification;

<sup>d</sup>Based on molecular analyses;

<sup>e</sup>Twelve other species were all negative

<https://doi.org/10.1371/journal.pntd.0011904.t007>

### *Fasciola* spp. in the snail hosts and on plant carriers

A total of 11 records, describing 15 studies, assessed *Fasciola* spp. infections in snail hosts and on plant carriers. Fourteen studies investigated the prevalence of *Fasciola* spp. in snails in five countries (Table 7), including Vietnam (n = 7), Indonesia (n = 2), Thailand (n = 2), the Philippines (n = 2), and Cambodia (n = 1). The detection of *Fasciola* spp. infections in snails was mainly based on crushing of the snails followed by microscopic examination (n = 11). In two other studies, cercarial shedding was deployed, and in one, dissection. The prevalence of *Fasciola* spp. in snails ranged between 0 and 66.2% by crushing, and between 0.0 and 1.3% by shedding. *Fasciola* spp. were found in unspecified lymnaeid snails in four studies, and in *Austropeplea viridis*/*Lymnaea viridis* [15,104], *Lymnaea swinhoei* [15], and *Lymnaea rubiginosa* [105–107]. Two more studies reported the presence of *Gymnocephalous* cercariae, a cercarial type to which *Fasciola* spp. belong (among others), in *Bithynia siamensis* [108] and *Radix rubiginosa* [106]. There is one study reporting *Fasciola* eggs on Chinese cabbage, thus indicating the presence of fecal matter on this plant [109]. However, none of the retrieved studies reported the presence of *Fasciola* spp. metacercariae, the infectious stage, on plant carriers.

### Risk of bias

All of the 87 unique population surveys underwent risk of bias assessment. For 35 out of these (40.2%), the sample was truly or somewhat representative for the target population. For the seven studies where applicable, only one (14.3%) provided information on non-respondents. Data on risk factors were provided in 26 studies, 25 (96.1%) of which used a validated measurement tool for exposure, and eight (30.8%) controlled for other factors, by means of a multivariable model. A total of 79 out of 87 (90.8%) detailed the method used for outcome assessment. Finally, 56 out of 87 (64.4%) described the statistical methods. Only 17 studies (19.5%) were deemed to be of high quality.

## Discussion

Aligned with the sustainable development goal 3, which focuses on promoting good health and well-being, the WHO has pledged to eliminate the epidemics of diverse infectious diseases, including neglected tropical diseases of zoonotic origin, such as fasciolosis (WHO, 2020). To achieve these targets set by the WHO, it is crucial to have an in-depth understanding about the epidemiology of such diseases in regions where they are prevalent, in order to develop effective control strategies. While Southeast Asia is recognized as an endemic region for fasciolosis [112], a systematic, comprehensive and detailed assessment of the current epidemiological landscape across various hosts was lacking up to now. Thus, this study aimed to systematically review recent literature on the occurrence of and risk factors associated with *Fasciola* spp. in all hosts involved in its life cycle within Southeast Asia.

### Southeast Asia serves as a typical example of *Fasciola* spp. transmission

Our study analyzed population surveys from various sources, revealing prevalence estimates of up to 66.7% in humans and as high as 97.8% in cattle and buffaloes. Although investigated in fewer studies (13 out of 100 studies on the final animal host), the presence of *Fasciola* spp. was also detected in numerous other domestic (e.g., goats and pigs) and wildlife species (e.g., deer, monkeys and elephants) that consume plants in the region. Among the intermediate snail hosts, the prevalence ranged up to 66.2%. For the plant carriers, the absence of studies examining their contamination with *Fasciola* spp. metacercariae was notable, particularly considering their role as the source of infection for humans and animals. Although more studies are crucial in assessing the infection risk, the lack of quick and effective tools for assessing the contamination of plants with *Fasciola* spp. is hampering this investigation. Overall, the reported prevalence estimates for humans surpass those summarized in a similar review centered on Africa, which reported a prevalence range of up to 20.9% [113]. Similarly, prevalence estimates for animal final hosts in Southeast Asia are higher compared to those reported in East and Southern Africa, with estimates ranging up to 58% [114]. To the best of our knowledge, no systematic review has yet been published summarizing prevalence estimates for *Fasciola* spp. in the snail host and/or plant carrier, so comparisons are difficult for these life cycle actors.

Our systematic review also reconfirms the occurrence of human *Fasciola* spp. infection at the country level as previously reported by the WHO [112]. For Laos and the Philippines, however, we did not identify any cases in our review, which may be attributed to the fact that WHO may rely on the reporting of cases prior to 2000 (the earliest year covered in our review) or non-bibliographic government reports. For the animal final hosts, fasciolosis is not listed as a notifiable disease by the World Organization for Animal Health (WOAH) [115]. However, compared to the review article by Mehmood et al. 2017 [2], our study expands the description of *Fasciola* spp. infection occurrence in ruminants beyond just Vietnam and Cambodia to seven countries in Southeast Asia, which is line with the findings in the review of Calvani and Šlapeta 2021 [7]. The same applied for the snail hosts: while Xiao et al. 2018 [116] reported *Lymnaea viridis* and *Radix swinhoei* as potential transmitters of *Fasciola* spp. in Thailand and Vietnam, our systematic review expanded this knowledge by identifying records reporting the presence of *Fasciola* spp. in four snail species across four countries.

### Significant challenges remain to accurately assess the epidemiology of *Fasciola* spp. in Southeast Asia

Despite Southeast Asia being confirmed as an endemic region for *Fasciola* spp., our systematic review also revealed significant challenges in accurately assessing the epidemiology based on

recent literature, primarily due to a dearth of well-designed studies. The risk of bias assessment conducted in our review indicated that less than a fifth of the population survey studies met the high-quality standards. The most commonly observed issues included the lack of information on non-respondents, not controlling for important factors in the analysis, and the lack of representativeness of the surveyed population compared to the national population. Consequently, the reported prevalence estimates may be subject to bias. Notably, certain studies also focused on specific subgroups such as, school children in Cambodia [18] and Vietnam [23], or hospital patients in Vietnam [19,24–26], potentially leading to an overestimation of the true underlying national prevalence. Moreover, there is a significant imbalance in the available literature, with a strong emphasis on specific host types. Among the different host species, the animal final hosts, particularly cattle, received the most attention, accounting for 46.2% (67 out of 145 studies) of the retrieved studies. Furthermore, a wide range of diagnostic tools were utilized in the studies, each with their own inherent limitations in terms of sensitivity and specificity. For instance, the microscopic examination of stool samples from the final human and animal hosts has a notoriously low sensitivity, due to the long prepatent period, low as well as intermittent egg shedding reported for *Fasciola* spp. [117]. The specificity can be hampered too in the presence of eggs of other trematodes such as, *Gastrodiscoides hominis*, *Paragonimus* spp., which are both prevalent in the study area. The Ab-ELISA, the most used technique for the human population surveys in Southeast Asia, on the other hand, is able to detect infections much earlier than copro-microscopy. However, the antigenic targets of the assays are often ill-defined, and the sensitivity and specificity not well characterized [118].

### **Addressing the challenge of *Fasciola* spp. infections in Southeast Asia necessitates the establishment of more ambitious and transdisciplinary partnerships**

Given the zoonotic nature of *Fasciola* spp., it is evident that comprehensive investigations incorporating One Health principles are crucial, especially in the light of more frequent reports on the parasites' resistance to triclabendazole, up to now the drug of choice for mass drug administration campaigns [5]. However, our systematic review revealed a notable gap in literature, as none of the retrieved studies assessed the occurrence of *Fasciola* spp. across all actors involved in its life cycle (including humans, animals, snails, and plant carriers) within the same geographical area. Only four studies in our review combined the assessment of *Fasciola* spp. occurrence in two hosts (cattle and snails [15], buffalo and snail [16], humans and snails [17], and humans and cattle [18]). Moreover, none of the studies retrieved conducted a very thorough investigation of risk factors, such as the investigation of the effect of certain culinary, sanitation and animal management practices, although this is essential to understand the transmission of *Fasciola* spp. Season is another important, yet poorly studied factor, next to the above mentioned designed related factors, explaining the wide variability in prevalence, even within the same country. Indeed, infection with *Fasciola* spp. is thought to be seasonal, as appropriate temperature and humidity levels are needed for egg development and metacercarial survival, and lymnaeid snail population maintenance. Moreover, certain management practices, typical for Southeast Asia, such as allowing animals to graze on rice stubble after harvest might further contribute to the seasonality [7]. Finally, the presence of both *Fasciola* species, *F. hepatica* and *F. gigantica*, as well as their hybrids has been reported in Southeast Asia [7], but their exact distribution is ill-described, and the impact on pathology, transmission, prevention and control in the region are therefore poorly understood. Consequently, there remains a significant lack of scientific evidence necessary for the development of targeted intervention strategies. To effectively combat *Fasciola* spp., it is imperative to foster transdisciplinary

collaborations that encompass all relevant hosts and incorporate a comprehensive exploration of risk factors, enabling the formulation of evidence-based interventions. In addition, the health impact of fasciolosis should be quantified by means of Disability Adjusted Life Years (DALY), an established WHO public health metric widely applied to express the burden of disease [119]. The socio-economic impact assessment should include direct as well as indirect costs related to human (e.g., costs for treatment; costs due to absence from work) and animal fasciolosis (e.g., costs for treatment and economic losses due to reduced meat and milk production and draught power, and condemnation of offal). Such an exercise would help in optimizing the allocation of resources towards the fight against this important disease.

Despite the strengths of our study in terms of conducting a systematic review comprising all the involved hosts in Southeast Asia, it is important to acknowledge certain limitations. Firstly, our language restriction may have resulted in the omission of relevant records published in languages other than English. This potential language bias could have impacted the comprehensiveness of our findings. Additionally, due to significant variations in prevalence estimates arising from differences in diagnostic tests, study designs, quality, and duration, we refrained from performing a meta-analysis to estimate national and regional prevalence of the disease. As is inherent to any systematic review, the estimated prevalence ranges are highly dependent on the quality of the included studies. Consequently, they may not accurately reflect the true prevalence ranges of *Fasciola* spp. infection in Southeast Asia, given the potential use of imperfect tests and limitations in the design of the studies included in our analysis.

## Author Contributions

**Conceptualization:** Vinh Hoang Quang, Bruno Levecke, Dung Do Trung, Brecht Devleesschauwer, Binh Vu Thi Lam, Katja Polman, Steven Callens, Pierre Dorny, Veronique Dermauw.

**Data curation:** Vinh Hoang Quang, Kathy Goossens, Veronique Dermauw.

**Formal analysis:** Vinh Hoang Quang.

**Funding acquisition:** Bruno Levecke, Dung Do Trung.

**Investigation:** Vinh Hoang Quang, Kathy Goossens, Veronique Dermauw.

**Project administration:** Vinh Hoang Quang, Veronique Dermauw.

**Supervision:** Bruno Levecke, Dung Do Trung, Veronique Dermauw.

**Visualization:** Vinh Hoang Quang.

**Writing – original draft:** Vinh Hoang Quang, Bruno Levecke, Pierre Dorny, Veronique Dermauw.

**Writing – review & editing:** Vinh Hoang Quang, Bruno Levecke, Dung Do Trung, Brecht Devleesschauwer, Binh Vu Thi Lam, Kathy Goossens, Katja Polman, Steven Callens, Pierre Dorny, Veronique Dermauw.

## References

1. Mas-Coma S, Bargues MD, Valero MA. Fascioliasis and other plant-borne trematode zoonoses. *International Journal for Parasitology*. 2005; 35(11):1255–78. <https://doi.org/10.1016/j.ijpara.2005.07.010> PMID: 16150452
2. Mehmood K, Zhang H, Sabir AJ, Abbas RZ, Ijaz M, Durrani AZ, et al. A review on epidemiology, global prevalence and economical losses of fasciolosis in ruminants. *Microbial Pathogenesis*. 2017; 109:253–62. <https://doi.org/10.1016/j.micpath.2017.06.006> PMID: 28602837

3. Fürst T, Keiser J, Utzinger J. Global burden of human food-borne trematodiasis: a systematic review and meta-analysis. *The Lancet Infectious Diseases*. 2012; 12(3):210–21. [https://doi.org/10.1016/S1473-3099\(11\)70294-8](https://doi.org/10.1016/S1473-3099(11)70294-8) PMID: 22108757
4. Organization WH. Report of the WHO Informal Meeting on use of triclabendazole in fascioliasis control Geneva, Switzerland: 2007 17–18 October 2006. Report No.: WHO/CDS/NTD/PCT/2007.1.
5. Kelley JM, Elliott TP, Beddoe T, Anderson G, Skuce P, Spithill TW. Current Threat of Triclabendazole Resistance in *Fasciola hepatica*. *Trends in Parasitology*. 2016; 32(6):458–69. <https://doi.org/10.1016/j.pt.2016.03.002> PMID: 27049013
6. Mas-Coma S, Bargues MD, Valero MA. Human fascioliasis infection sources, their diversity, incidence factors, analytical methods and prevention measures. *Parasitology*. 2018; 145(13):1665–99. Epub 2018/07/11. <https://doi.org/10.1017/S0031182018000914> PMID: 29991363
7. Calvani NED, Šlapeta J. *Fasciola gigantica* and *Fasciola* hybrids in Southeast Asia: CABI International; 2021. 423–60 p.
8. Sripa B, Kaewkes S, Intapan PM, Maleewong W, Brindley PJ. Food-borne trematodiasis in Southeast Asia epidemiology, pathology, clinical manifestation and control. *Adv Parasitol*. 2010; 72:305–50. [https://doi.org/10.1016/S0065-308X\(10\)72011-X](https://doi.org/10.1016/S0065-308X(10)72011-X) PMID: 20624536.
9. Mas-Coma S, Valero MA, Bargues MD. Climate change effects on trematodiasis, with emphasis on zoonotic fascioliasis and schistosomiasis. *Vet Parasitol*. 2009; 163(4):264–80. Epub 20090326. <https://doi.org/10.1016/j.vetpar.2009.03.024> PMID: 19375233.
10. Hoang Quang V, Levecke B, Do Trung D, Devleeschauwer B, Lam BVT, Polman K, et al. *Fasciola* spp. in Southeast Asia: a systematic review and meta-analysis protocol. *Systematic Reviews*. 2022; 11(1):138. <https://doi.org/10.1186/s13643-022-02013-3> PMID: 35790977
11. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. *International Journal of Surgery*. 2021; 88:105906. <https://doi.org/10.1016/j.ijsu.2021.105906> PMID: 33789826
12. (2021) RCT. A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>.
13. Patra J, Bhatia M, Suraweera W, Morris SK, Patra C, Gupta PC, et al. Exposure to second-hand smoke and the risk of tuberculosis in children and adults: a systematic review and meta-analysis of 18 observational studies. *PLoS Med*. 2015; 12(6):e1001835; discussion e. Epub 20150602. <https://doi.org/10.1371/journal.pmed.1001835> PMID: 26035557.
14. Wells GA, Wells G, Shea B, Shea B, O'Connell D, Peterson J, et al., editors. The Newcastle-Ottawa Scale (NOS) for Assessing the Quality of Nonrandomised Studies in Meta-Analyses 2014.
15. Nguyen ST, Nguyen DT, Van Nguyen T, Huynh VV, Le DQ, Fukuda Y, et al. Prevalence of *Fasciola* in cattle and of its intermediate host *Lymnaea* snails in central Vietnam. *Tropical Animal Health and Production*. 2012; 44(8):1847–53. <https://doi.org/10.1007/s11250-012-0147-8> PMID: 22549769
16. Portugaliza HP, Balaso IMC, Descallar JCB, Lañada EB. Prevalence, risk factors, and spatial distribution of *Fasciola* in carabao and intermediate host in Baybay, Leyte, Philippines. *Veterinary Parasitology: Regional Studies and Reports*. 2019; 15:100261. <https://doi.org/10.1016/j.vprsr.2018.100261> PMID: 30929938
17. Quy T, Yeatman H, Flood VM, Chuong N, Tuan B. Prevalence and risks of fascioliasis among adult cohorts in Binh Dinh and Quang Ngai provinces-central Viet Nam. 2015.
18. Bless PJ, Schär F, Khieu V, Kramme S, Muth S, Marti H, et al. High prevalence of large trematode eggs in schoolchildren in Cambodia. *Acta Tropica*. 2015; 141(Part B):295–302. <https://doi.org/10.1016/j.actatropica.2014.09.007> PMID: 25250828
19. Van De N. Fascioliasis infection in tumour liver patients in Hanoi hospitals 2006–2010. *Tropical Medicine and International Health*. 2011; 16:274.
20. Ngui R, Ishak S, Chuen CS, Mahmud R, Lim YAL. Prevalence and Risk Factors of Intestinal Parasitism in Rural and Remote West Malaysia. *PLOS Neglected Tropical Diseases*. 2011; 5(3):e974. <https://doi.org/10.1371/journal.pntd.0000974> PMID: 21390157
21. Kitvatanachai S, Taylor A, Rhongbuttri P, Pongstaporn W. Determine the prevalence of intestinal and soil-transmitted helminths using different copromicroscopic techniques in Krabi Province, Thailand. *Asian Pac J Trop Dis*. 2017; 7(12):719–23. <https://doi.org/10.12980/apjtd.7.2017D7-196>
22. Najib MA, Noor-Izani NJ, Wan-Nor-amilah WAW, Wong WK, Faez AM. High seroprevalence of anti-*Fasciola* antibody among cattle farm workers and dwellers in Kelantan. *Tropical Biomedicine*. 2020; 37(2):389–96..
23. Uga S, Hoa NTV, Thuan LK, Noda S, Fujimaki Y. Intestinal parasitic infections in schoolchildren in a suburban area of Hanoi, Vietnam. *Southeast Asian Journal of Tropical Medicine and Public Health*. 2005; 36(6):1407–11.. PMID: 16610642

24. Nguyen TGT, Le TH, De NV, Doan TT, Dao THT, Vercruysse J, et al. Assessment of a 27-kDa antigen in enzyme-linked immunosorbent assay for the diagnosis of fasciolosis in Vietnamese patients. *Tropical Medicine and International Health*. 2010; 15(4):462–7. <https://doi.org/10.1111/j.1365-3156.2010.02468.x> PMID: 20149166
25. Nguyen T, Cheong FW, Liew JWK, Lau YL. Seroprevalence of fascioliasis, toxocariasis, strongyloidiasis and cysticercosis in blood samples diagnosed in Medic Medical Center Laboratory, Ho Chi Minh City, Vietnam in 2012. *Parasites & Vectors*. 2016; 9(1):486. <https://doi.org/10.1186/s13071-016-1780-2> PMID: 27595647
26. De NV, Minh PN, Bich NN, Chai JY. Seroprevalence of Tissue and Luminal Helminths among Patients in Hanoi Medical University Hospital, Vietnam, 2018. *Korean J Parasitol*. 2020; 58(4):387–92. Epub 20200825. <https://doi.org/10.3347/kjp.2020.58.4.387> PMID: 32871632.
27. Tran VH, Tran TK, Nguye HC, Pham HD, Pham TH. Fascioliasis in Vietnam. *The Southeast Asian journal of tropical medicine and public health*. 2001; 32 Suppl 2:48–50. PMID: 12041604.
28. Thu HN, Dermauw V, Huy TT, Roucher C, Dorny P, Thi HN, et al. Diagnosing Human Fascioliasis Using ELISA Immunoassays at a Tertiary Referral Hospital in Hanoi: A Cross-Sectional Study. *Tropical Medicine and Infectious Disease*. 2022; 7(5). <https://doi.org/10.3390/tropicalmed7050076> PMID: 35622703
29. Puastrup S. Fasciola liver abscess: An unrecognized and fatal but curable disease. *Hepatology International*. 2017; 11(1):S581–S2.
30. Pisespongsa P, Thongsawat S, Leerapun A, Praisontarangkul OA. Characteristics of ERCP findings in hepatobiliary fascioliasis. *Gastrointestinal Endoscopy*. 2011; 73(4):AB367. <https://doi.org/10.1016/j.gie.2011.03.806>
31. Naresh G, Gomez PA, Salmah B, Suryati MY. Fasciolosis (liver fluke) of the breast in a male patient: A case report. *The Breast*. 2006; 15(1):103–5. <https://doi.org/10.1016/j.breast.2005.02.009> PMID: 16024249
32. Aroonroch R, Worawichawong S, Nitiyanant P, Kanchanapitak A, Bunyaratvej S. Hepatic fascioliasis due to *Fasciola hepatica*: a two-case report. *J Med Assoc Thai*. 2006; 89(10):1770–74.
33. Kanoksil W, Wattanatrano D, Wilasrusmee C, Mingphrueh S, Bunyaratvej S. Endoscopic removal of one live biliary *Fasciola gigantica*. *JOURNAL-MEDICAL ASSOCIATION OF THAILAND*. 2006; 89(12):2150.
34. Tippawangkosol P, Saingamsook J, Kijdamrongthum P, Thinrungron N, Jariyawat K, Somboon P. Endoscopic management of biliary fascioliasis and identification of *Fasciola* fluke based on mitochondrial DNA: a case report from Thailand. *The Southeast Asian Journal of Tropical Medicine and Public Health*. 2017; 48(3):524–31.
35. LE THI X, Hung NT, Waikagul J. Cutaneous fascioliasis: a case report in Vietnam. *The American journal of tropical medicine and hygiene*. 2005; 72(5):508–9. PMID: 15891121
36. De NV, Le TH, Agramunt VH, Mas-Coma S. Early Postnatal and Preschool-Age Infection by *Fasciola* spp.: Report of Five Cases from Vietnam and Worldwide Review. *Am J Trop Med Hyg*. 2020; 103(4):1578–89. <https://doi.org/10.4269/ajtmh.20-0139> PMID: 32618259.
37. Sothoeun S, Davun H, Copeman B. Abattoir study on *Fasciola gigantica* in Cambodian cattle. *Tropical Animal Health and Production*. 2006; 38(2):113–5. <https://doi.org/10.1007/s11250-006-4302-y> PMID: 17682595
38. Suon S, Hol D, Siek S, McLean M, Copeman B. Seasonal differences in the incidence of infection with *Fasciola gigantica* in cambodian cattle. *Tropical Animal Health and Production*. 2006; 38(1):23–8. <https://doi.org/10.1007/s11250-006-4340-5> PMID: 17405625
39. Young J, Suon S, Leoung V, Kea P, Hout S, Thong S, et al. Parasitic infections of large ruminants in Cambodia. Cattle health, production and trade in Cambodia, ed by Young JR, Rast L, Suon S and Windsor PA, ACIAR Proceedings. 2013;(138):53–8.
40. Dorny P, Stoliaroff V, Charlier J, Meas S, Sorn S, Chea B, et al. Infections with gastrointestinal nematodes, *Fasciola* and *Paramphistomum* in cattle in Cambodia and their association with morbidity parameters. *Veterinary Parasitology*. 2011; 175(3):293–9. <https://doi.org/10.1016/j.vetpar.2010.10.023> PMID: 21071148
41. Loeurng V, Chea B, Tum S, Seng MOM. Challenge and Prevalence of Fasciolosis in Cattle in Pursat Province, Cambodia. *International Journal of Environmental and Rural Development*. 2016; 7(1):70–6. [https://doi.org/10.32115/ijerd.7.1\\_70](https://doi.org/10.32115/ijerd.7.1_70)
42. Ananta SM, Suharno, Hidayat A, Matsubayashi M. Survey on gastrointestinal parasites and detection of *Cryptosporidium* spp. on cattle in West Java, Indonesia. *Asian Pacific Journal of Tropical Medicine*. 2014; 7(3):197–201. [https://doi.org/10.1016/S1995-7645\(14\)60020-1](https://doi.org/10.1016/S1995-7645(14)60020-1) PMID: 24507639

43. Purwaningsih, Widayati I, Nurhayati D, Baaka A. Differences in the Rearing System Toward Bali Cattle Gastrointestinal Helminths Infestation in Prati District, Manokwari Regency, West Papua Province, Indonesia. *KnE Life Sciences*. 2019; 4(11). <https://doi.org/10.18502/cls.v4i11.3872>
44. Sawitri DH, Wardhana AH, Martindah E, Ekawasti F, Dewi DA, Utomo BN, et al. Detections of gastrointestinal parasites, including *Giardia intestinalis* and *Cryptosporidium* spp., in cattle of Banten province, Indonesia. *Journal of Parasitic Diseases*. 2020; 44(1):174–9. <https://doi.org/10.1007/s12639-019-01179-3> PMID: 32174722
45. Rinca KF, Prastowo J, Widodo DP, Nugraheni YR. Trematodiasis occurrence in cattle along the Progo River, Yogyakarta, Indonesia. *Vet World*. 2019; 12(4):593–7. Epub 20190422. <https://doi.org/10.14202/vetworld.2019.593-597> PMID: 31190716.
46. Awaludin A, Nurkholis, Nusantara S. Identify the diversity of helminth parasites in cattle in Jember district (East Java—Indonesia). *IOP Conference Series: Earth and Environmental Science*. 2018; 207(1):012032. <https://doi.org/10.1088/1755-1315/207/1/012032>
47. Kurnianto H, Ramanoon SZ, Aziz NAA, Indarjulianto S. Prevalence, risk factors, and infection intensity of fasciolosis in dairy cattle in Boyolali, Indonesia. *Vet World*. 2022; 15(6):1438–48. Epub 20220612. <https://doi.org/10.14202/vetworld.2022.1438-1448> PMID: 35993064.
48. Nurtjahyani SD, Agustin DS. Comparison of parasite infection degree in cattle (*Bos* sp.) using faecal egg counting method in two East Java regions, Lamongan and Gresik. *Asian Pacific Journal of Tropical Disease*. 2015; 5(8):614–6. [https://doi.org/10.1016/S2222-1808\(15\)60899-4](https://doi.org/10.1016/S2222-1808(15)60899-4)
49. Setiawan H, Kusnoto MuH. The Effect of Breeding Management on the Prevalence and Trematode Infection Degree in calves. *SCOPUS IJPHRD CITATION SCORE*. 2019; 10(9):433.
50. Pratiwi KW, Koesdarto S, Harijani N. Fasciolosis Prevalence on Several cattle Breeds. *SCOPUS IJPHRD CITATION SCORE*. 2019; 10(9):376.
51. Mukhlisi, Sirupang M, Yanuar A, Sayektiningsih T. Wildlife diversity and identification of potential zoonosis around the Sumatran Rhino (*Dicerorhinus Sumatrensis* Harrissoni) sanctuary, East Kalimantan, Indonesia. *IOP Conference Series: Earth and Environmental Science*. 2020; 457(1):012029. <https://doi.org/10.1088/1755-1315/457/1/012029>
52. Hambal M, Ayuni R, Vanda H, Amiruddin A, Athaillah F. Occurrence of *Fasciola gigantica* and *Paramphistomum* spp. Infection in Aceh Cattle. *E3S Web Conf*. 2020; 151:01025.
53. Tanjung M, Thahira D. Endoparasites of cattle raised under intensive and semi-intensive system at Klumpang Kebon Village, North Sumatra. *IOP Conference Series: Earth and Environmental Science*. 2021; 713(1):012057. <https://doi.org/10.1088/1755-1315/713/1/012057>
54. Phomhaksa S, Thamrongyoswittayakul C, Chanlun A, Chanlun S, Somphol N, Yeanpet C. Prevalence of *Fasciola gigantica* infestation in beef cattle at Nongduang slaughterhouse in Vientiane capital of Lao People's Democratic Republic using iELISA approach. *KKU Veterinary Journal*. 2012; 22(2):155–66.
55. Gueguen E, Leperre P, Petit H, Merlin A, Chartier C. A coproscopical study of helminthosis in domestic ruminants (cattle, buffalo, goat) in Khammouane Province (Lao PDR). *Revue de Médecine Vétérinaire*. 2016; 167(11–12):310–5.
56. Windsor PA, Nampanya S, Kinnavong B, Phommasone P, Bush RD, Khounsy S. Do triclabendazole medicated molasses blocks have a role in control of *Fasciola gigantica* in smallholder cattle production in Lao PDR? *Animal Production Science*. 2019; 59(4):787–93. <https://doi.org/10.1071/AN17255>
57. Diyana JNA, Lokman IH, Fazila SHN, Latiffah H, Ibitoye EB, Hazfalinda HN, et al. A Retrospective Study on Bovine Fascioliasis in Veterinary Regional Laboratories in Peninsular Malaysia. *Journal of Parasitology Research*. 2019; 2019:7903682. <https://doi.org/10.1155/2019/7903682> PMID: 31354983
58. Kamaruddin NC, Razali MAI, Busayo IE, Hamzah NH, Idris LH, Md Isa NM. The First Report of Ruminant Fascioliasis in Sabah, East Malaysia. *Journal of Parasitology Research*. 2021; 2021:6691483. <https://doi.org/10.1155/2021/6691483> PMID: 33953962
59. Rita N, Mursyidah AK, Khadijah S. The prevalence of helminthiasis in cattle, Terengganu, Peninsular Malaysia. *Tropical biomedicine*. 2017; 34(2):324–31. PMID: 33593012.
60. Ahmad-Najib M, Wan-Nor-Amilah WAW, Kin WW, Arizam MF, Noor-Izani NJ. Prevalence and Risk Factors of Bovine Fascioliasis in Kelantan, Malaysia: A Cross-Sectional Study. *Trop Life Sci Res*. 2021; 32(2):1–14. Epub 20210629. <https://doi.org/10.21315/tlsr2021.32.2.1> PMID: 34367511.
61. Khadijah S, Mohammad Faiz M, Rita N, Mursyidah AK, Quaza Nizamuddin H. High prevalence fluke infection at four cattle farms located in Kuala Terengganu, Malaysia. *Tropical Life Sciences Research*. 2019; 30(3):59–67. <https://doi.org/10.21315/tlsr2019.30.3.4>
62. Gordon CA, Acosta LP, Gobert GN, Jiz M, Olveda RM, Ross AG, et al. High Prevalence of *Schistosoma japonicum* and *Fasciola gigantica* in Bovines from Northern Samar, the Philippines. *PLOS Neglected Tropical Diseases*. 2015; 9(2):e0003108. <https://doi.org/10.1371/journal.pntd.0003108> PMID: 25643317

63. Besana C, Paller V. Evaluation of Selected Slaughterhouses and Parasites of Slaughtered Livestock in Cotabato Province, Mindanao, Philippines. *Journal of Livestock Science* (ISSN online 2277–6214). 11:67–76.
64. Molina EC, Gonzaga EA, Lumbao LA. Prevalence of Infection with *Fasciola gigantica* and Its Relationship to Carcase and Liver Weights, and Fluke and Egg Counts in Slaughter Cattle and Buffaloes in Southern Mindanao, Philippines. *Tropical Animal Health and Production*. 2005; 37(3):215–21. <https://doi.org/10.1023/b:trop.0000049294.87048.48> PMID: 15747858.
65. Domingo CY, editor Epidemiology of fasciolosis in Nueva Ecija Province, Northern Philippines, including an assessment of ecological factors and recommended control measures 2014: Citeseer.
66. Jittapalapong S, Sangwaranond A, Nimsuphan B, Inpankaew T, Phasuk C, Pinyopanuwat N, et al. Prevalence of gastro-intestinal parasites of dairy cows in Thailand. *Kasetsart Journal, Natural Sciences*. 2011; 45(1):40–5.
67. Thanasuwan S, Piratae S, Tankathok A, Kunthapom S, Daungmawong S. Prevalence of gastrointestinal parasites in cattle, Mueng District, Kalasin province, Thailand. *Kaen Kaset = Khon Kaen Agriculture Journal*. 2019; 47(6):1151–62. <https://doi.org/10.14456/kaj.2019.105>
68. Kaewnoi D, Wiriaprom R, Indoung S, Ngasaman R. Gastrointestinal parasite infections in fighting bulls in South Thailand. *Vet World*. 2020; 13(8):1544–8. Epub 20200810. <https://doi.org/10.14202/vetworld.2020.1544-1548> PMID: 33061225.
69. Japa O, Siriwechviriya P, Prakhammin K. Occurrence of fluke infection in beef cattle around Phayao Lake, Phayao, Thailand. *Vet World*. 2020; 13(2):334–7. Epub 20200220. <https://doi.org/10.14202/vetworld.2020.334-337> PMID: 32255976.
70. Pornsukarom S, Sanguanprasit B, Sangwittayanon K, Chan-On M, Tyobamrung P, Singasa K, et al. Prevalence and analysis of risk factors for gastrointestinal parasites in beef cattle herds in Aranya-prathet district, Sa Kaeo province, Thailand. *Japanese Journal of Veterinary Research*. 2020; 68(2):69–76. <https://doi.org/10.14943/jivr.68.2.69>
71. Thanasuwan S, Tankathok A. Comparison of polymerase chain reaction and microscopy for the detection of *Fasciola* spp. in the fecal matter of domestic bovines in Kalasin Province, Thailand. *Vet World*. 2021; 14(11):2878–82. Epub 20211110. PMID: 35017834.
72. Boonae B, Chimnoi W, Virochasaengaroorn V, Mungmuang T, Jittapalapong S, Nimsuphan B. Prevalence of a liver fluke infection (*Fasciola*) in cows and buffaloes nearby Songkhla Lake. Bangkok: Kasetsart University; 2013. p. O192.
73. Holland WG, Luong TT, Nguyen LA, Do TT, Vercruysse J. The epidemiology of nematode and fluke infections in cattle in the Red River Delta in Vietnam. *Veterinary Parasitology*. 2000; 93(2):141–7. [https://doi.org/10.1016/s0304-4017\(00\)00363-0](https://doi.org/10.1016/s0304-4017(00)00363-0) PMID: 11035232
74. Suzuki K, Kanameda M, Inui K, Ogawa T, Nguyen VK, Dang TTS, et al. A longitudinal study to identify constraints to dairy cattle health and production in rural smallholder communities in Northern Vietnam. *Research in Veterinary Science*. 2006; 81(2):177–84. <https://doi.org/10.1016/j.rvsc.2005.12.002> PMID: 16481015
75. Geurden T, Somers R, Thanh NTG, Vien LV, Nga VT, Giang HH, et al. Parasitic infections in dairy cattle around Hanoi, northern Vietnam. *Veterinary Parasitology*. 2008; 153(3):384–8. <https://doi.org/10.1016/j.vetpar.2008.01.031> PMID: 18328629
76. Nguyen TGT, Le TH, Dao THT, Tran TLH, Praet N, Speybroeck N, et al. Bovine fasciolosis in the human fasciolosis hyperendemic Binh Dinh province in Central Vietnam. *Acta Tropica*. 2011; 117(1):19–22. <https://doi.org/10.1016/j.actatropica.2010.09.003> PMID: 20920452
77. Lan Anh NT, Thanh DTH, Hoan DH, Thuy DT, Khong NV, Anderson N. The transmission of *Fasciola* spp. to cattle and contamination of grazing areas with *Fasciola* eggs in the Red River Delta region of Vietnam. *Tropical Animal Health and Production*. 2014; 46(4):691–6. <https://doi.org/10.1007/s11250-014-0542-4> PMID: 24510222
78. Nguyen NT, Le TC, Vo MDC, H VAN C, Nguyen LT, Ho KT, et al. High prevalence of cattle fascioliasis in coastal areas of Thua Thien Hue province, Vietnam. *J Vet Med Sci*. 2017; 79(6):1035–42. Epub 20170501. <https://doi.org/10.1292/jvms.16-0331> PMID: 28458272.
79. Nurhidayah N, Satrija F, Retnani EB, Astuti DA, Murtini S. Prevalence and risk factors of trematode infection in swamp buffaloes reared under different agro-climatic conditions in Java Island of Indonesia. *Vet World*. 2019; 13(4):687–94. Epub 20200415. <https://doi.org/10.14202/vetworld.2020.687-694> PMID: 32546913.
80. Ishihara S, Ishida A, Barrio D, Lapitan RM, Atabay E, Boyles RM, et al. Field survey on Tamaraw (*Bubalus mindorensis*)—Present population size and herd behavior in wild. *Italian Journal of Animal Science*. 2007; 6(sup2):1249–51. <https://doi.org/10.4081/ijas.2007.s2.1249>

81. Navarro RC, de Cadiz AE, Fronda JM, Ong LAD, Belizario VY Jr. Prevalence of *Schistosoma japonicum* infection in water buffaloes in selected areas in Davao del Norte and Davao de Oro, the Philippines. *International Journal of One Health*. 2021; 7(1):12–9.
82. Mingala CN, Gundran RS. Assessment of water buffalo health and productivity in a communal management system in the Philippines. *Tropical Animal Health and Production*. 2008; 40(1):61–8. <https://doi.org/10.1007/s11250-007-9054-9> PMID: 18551780
83. Silvestre RC, Camalig FM, Silvestre JQ. Prevalence of ectoparasites and endoparasites of carabao in Region I. *E-International Scientific Research Journal*. 2013; 5(3):177–92.
84. Tum S, Puotinen ML, Copeman DB. A geographic information systems model for mapping risk of fasciolosis in cattle and buffaloes in Cambodia. *Veterinary Parasitology*. 2004; 122(2):141–9. <https://doi.org/10.1016/j.vetpar.2004.03.016> PMID: 15177719
85. Tum S, Puotinen ML, Skerratt LF, Chan B, Sothoeun S. Validation of a geographic information system model for mapping the risk of fasciolosis in cattle and buffaloes in Cambodia. *Veterinary Parasitology*. 2007; 143(3):364–7. <https://doi.org/10.1016/j.vetpar.2006.08.033> PMID: 17045401
86. Rast L, Nampanya S, Khounsy S, Toribio J-A, Windsor P. Liver fluke in large ruminants in northern Lao PDR. *Cattle health, production and trade in Cambodia*. 2013; 60.
87. Rast L, Nampanya S, Toribio J-ALML, Khounsy S, Windsor PA. *Fasciola gigantica* infection in large ruminants in northern Laos: smallholder knowledge and practices. *Animal Production Science*. 2017; 57(1):141–6. <https://doi.org/10.1071/AN141032>
88. Mursyidah AK, Khadijah S, Rita N. *Fasciola* and *Paramphistomum* infections in small ruminants (sheep and goat) in Terengganu. *Malaysian Journal of Veterinary Research*. 2017; 8(2):8–12.
89. Padilla M, Ducusin RJT. Herd-level prevalence and factors for risk of endoparasite infection in small-holder pig farms in Sariaya, Quezon, Philippines. *Philippine Journal of Veterinary Medicine*. 2015; 52:99–106.
90. Purnama MTE, Dewi WK, Triana NM, Ooi HK. Serum liver enzyme profile in Timor deer (*Cervus timorensis*) with fascioliasis in Indonesia. *Trop Biomed*. 2021; 38(1):57–61. <https://doi.org/10.47665/tb.38.1.010> PMID: 33797525.
91. Hing S, Othman N, Nathan S, Fox M, Fisher M, Goossens B. First parasitological survey of Endangered Bornean elephants *Elephas maximus borneensis*. *Endangered Species Research*. 2013; 21(3):223–30.
92. Adrus M, Zainuddin R, Ahmad Khairi NH, Ahamad M, Abdullah M-T. Helminth parasites occurrence in wild proboscis monkeys (*Nasalis larvatus*), endemic primates to Borneo Island. *Journal of medical primatology*. 2019; 48(6):357–63. <https://doi.org/10.1111/jmp.12437> PMID: 31486088.
93. Lo CLC, Fernandez DAP, de Luna MCT, de Guia APO, Paller VGV. Diet, parasites, and other pathogens of Sunda leopard cats (*Prionailurus javanensis* Desmarest 1816) in Aborlan, Palawan Island, Philippines. *Journal of Parasitic Diseases*. 2021; 45(3):627–33. <https://doi.org/10.1007/s12639-020-01335-0> PMID: 34475642
94. Loeurng V, Ichikawa-Seki M, Wannasan A, Sothyra T, Chaisowwong W, Tiwananthagorn S. Genetic Characterization of Cambodian *Fasciola gigantica* and Dispersal Direction of the Species in Asia. *Veterinary Parasitology*. 2019; 273:45–51. <https://doi.org/10.1016/j.vetpar.2019.07.013> PMID: 31442892
95. Zalizar L, Rahmawati K, Yaro A. Fasciolosis Infection Level of Various Breed Cattle in Batu and Pujon District, East Java, Indonesia. *E3S Web Conf*. 2021; 226:00029.
96. Estuningsih E, Spithill T, Raadsma H, Law R, Adiwinata G, Meeusen E, et al. Development and application of a fecal antigen diagnostic sandwich ELISA for estimating prevalence of *Fasciola gigantica* in cattle in central Java, Indonesia. *Journal of Parasitology*. 2009; 95(2):450–5. <https://doi.org/10.1645/GE-1672.1> PMID: 18763850
97. Muhammad Rofi P, Setiawan K, Lastuti NDR, Lucia Tri S, Kusnoto, Muchammad Y. Study on the morphology of *Fasciola gigantica* and economic losses due to fasciolosis in Berau, East Kalimantan. *Biosaintifika: Journal of Biology & Biology Education*. 2019; 11(1):156–61. <https://doi.org/10.15294/biosaintifika.v11i1.18201>
98. Calvani NED, Ichikawa-Seki M, Bush RD, Khounsy S, Šlapeta J. Which species is in the faeces at a time of global livestock movements: single nucleotide polymorphism genotyping assays for the differentiation of *Fasciola* spp. *International Journal for Parasitology*. 2020; 50(2):91–101. <https://doi.org/10.1016/j.ijpara.2019.12.002> PMID: 32006549
99. Zainalabidin FA, Azmi MS, Bakri WN, Sathaya G, Ismail MI. Screening for Zoonotic Fascioliasis in Slaughtered Large Ruminants in Abattoirs in Perak, Malaysia. *Trop Life Sci Res*. 2015; 26(2):121–4. PMID: 26868715.
100. Phalee A, Wongsawad C. Prevalence of infection and molecular confirmation by using ITS-2 region of *Fasciola gigantica* found in domestic cattle from Chiang Mai province, Thailand. *Asian Pacific Journal*

- of Tropical Medicine. 2014; 7(3):207–11. [https://doi.org/10.1016/S1995-7645\(14\)60022-5](https://doi.org/10.1016/S1995-7645(14)60022-5) PMID: 24507641
101. Diyana JNA, Mahiza MIN, Latiffah H, Fazila SHN, Lokman IH, Hazfalinda HN, et al. Occurrence, Morphometric, and Molecular Investigation of Cattle and Buffalo Liver Adult Fluke in Peninsular Malaysia Main Abattoirs. *Journal of Parasitology Research*. 2020; 2020:5436846. <https://doi.org/10.1155/2020/5436846> PMID: 32577310
  102. Rosilawati K, Ramli S, Saipul Bahari AR. Fascioliasis in an adult draught buffalo in Malaysia: a case report. *Malaysian Journal of Veterinary Research*. 2017; 8(1):169–72.
  103. Portugaliza HP, Romero SN, Bagot MA. Two potentially zoonotic parasites infecting Philippine brown deer (*Cervus mariannus desmarest*, 1822) in Leyte Island. *Journal of Advanced Veterinary and Animal Research*. 2015; 2(4):489–93.
  104. Dung BT, Doanh PN, The DT, Loan HT, Losson B, Caron Y. Morphological and molecular characterization of lymnaeid snails and their potential role in transmission of *Fasciola* spp. in Vietnam. *Korean J Parasitol*. 2013; 51(6):657–62. Epub 20131231. <https://doi.org/10.3347/kjp.2013.51.6.657> PMID: 24516270.
  105. Suhardono D, Copeman D. Population dynamics of snail *Lymnaea rubiginosa* in rice fields and its infection with larvae of trematodes in the subdistrict of Surade, West Java. *J Ilmu Ternak dan Vet*. 2000; 5:241–9.
  106. Ngoen-klan R, Piangjai S, Somwang P, Moophayak K, Sukontason K, Sukontason KL, et al. Emerging helminths infection in snails and cyprinoid fish in sewage treatment wetlands waters in Cambodia. *Asian Journal of Water, Environment and Pollution*. 2010; 7(3):13–21.
  107. Suhardono Roberts JA, Copeman DB. Biological control of *Fasciola gigantica* with *Echinostoma revolutum*. *Veterinary Parasitology*. 2006; 140(1):166–70. <https://doi.org/10.1016/j.vetpar.2006.02.028> PMID: 16854529
  108. Chontanarath T, Wongsawad C. Epidemiology of cercarial stage of trematodes in freshwater snails from Chiang Mai province, Thailand. *Asian Pacific Journal of Tropical Biomedicine*. 2013; 3(3):237–43. [https://doi.org/10.1016/S2221-1691\(13\)60058-1](https://doi.org/10.1016/S2221-1691(13)60058-1) PMID: 23620846
  109. Chaiphongpachara T, Katsarila P, Wassanasompong W, Ayudhaya WSN, Damapong P, Damapong P. Detection of parasitic contamination in raw vegetables at a local market in Samut Songkhram province, Thailand. *BIOSCIENCE RESEARCH*. 2019; 16(3):2751–5.
  110. Prastowo J, Priyowidodo D, Sahara A, Nurcahyo W, Nugraheni YR, Awaludin A. Molecular identification of cercaria *Fasciola gigantica* in lymnaeid snails in Kulon Progo, Yogyakarta. *Veterinary Parasitology: Regional Studies and Reports*. 2022; 30:100707. <https://doi.org/10.1016/j.vprsr.2022.100707> PMID: 35431065
  111. Martin I GL, Cabrera EC. Morphological Characterization of Emerging Cercariae among Lymnaeid Snails from Barangay Cawongan, Padre Garcia, Batangas, Philippines. *Journal of Parasitology Research*. 2018; 2018:5241217. <https://doi.org/10.1155/2018/5241217> PMID: 30275988
  112. Organization WH. Investing to overcome the global impact of neglected tropical diseases: third WHO report on neglected tropical diseases 2015: World Health Organization; 2015.
  113. Dermauw V, Muchai J, Al Kappany Y, Fajardo Castaneda AL, Dorny P. Human fascioliasis in Africa: A systematic review. *PLOS ONE*. 2021; 16(12):e0261166. <https://doi.org/10.1371/journal.pone.0261166> PMID: 34882738
  114. Malatji MP, Pfukenyi DM, Mukaratirwa S. *Fasciola* species and their vertebrate and snail intermediate hosts in East and Southern Africa: a review. *Journal of Helminthology*. 2020; 94:e63. Epub 2019/07/23. <https://doi.org/10.1017/S0022149X19000531> PMID: 31331410
  115. WOA. OIE-WAHIS Database: World Organisation for Animal Health; 2023. <https://wahis.woah.org/#/dashboards/country-or-disease-dashboard>.
  116. Lu X-T, Gu Q-Y, Limpanont Y, Song L-G, Wu Z-D, Okanurak K, et al. Snail-borne parasitic diseases: an update on global epidemiological distribution, transmission interruption and control methods. *Infectious Diseases of Poverty*. 2018; 7(1):28. <https://doi.org/10.1186/s40249-018-0414-7> PMID: 29628017
  117. Mas-Coma S, Bargues MD, Valero MA. Diagnosis of human fascioliasis by stool and blood techniques: update for the present global scenario. *Parasitology*. 2014; 141(14):1918–46. Epub 2014/07/31. <https://doi.org/10.1017/S0031182014000869> PMID: 25077569
  118. Caravedo MA, Cabada MM. Human Fascioliasis: Current Epidemiological Status and Strategies for Diagnosis, Treatment, and Control. *Research and Reports in Tropical Medicine*. 2020; 11:149–58. <https://doi.org/10.2147/RRTM.S237461> PMID: 33273878
  119. Organization WH. WHO methods and data sources for global burden of disease estimates 2000–2019. World Health Organization: Geneva, Switzerland. 2020.