

BrainPepPass

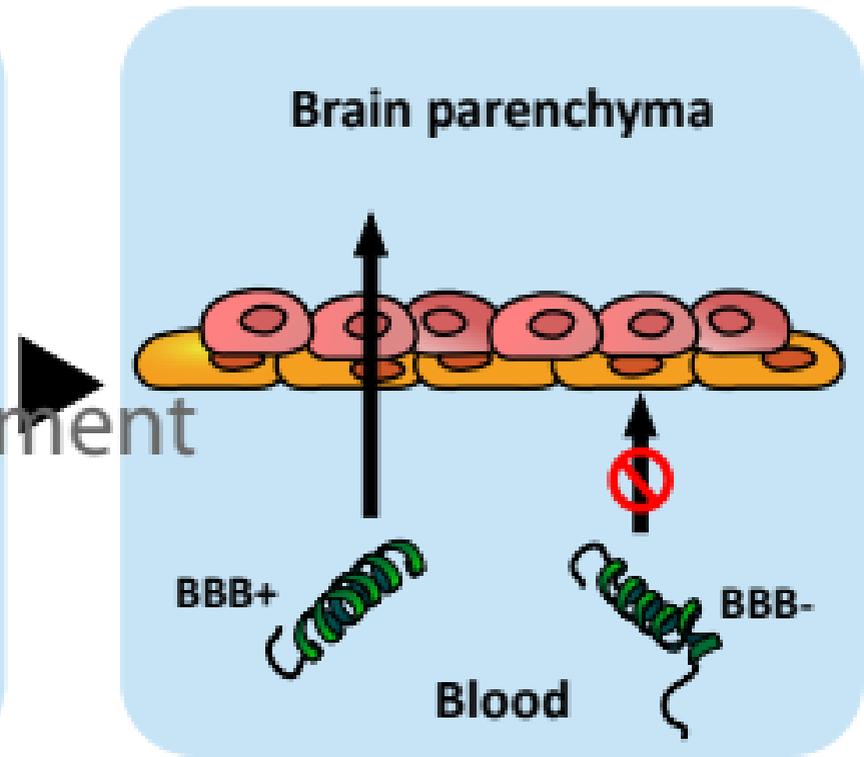
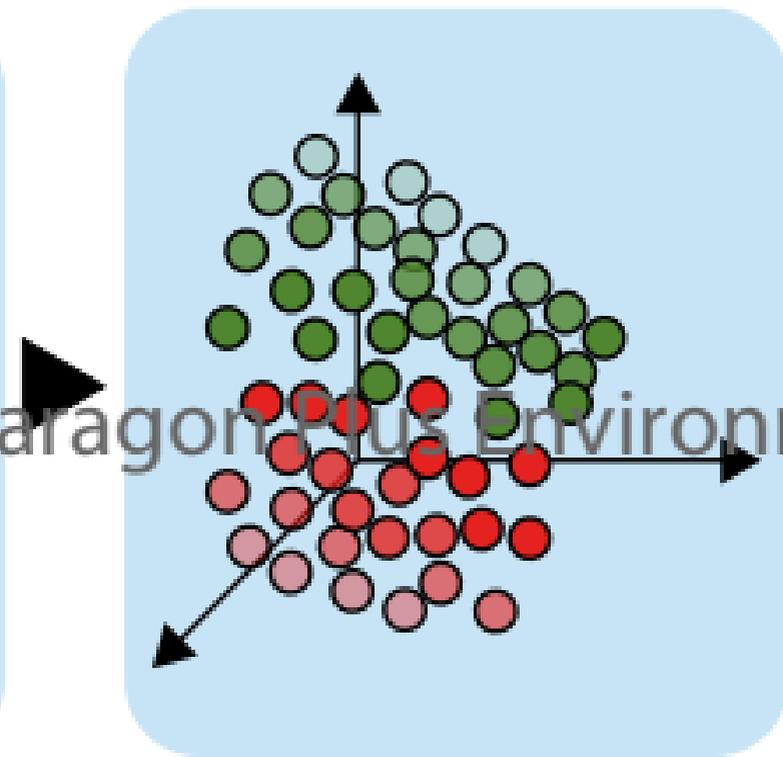
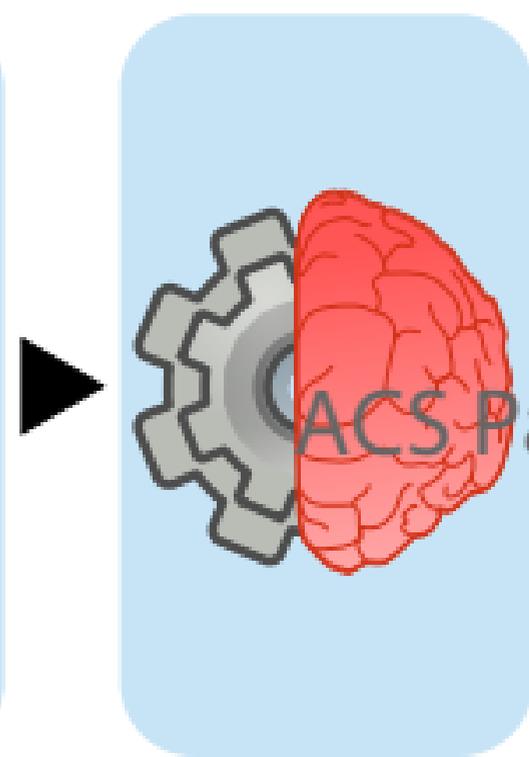
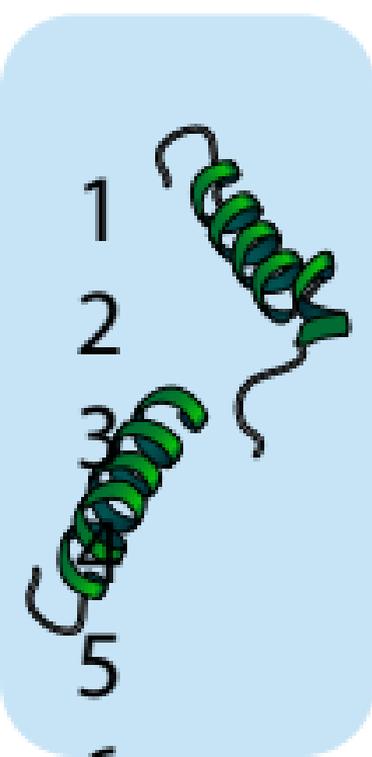
Page 1 of 70 Journal of Chemical Information and Modeling

Query Peptides

Machine Learning

Dimensionality Reduction

Peptide Classification



1
2
3
4
5
6
7 1 BrainPepPass: A Framework Based on Supervised
8
9
10
11 2 Dimensionality Reduction for Predicting Blood-
12
13
14
15 3 Brain Barrier-Penetrating Peptides
16
17
18
19
20

21 4 *Ewerton Cristhian Lima de Oliveira^{1,5*}, Hannah Hirmz^{2*}, Evelien Wynendaele², Juliana*

22
23
24
25 5 *Auzier Seixas Feio¹, Igor Matheus Souza Moreira¹, Kauê Santana da Costa^{3, ‡}, Anderson*

26
27
28
29 6 *H. Lima⁴, Bart De Spiegeleer^{2, ‡}, Claudomiro de Souza de Sales Júnior^{1, ‡}*

30
31
32
33
34 7 ¹Laboratório de Inteligência Computacional e Pesquisa Operacional, Campos Belém,

35
36
37 8 Instituto de Tecnologia, Universidade Federal do Pará, 66075-110, Belém, Pará, Brasil

38
39
40
41 9 ² Drug Quality and Registration (DruQuaR) Group, Faculty of Pharmaceutical Sciences,

42
43
44
45 10 Ghent University, Ottergemsesteenweg 460, B-9000 Ghent, Belgium.

46
47
48
49 11 ³Laboratório de Simulação Computacional, Campos Marechal Rondon, Instituto de

50
51
52 12 Biodiversidade, Universidade Federal do Oeste do Pará, 68040-255, Santarém, Pará, Brasil.
53
54
55
56
57
58
59
60

1
2
3
4 13 ⁴Laboratório de Planejamento e Desenvolvimento de Fármacos, Instituto de Ciências Exatas
5
6
7 14 e Naturais, Universidade Federal do Pará, 66075-110, Belém, Pará, Brasil.
8
9

10
11 15 ⁵Instituto Tecnológico Vale, 66055-090, Belém, Pará, Brasil.
12
13

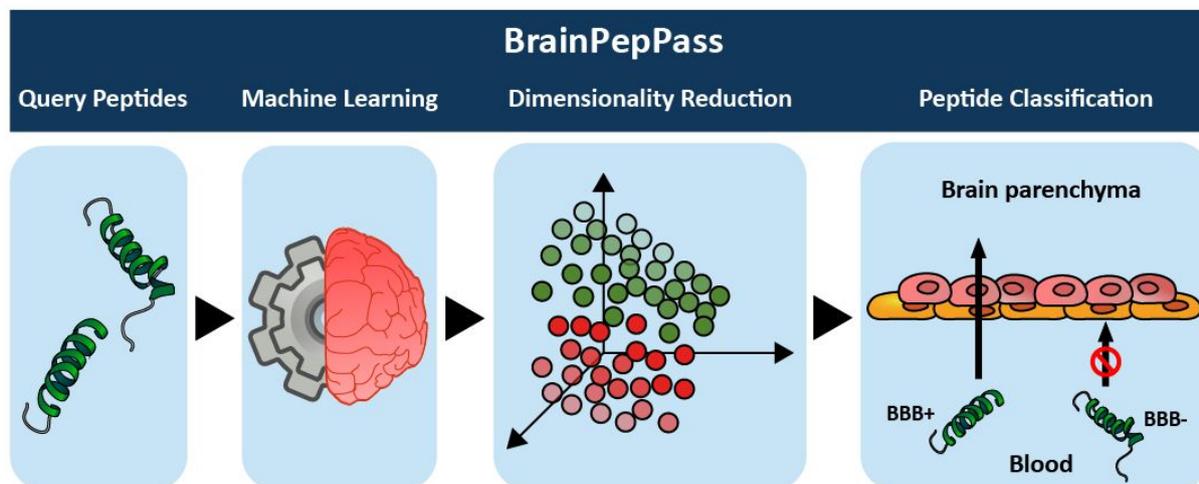
14
15 16 * Authors contributed equally to this work.
16

17 17 ‡ Corresponding authors
18

19
20 18 This paper is dedicated to the memory of a great biomedical scientist, Professor Abba
21 19 Jeremiah Kastin, who passed away last year.
22
23

24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

21 GRAPHICAL ABSTRACT



22

23

24 ABSTRACT

25 Peptides that pass through the blood-brain barrier (BBB) are not only implicated in brain-
26 related pathologies but are also promising therapeutic tools for treating brain diseases, e.g., as
27 shuttles carrying active medicines across the BBB. Computational prediction of BBB-
28 penetrating peptides (B3PPs) has emerged as an interesting approach because of its ability to
29 screen large peptide libraries in a cost-effective manner. In this study, we present
30 BrainPepPass, a machine learning (ML) framework that utilizes supervised manifold
31 dimensionality reduction and extreme gradient boosting (XGB) algorithms to predict natural
32 and chemically modified B3PPs. The results indicate that the proposed tool outperforms other
33 classifiers, with average accuracies exceeding 94% and 98% in 10-fold cross-validation and

1
2
3
4 34 leave-one-out cross-validation (LOOCV), respectively. In addition, accuracy values ranging
5
6
7 35 from 45% to 97.05% were achieved in the independent tests. The BrainPepPass tool is available
8
9
10 36 in a public repository for academic use (<https://github.com/ewerton-cristhian/BrainPepPass>).
11
12
13
14
15 37

16 37

17 38 INTRODUCTION

19
20
21 39 Blood brain-penetrating peptides are oligopeptide chains that can naturally traverse the
22
23
24
25 40 blood-brain barrier (BBB); thus, for example facilitating the enhanced uptake of molecular
26
27
28
29 41 cargoes in a non-selective way. Hence, they are also called BBB shuttle peptides.¹⁻⁴ Until the
30
31
32 42 1970s, peptides were believed not to cross the BBB. The late Abba J Kastin († in 2022) was
33
34
35 43 the first researcher who experimentally tried to refute this assumption. After injecting
36
37
38
39 44 radiolabeled peptides such as ¹²⁵I-Met-enkephalin and ³H- α -melanocyte-stimulating hormone
40
41
42 45 into the carotid artery of mice, Kastin and colleagues observed radioactivity in different brain
43
44
45 46 regions, providing the first indications that certain endogenous peptides cross the BBB.⁵⁻⁷
46
47
48 47 William Banks continued and expanded this research, becoming a protagonist in the field of
49
50
51
52 48 BBB permeability of peptides. Their research shed light on the function of these endogenous
53
54
55 49 peptides as they showed that in crossing the BBB, peptides act as informational molecules that
56
57
58
59
60

1
2
3
4 50 inform the brain of peripheral events. Conversely, peptides crossing from the brain to the blood
5
6
7 51 can deliver information in the brain-to-blood direction.⁸ Not only physiological functions but
8
9
10 52 also pathologies are attributed to the BBB passage of certain peptides. For instance, BBB
11
12
13 53 dysfunction results in amyloid- β disposition in the brain by preventing its normal transport
14
15
16 54 through the BBB. Amyloid plaques formed by amyloid- β aggregation are considered
17
18
19
20 55 pathological triggers of Alzheimer's disease.⁹ Another example is the transport of insulin
21
22
23 56 through the BBB, which is decreased in obese people^{10,11} but seems to be increased in people
24
25
26
27 57 with diabetes mellitus.^{12,13} BBB-penetrating peptides (B3PPs) are being explored in drug
28
29
30 58 development as potential shuttle molecules capable of transporting bio-active drugs across the
31
32
33
34 59 BBB. In addition, some B3PPs may serve as cell-penetrating peptides.¹⁴ Peptides, including
35
36
37 60 the B3PPs, show low immunogenicity and toxicity, and are amenable to chemical synthesis,
38
39
40
41 61 offering a plethora of possibilities for functional modifications and improvements. Therefore,
42
43
44 62 B3PPs have opened up new therapeutic and diagnostic horizons.^{2,4,15}

45
46
47 63 Determining whether and to what extent peptides can cross the BBB is a challenge that
48
49
50
51 64 requires the development of appropriate *in vitro* and *in vivo* techniques to address the technical
52
53
54 65 difficulties in studying these molecules. Various experimental methods have been utilized to
55
56
57 66 determine the permeability of peptides across the BBB, including static *in vitro* models

1
2
3
4 67 encompassing transwell monoculture models, co-culture models, and triple-cell co-culture
5
6
7 68 models. These are straightforward and inexpensive methods that do not capture the
8
9
10 69 physiological complexity involved in BBB permeation. More advanced *in vitro* models such
11
12
13 70 as the blood-brain barrier specific parallel artificial membrane permeability (BBB-PAMPA),
14
15
16 71 bovine brain microvessel endothelial cells (BBMEC), dynamic *in vitro* models, microfluidic
17
18
19
20 72 models, and induced pluripotent stem cells (iPSC)-based models, have been developed to more
21
22
23 73 closely mimic the *in vivo* situation.¹⁶⁻¹⁹ However, these models are expensive and involve
24
25
26 74 complex and rigid procedures, some of which are not well established.^{18,19} Finally, *in vivo*
27
28
29
30 75 experimental methods include the brain uptake index (BUI), multiple time regression (MTR)
31
32
33 76 analysis or Gjedde-Patlak plot, in situ brain perfusion, brain microdialysis, and quantitative
34
35
36 77 radiography. These experimental methods involve more complex, cost- and time-consuming,
37
38
39
40 78 and labor-intensive techniques when compared to computational tools, but provide the most
41
42
43
44 79 complete and detailed quantitative information.^{18,20-22}

45
46 80 With the development of artificial intelligence technology, machine learning (ML) models
47
48
49 81 have been applied in many biochemistry research fields, including protein and protein-like
50
51 82 molecule analysis²³⁻²⁷. For instance, Bao et al. developed several tools covering a wide range
52
53
54 83 of applications, such as Golgi_DF, which classifies Golgi proteins using deep forest
55
56
57
58
59
60

1
2
3 84 algorithms,²⁸ and Phage_UniR_LGBM, which classifies phage virion proteins using UniRep
4
5 85 features and the LightGBM algorithm.²⁴
6
7

8 86 Information on BBB permeability is also often difficult to interpret, because of the multitude
9
10
11 87 of research methods used. Their corresponding output responses ensure that BBB permeability
12
13
14 88 information is not always straightforward to compare, especially in the absence of generally
15
16
17
18 89 agreed controls such as BSA as a negative control and dermorphin as a positive control. To
19
20
21 90 circumvent this problem and allow a direct comparison of BBB influx results, Stalmans et al.
22
23
24 91 introduced a classification method and unified the response of BBB influx transport. The
25
26
27
28 92 results of different BBB influx response types, which quantitatively express brain influx, were
29
30
31 93 classified into five classes of BBB influx magnitude based on the distribution of the results for
32
33
34 94 individual response types. This classification can be immediately applied to new BBB influx
35
36
37
38 95 results of peptides and allows the direct comparison and ranking of peptides independent of the
39
40
41 96 response type.²⁹
42
43
44

45 97 Owing to expensive, time-consuming, and labor-intensive experimental methods, there is an
46
47
48 98 imminent need for efficient *in silico* methods to estimate the BBB permeability of peptides.
49
50
51 99 Several computational methods for estimating the BBB permeability of small molecules
52
53
54
55 100 (excluding peptides) have already been developed.³⁰⁻³⁷ However, for estimating the BBB
56
57
58
59
60

1
2
3
4 101 permeability of peptides, *in silico* methods have only been sparsely investigated.³⁷ Dai et al.
5
6
7 102 presented a sequence-based prediction approach to identify whether a peptide can penetrate the
8
9
10 103 BBB. Using a benchmark dataset, a feature representation learning strategy was designed to
11
12
13 104 characterize sequence-based features from a wide variety of feature descriptors³⁸.
14
15
16
17 105 Subsequently, a three-step feature-selection method was adopted to filter irrelevant and
18
19
20 106 redundant features, resulting in seven optimal features. Based on the optimal features, a
21
22
23
24 107 predictive model was developed using logistic regression (LR).³⁷ Zou employed
25
26
27 108 physicochemical properties of amino acids to represent peptide sequences, and the maximal
28
29
30 109 information coefficient (MIC) and Pearson's correlation coefficient (PCC) were used to extract
31
32
33
34 110 useful information from them. A similarity network fusion algorithm was utilized to integrate
35
36
37 111 these two different types of features, followed by the Fisher algorithm to select the
38
39
40 112 discriminative features. Finally, these selected features were input into support vector machine
41
42
43
44 113 (SVM) to distinguish B3PPs from non-B3PPs.³⁹ However, despite the valuable information
45
46
47 114 provided by these *in silico* methods for predicting peptide penetration of the BBB, the lack of
48
49
50 115 computational tools to predict this pharmacokinetic property for both natural and chemically
51
52
53
54 116 modified peptides hampered the efficient exploration of biotechnological and pharmaceutical
55
56
57 117 applications of these molecules against several brain diseases.
58
59
60

1
2
3
4 118 In this article, we describe BrainPepPass, a novel ML-based framework dedicated to
5
6
7 119 predicting not only whether natural peptides can cross or not the BBB, but also whether
8
9
10 120 chemically modified peptides have this property. To the best of our knowledge, BrainPepPass
11
12
13
14 121 is the first tool in this research field that employs a supervised manifold dimensionality
15
16
17 122 reduction algorithm in the preprocessing stage, in combination with extreme gradient boosting
18
19
20 123 (XGB) models. The recently extended Brainpeps database was used as the most up-to-date and
21
22
23
24 124 complete data source for this study. Moreover, we investigated how distinct groups of
25
26
27 125 molecular descriptors, including physicochemical and structural properties, correlate with the
28
29
30 126 BBB permeability of peptides. In addition, we have provided a repository with the tool
31
32
33
34 127 developed in this study to predict the BBB permeability of peptides.
35
36

37 128

39 129 MATERIAL AND METHODS

43 130 Datasets

47 131 Brainpeps consolidates extensive information related to peptides that interact with and
48
49
50
51 132 penetrate the BBB. The database contains pertinent details of peptides, such as their
52
53
54 133 nomenclature, primary structure, bibliographical references, as well as pharmacokinetic
55
56
57
58
59
60

1
2
3
4 134 indicators. The molecules included in this database were subjected to experimental evaluation,
5
6
7 135 where their BBB penetration characteristics were assessed using a diverse range of methods.⁴⁰
8
9

10 136 A total of 328 peptides from Brainpeps were extracted in the MOL format, which was
11
12
13
14 137 essential for our study because this file format encodes the chemical modifications in the
15
16
17 138 peptide's structure, in addition to encoding the cyclic peptides present in this database. The
18
19
20 139 peptides were classified into two categories: blood-brain barrier permeable (BBB+) or
21
22
23
24 140 nonpermeable (BBB-). This designation was based on experimental indicators used to evaluate
25
26
27 141 the brain penetration levels of the peptides. The six indicators employed in this study included
28
29
30 142 the unidirectional influx constant (K_{in}), measured by multiple time regression (MTR) as well
31
32
33
34 143 as by *in situ* brain perfusion methods; BBB permeability (P), both *in vitro* and *in vivo*;
35
36
37 144 endothelial permeability (P_e) (measured using the parallel artificial membrane permeability
38
39
40 145 assay [PAMPA]); and apparent permeability coefficient (P_{app}).
41
42

43
44 146 The parameter K_{in} or unidirectional influx constant is an indicator that characterizes the
45
46
47 147 steady-state unidirectional influx transfer of peptides from the bloodstream to the brain after a
48
49
50 148 single passage, and is measured in mL/(g.min). This indicator is obtained by performing either
51
52
53
54 149 multiple time regression (MTR) or an *in situ* brain perfusion experiment. After intravenously
55
56
57 150 injecting with a radiolabeled compound, the brain and plasma or perfusate concentrations is
58
59
60

1
2
3
4 151 measured at several time points, which allows the construction of a concentration-time profile,
5
6
7 152 where the slope of this linear regression measures K_{in} .⁴¹ This *in vivo* method has been utilized
8
9
10 153 to classify the penetration of specific peptides such as conotoxin cVc1.1,⁴² or somatropin-
11
12
13 154 derived or modified peptides,⁴³ and quorum-sensing peptides such as PapRIV.⁴⁴

15 155 Conversely, the permeability indicator P , which can be measured using both *in vitro* and *in*
16
17 156 *vivo* techniques, expresses the rate at which a peptide moves from the blood to the brain in
18
19
20 157 units of distance per time, usually cm/s.²⁹ $P_{in vitro}$ data is acquired using the brain microvessel
21
22
23 158 endothelial cell (BMEC) culture model. In this technique, bovine, porcine, mouse, rat, or
24
25
26 159 human BMEC form a monolayer on a rat-tail collagen-coated filter or a microporous membrane
27
28
29 160 placed in a diffusion apparatus consisting of a donor and receptor chamber. The test peptide is
30
31
32 161 placed in the donor compartment and the amount of peptide is measured in samples periodically
33
34
35 162 taken from the acceptor compartment. The amount of test peptide in the acceptor compartment
36
37
38 163 can then be plotted as a function of time to calculate $P_{in vitro}$, which is dependent on the initial
39
40
41 164 concentration of the test peptide in the donor compartment as well as the membrane surface
42
43
44 165 area. $P_{in vivo}$ is measured by performing *in situ* brain perfusion experiments in animal models,
45
46
47 166 such as rats.^{43,45}

1
2
3
4 167 Two other permeability indicators, related to P , determined using slightly different methods
5
6
7 168 and calculations are P_e and P_{app} . P_e evaluates the *in vitro* ability of a molecule to penetrate the
8
9
10 169 endothelial cell layer, which represents the primary barrier to the entry of substances into the
11
12
13 170 brain, and is measured in 10^{-6} cm/s. Di et al. proposed a classification system for molecules
14
15
16
17 171 based on their P_e values, where values greater than $4 \cdot 10^{-6}$ cm/s indicate high penetration, values
18
19
20 172 less than $2 \cdot 10^{-6}$ cm/s indicate low penetration, and values between $2 \cdot 10^{-6}$ and $4 \cdot 10^{-6}$ cm/s
21
22
23 173 indicate uncertain permeation.⁴⁶ These ranges were derived empirically using parallel artificial
24
25
26
27 174 membrane permeation assays (PAMPA). This technique utilizes a porcine polar brain lipid
28
29
30 175 artificial membrane between the donor and acceptor compartments to predict the blood-brain
31
32
33 176 barrier permeation of molecular compounds, including peptides. This technique, in its different
34
35
36
37 177 variants, has been used to classify the permeability of various compounds in the brain,
38
39
40 178 including 3-hydroxy-2-pyridinealldoxime,⁴⁷ furosemide, ranitidine, donepezil, tacrine,⁴⁸
41
42
43 179 platyphyllenone, alnusone,⁴⁹ gingerol, and shogaol derivatives.⁵⁰

44
45
46
47 180 The apparent permeability coefficient (P_{app}) is a similar indicator used to evaluate the *in-vitro*
48
49
50 181 ability of a molecule to traverse a cell-barrier, such as the blood-brain barrier, and is expressed
51
52
53 182 in units of cm/s. *In vitro* models of the BBB, such as monolayers of endothelial cells, are
54
55
56
57 183 commonly employed to evaluate the permeability of compounds using P_{app} .⁵¹ In a similar

1
2
3
4 184 manner to P_c , a classification system for molecules based on their P_{app} values was proposed by
5
6
7 185 Yoon et al., where values greater than $20 \cdot 10^{-6}$ cm/s are considered indicative of high
8
9
10 186 permeability, while values lower than $2 \cdot 10^{-6}$ cm/s suggest low permeability.⁵² Each of these
11
12
13 187 BBB influx response describes the BBB influx from a different viewpoint using different
14
15
16
17 188 techniques, thereby providing different information.

18
19
20 189 As previously discussed, studies investigating the entry of peptides and other compounds
21
22
23
24 190 into the BBB have employed established limits on physicochemical indicators to determine
25
26
27 191 their permeability. Specifically, we propose to categorize peptides as belonging to either the
28
29
30 192 BBB+ or BBB- class based on the limits described for each indicator. For P_c and P_{app} , high
31
32
33
34 193 penetration was designated as belonging to the BBB+ class, whereas low penetration was
35
36
37 194 classified as BBB-. Peptides with penetration rates between low and high were classified
38
39
40 195 according to the proximity of their respective values to one of the two thresholds (high or low).
41
42
43
44 196 For example, a peptide with a penetration level numerically closer to a low threshold was
45
46
47 197 classified as BBB-. In terms of K_{in} and P , we dichotomously classified peptides based on the
48
49
50 198 five groups proposed by Stalmans et al. to evaluate the influx of peptides across the BBB.²⁹
51
52
53
54 199 Peptides belonging to the very low and low influx categories were classified as BBB- class,
55
56
57 200 while those in the medium, high, and very high influx categories were classified as BBB+. The
58
59
60

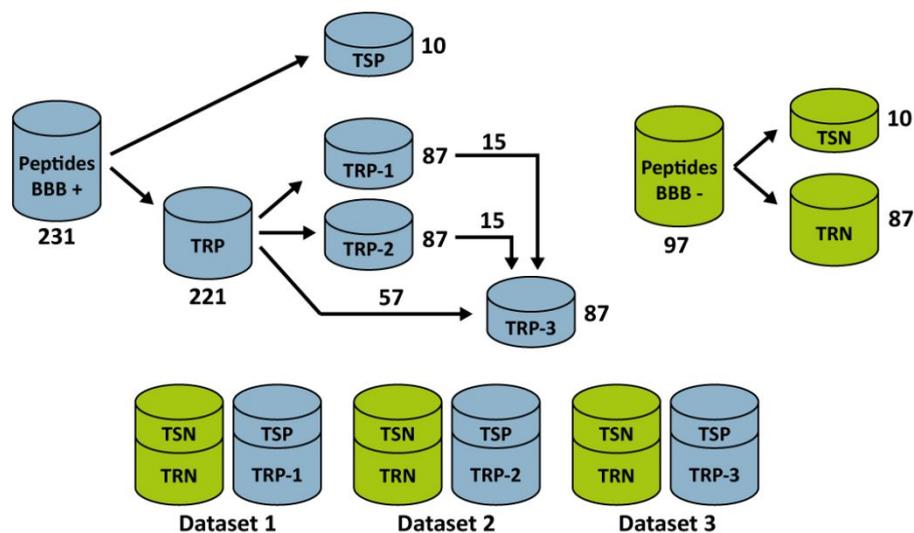
1
2
3
4 201 proposed classification system is expected to facilitate a more direct understanding of BBB
5
6
7 202 permeability and contribute to the development of more effective permeability prediction
8
9
10 203 strategies.

11
12
13 204 The implementation of the proposed criteria for classifying peptides based on their
14
15
16
17 205 permeability in the BBB using physicochemical indicators in our study yielded a database
18
19
20 206 containing 231 BBB+ and 97 BBB- peptides. However, the database was unbalanced in terms
21
22
23
24 207 of the number of peptides belonging to each class, necessitating
25
26
27 208 the division of the data into three balanced datasets to avoid issues associated with overfitting.
28
29
30 209 Each dataset comprised the same 97 BBB- peptides and 97 randomly sampled BBB+ peptides.

31
32
33 210 Of the 231 peptides classified as BBB+ in the complete database, 10 were randomly selected
34
35
36
37 211 to constitute the test sample of permeable molecules (TSP), whereas the remaining 221
38
39
40 212 peptides were used to compose the training samples of permeable molecules (TRP). The BBB+
41
42
43
44 213 peptides in Dataset 1 were composed of 87 molecules randomly sampled from TRP (TRP-1)
45
46
47 214 and 10 peptides from TSP, totaling 97 BBB+ peptides. Similarly, the BBB+ peptides in Dataset
48
49
50 215 2 were a combination of distinct 87 molecules randomly sampled from the TRP (TRP-2) and
51
52
53
54 216 10 peptides from the TSP. The training samples of BBB+ peptides in Dataset 3 (TRP-3) were
55
56
57 217 composed of the remaining 57 samples from TRP with 15 randomly selected peptides from
58
59
60

218 TRP-1 and 15 peptides randomly selected from TRP-2, while the test samples include the same
 219 molecules from TSP, totaling 97 BBB+ peptides (see Figure 1).

220



221

222 **Figure 1.** Construction of the three balanced datasets. TRP-[1,2,3]: training samples of BBB+
 223 peptides, TSP: test samples of BBB+ peptides, TRN: training samples of BBB- peptides, TSN:
 224 test samples of BBB- peptides.

225

226 Overall, each of the three datasets comprised a balanced number of samples from both
 227 classes, with 97 BBB+ and 97 BBB- peptides. In each dataset, 174 peptides (87 BBB+ and 87
 228 BBB-) were dedicated to training, while 20 peptides (10 BBB+ and 10 BBB-) were used for
 229 testing. This approach ensured that each dataset could be utilized for both training and testing

1
2
3
4 230 and that the models developed using these datasets were adequately validated. Supplementary

5
6
7 231 Table S1 provides information regarding the peptides used in each dataset.

8
9
10 232

11
12 233 Molecular Properties

13
14
15
16 234 In this study, we investigated the permeability of peptides across the blood-brain barrier by

17
18
19
20 235 analyzing a set of molecular properties. The molecular properties were grouped into four

21
22
23 236 distinct feature compositions (FCs). The first feature composition (FC-1) comprised several

24
25
26 237 key descriptors including molecular weight (MW), calculated water-octanol partition

27
28
29
30 238 coefficient (LogP), calculated octanol-water distribution coefficient (LogD) at pH 7.4,

31
32
33 239 topological polar surface area (TPSA), number of hydrogen bond acceptors (HBA), donors

34
35
36 240 (HBD), nitrogen count (nN), oxygen count (nO), and nitrogen plus oxygen count (nN+nO).

37
38
39
40 241 Previous studies have highlighted the importance of these descriptors in filtering molecules

41
42
43 242 that are likely to reach the central nervous system (CNS).^{1,53-55} Furthermore, some of these

44
45
46 243 descriptors have also been linked to the oral bioavailability of compounds, as proposed by

47
48
49
50 244 Lipinski's rule of five and other related studies on bioavailability and biomembrane

51
52
53 245 permeability.⁵⁶⁻⁵⁹

1
2
3
4 246 The second feature composition (FC-2) comprised Mordred's molecular descriptors, which
5
6
7 247 consist of a combination of structural and physicochemical descriptors. Mordred is a Python
8
9
10 248 library for molecular descriptor calculations that encompasses 2D, 3D, constitutional, and
11
12
13 249 electronic descriptors.⁶⁰ The 749 descriptors in this FC were extracted from the 231 molecules
14
15
16
17 250 using this package, after filtering out molecular properties with missing, non-numeric, or non-
18
19
20 251 Boolean values.

22
23 252 The third feature composition (FC-3) was constructed by selecting the ten best-correlated
24
25
26
27 253 molecular descriptors from FC-2 using Kendall's correlation coefficient. The fourth
28
29
30 254 feature composition (FC-4) was obtained by combining FC-1 and FC-3. Supplementary Tables
31
32
33
34 255 S2, S3, and S4 provide information on the molecular descriptors of FC-1, FC-2, and FC-3.

35
36
37 256 To calculate the FC-1 descriptors, we utilized the RDKit package in Python to extract the
38
39
40 257 properties from the peptides, except for LogD, which was determined
41
42
43
44 258 for each molecule using the Instant JChem software. The other descriptors were calculated
45
46
47 259 using the Mordred package.

50 260

52 261 Proposed Machine Learning Framework

56 262

1
2
3
4 263 The ML-based framework proposed in this study for predicting B3PPs, BrainPepPass, is a
5
6
7 264 generic architecture, comprising two stages: dimensionality reduction pattern learning (DRPL)
8
9
10 265 and classification. The DRPL step involves projecting a high-dimensional dataset of molecular
11
12
13 266 descriptors onto a three-dimensional (3D) space, with the dual objective of facilitating low-
14
15
16
17 267 dimensional visualization of peptides and enabling the clustering of molecules based on their
18
19
20 268 BBB+ or BBB- class labels. To this end, we employed a supervised Laplacian eigenmaps (sLE)
21
22
23
24 269 algorithm, which has been demonstrated to be effective in reducing high-dimensional data with
25
26
27 270 class labels.⁶¹ However, the original sLE algorithm, like t-SNE, lacks the capacity for
28
29
30 271 independent dataset dimensionality reduction, which renders it unsuitable for BrainPepPass
31
32
33
34 272 prediction of the permeability of new peptides. To overcome this limitation, we propose using
35
36
37 273 an XGB regression (XGBr) algorithm.

40 274 As shown in Figure 2a, the DRPL stage entails the dimensionality reduction of a high-
41
42
43
44 275 dimensional dataset consisting of molecular descriptors by FC to a 3D representation.
45
46
47 276 Specifically, the same original n-dimensional data was used as input, and their 3D projection
48
49
50 277 was used as the target to train an XGBr algorithm to learn the DR pattern performed by sLE.
51
52
53
54 278 To select the optimal hyperparameters for the XGBr, we performed a grid search with a
55
56
57 279 predetermined range of values and utilized a 10-fold cross-validation technique to compute the
58
59
60

1
2
3
4 280 average accuracy metric. Supplementary Table S5 presents the search range and best
5
6
7 281 hyperparameters obtained through this process.
8
9

10 282 The subsequent stage of our ML-based framework involved the classification of molecules
11
12
13 283 using an XGB classifier (XGBc). As illustrated in Figure 2b, the training of the XGBc model
14
15
16
17 284 leveraged the 3D data generated by the DRPL stage as input, with the target being the class
18
19
20 285 labels of the peptides (BBB+ or BBB-). To optimize the performance of XGBc, we performed
21
22
23
24 286 a grid search for the optimal set of hyperparameters, similar to the approach used for XGBr.
25
26
27 287 Supplementary Table S6 presents the search range and best hyperparameters obtained through
28
29
30 288 this process.
31
32

33
34 289 To summarize, the ML-based framework proposed in this study for predicting peptide
35
36
37 290 penetration across the BBB consists of a pipeline comprising the XGBr and XGBc algorithms.
38
39
40 291 The former algorithm was trained to learn the DR pattern produced by the sLE algorithm,
41
42
43
44 292 whereas the latter was responsible for predicting whether a given peptide could penetrate the
45
46
47 293 BBB, as shown in Figure 2c. Importantly, BrainPepPass also facilitates 3D visualization of
48
49
50 294 new data, which is a key feature for analyzing the extent of separation of a given peptide from
51
52
53
54 295 its original cluster.
55
56
57
58
59
60

296 The methodology proposed in this study is summarized in the flowchart shown in Figure 3.

297 Figure 3a illustrates the data extraction step from Brainpeps, where peptides are classified into

298 BBB+ and BBB- based on physicochemical indicators, the three datasets are structured, and

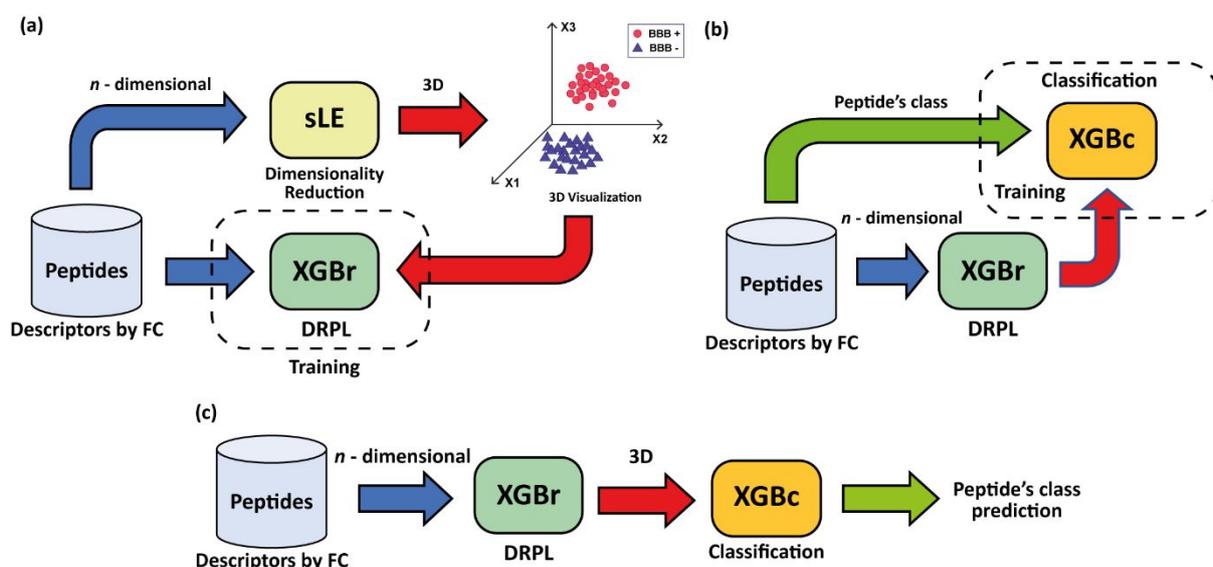
299 the molecular descriptors are calculated. Figure 3b shows the preprocessing step in which the

300 FCs are constructed for each dataset. Figure 3c shows the final step, in which the data are

301 partitioned into training and testing sets, and each of the algorithms analyzed in this study is

302 trained and tested for their respective comparisons using various metrics.

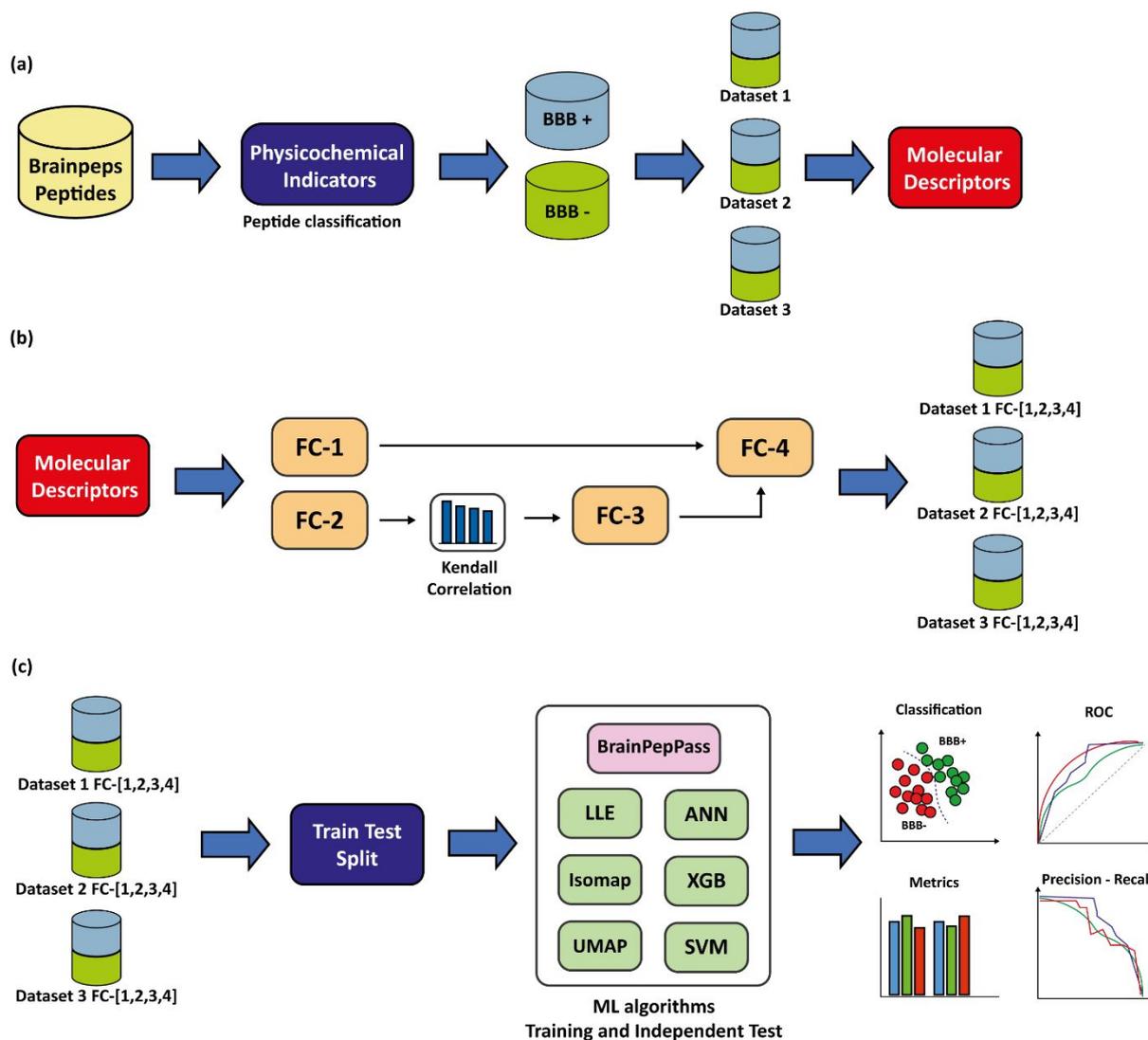
303



304

305 **Figure 2.** Stages of the BrainPepPass. (a) DRPL stage. (b) Training of the XGBc for predicting

306 peptides permeability. (c) Final architecture of the BrainPepPass.



307
308 **Figure 3.** Flowchart of the proposed method to predict B3PPs. (a) Data extraction and
309 molecular properties calculation. (b) Data preprocessing to generate the FCs. (c) B3PPs
310 prediction with BrainPepPass and comparison with other algorithms.

311
312 **RESULTS AND DISCUSSION**

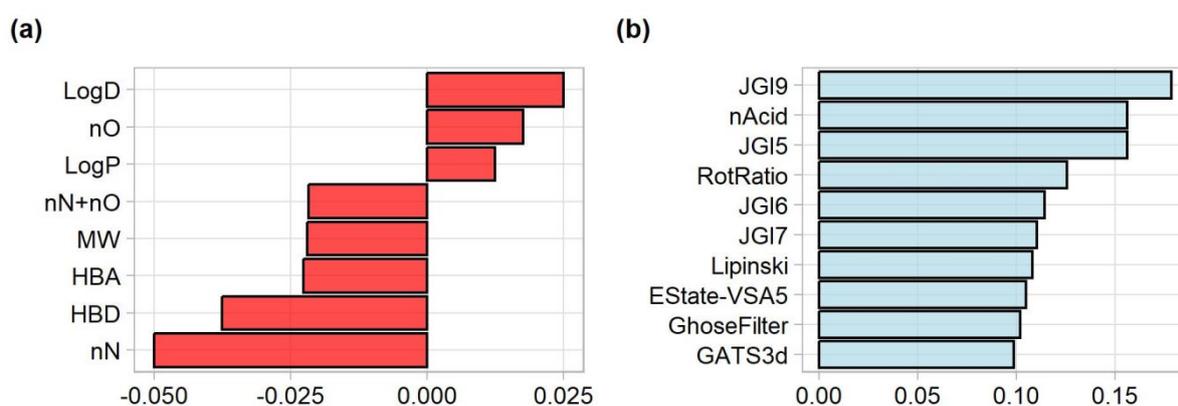
313

1
2
3
4 314 We conducted a thorough analysis using BrainPepPass to predict the permeability of peptides
5
6
7 315 across the BBB by considering multiple factors. First, we performed a Kendall correlation
8
9
10 316 analysis to examine the relationship between the selected structural and physicochemical
11
12
13
14 317 properties of the peptides and their corresponding permeability classes for each FC. This
15
16
17 318 analysis provided insights into the behavior of previously studied permeability properties in
18
19
20 319 our dataset and helped identify the most relevant descriptors to compose FC-3 and FC-4. Thus,
21
22
23
24 320 the present study evaluated the performance of the proposed ML-based framework trained with
25
26
27 321 three datasets for each FC using 10-fold cross-validation and independent testing. Additionally,
28
29
30 322 leave-one-out cross-validation (LOOCV) was used to investigate the predictive generalization
31
32
33
34 323 of BrainPepPass. Moreover, the proposed method was compared with state-of-the-art
35
36
37 324 classifiers previously used in the same research field, such as artificial neural network (ANN),
38
39
40 325 SVM, and XGB. We also compared BrainPepPass using the same ML architecture as shown
41
42
43
44 326 in Figure 2 but with other manifold DR algorithms such as locally linear embedding (LLE),
45
46
47 327 isometric mapping (Isomap), and uniform manifold approximation and projection (UMAP).
48
49
50 328 These comparisons assessed the predictive power and information content of the molecular
51
52
53
54 329 descriptors in discriminating permeability classes of peptides.

330

331 Correlation Analysis and Feature Selection

332 We employed the Kendall correlation to evaluate the association between the molecular
 333 descriptors investigated in FC-1 (see Figure 4a) and all 749 descriptors calculated in FC-2 and
 334 their corresponding permeability classes. Based on these results, we identified the top ten
 335 descriptors with the highest correlation values to form FC-3 (see Figure 4b).



336
 337 **Figure 4.** Kendall correlation analysis on molecular descriptors regarding to permeability
 338 across the BBB. (a) Molecular descriptors previously reported as associated with the
 339 permeability of small molecules across the BBB (FC-1). (b) 10 most correlated Mordred's
 340 descriptors (FC-3).

341
 342 The findings depicted in Figure 4 highlight some aspects of the relationship between the
 343 molecular properties and permeability of peptides across the BBB. Specifically, the properties

1
2
3
4 344 included in FC-1, which have been previously reported to exhibit the highest correlation with
5
6
7 345 the permeability of this biological barrier for small molecules, displayed comparatively lower
8
9
10 346 correlation values in the Kendall correlation analysis than other structural and physicochemical
11
12
13
14 347 properties derived from Mordred.

15
16
17 348 Figure 4b shows some properties related to the electrical properties of the investigated
18
19
20 349 molecules. For example, the charge index (JGI_x) is a topological descriptor that characterizes
21
22
23 350 the molecular charge distribution on the x -th order.⁶² Notably, values of 0.156, 0.114, 0.11, and
24
25
26
27 351 0.178 were obtained for JGI_5 , JGI_6 , JGI_7 , and JGI_9 , respectively. In addition, Estate-VSA
28
29
30 352 calculates the sum of the van der Waals surface area contributions to the electron topological
31
32
33
34 353 states within a specific range.⁶³ A value of 0.104 was observed for *Estate-VSA5*. The Geary
35
36
37 354 coefficient ($GATSy_d$) is a general index of 2D-autocorrelation with lag y applied to a molecular
38
39
40 355 graph.⁶⁴ It describes the topology of a molecule associated with atomic masses, polarizabilities,
41
42
43
44 356 and Sanderson electronegativities, weighted by sigma electrons. The descriptor *GATS3d*
45
46
47 357 achieved a correlation value of 0.098.

48
49
50 358 Although no study has focused on the relationship between JGI_x and BBB penetration,
51
52
53
54 359 studies based on lipid bilayer membrane models to evaluate this pharmacokinetic property of
55
56
57 360 compounds have revealed that charged molecules can modify the dipole potential of the
58
59
60

1
2
3
4 361 membrane through electrostatic interactions and interact with the BBB through attraction and
5
6
7 362 repulsion.⁵⁵ Similarly, estate-VSA descriptors have not been investigated in depth regarding
8
9
10 363 their association with the penetration of this barrier. However, Liu et al. concluded that high
11
12
13 364 van der Waals surface area values are associated with good permeability of molecules in the
14
15
16
17 365 BBB because molecules with high values tend to protonate and carry positive charges in
18
19
20 366 molecules.⁶⁵ Similarly, the correlation between the Geary coefficient and BBB penetration has
21
22
23
24 367 not yet been investigated in previous studies.

25
26
27 368 Additionally, the correlations of other selected features related to the structural properties of
28
29
30 369 the molecules were investigated. For instance, *RotRatio*, which represents the ratio of the
31
32
33
34 370 number of rotatable bonds to the total number of bonds in the molecule, showed a correlation
35
36
37 371 value of 0.125. Similarly, the *nAcid* property, representing the number of acidic groups,
38
39
40 372 showed a correlation value of 0.156. Some studies have indicated that the smaller the number
41
42
43
44 373 of rotational bonds, that is, five or fewer bonds, the greater the permeability of the molecule in
45
46
47 374 the CNS.⁵⁴ However, the number of acidic groups was not directly evaluated as a parameter
48
49
50
51 375 to filter molecules that could penetrate the BBB, although Dichiara et al. observed in their
52
53
54 376 studies that acidic compounds are among the least permeable across the barrier.⁵³
55
56
57
58
59
60

1
2
3
4 377 Several of the descriptors selected for this analysis were linked to the drug-likeness of the
5
6
7 378 molecules and other molecular properties. For example, the descriptor *Lipinski* represents a
8
9
10 379 logical feature based on Lipinski's rule of five, which determines whether a molecule can be
11
12
13
14 380 considered an orally available drug by satisfying specific numerical criteria for MW, LogP,
15
16
17 381 HBA, and HBD.⁵⁷ Another cheminformatics filter, *GhoseFilter*, defines drug-likeness
18
19
20 382 constraints for molecules based on their LogP and MW values, total number of atoms, and
21
22
23
24 383 molar refractivity.⁶⁶ The correlation values for *Lipinski* and *GhoseFilter* were found to be 0.108
25
26
27 384 and 0.101, respectively.

28
29
30 385 The molecular descriptors present in FC-1 have a long history of investigation regarding their
31
32
33
34 386 correlation with the pharmacokinetic property of small molecules in crossing the BBB.
35
36
37 387 However, our results indicated that the correlation values for these properties were
38
39
40
41 388 approximately two to four times lower than those for *GATS3d*, which had the lowest
42
43
44 389 correlation value in FC-3. This finding suggests that the descriptive properties of charge
45
46
47 390 distribution in peptides are closely related to their ability to penetrate the BBB. Notably, our
48
49
50
51 391 analysis also demonstrated that *Lipinski* and *GhoseFilter* descriptors are also significantly
52
53
54 392 associated with peptide penetration of the BBB.
55
56
57
58
59
60

1
2
3
4 393 The results of this investigation also contribute to refining the selection of descriptors for
5
6
7 394 FC-4, which combines the properties of FC-1 and FC-3. FC-4 is essential for evaluating
8
9
10 395 whether combining these descriptors will provide information gain to correctly classify
11
12
13
14 396 molecules that cross the BBB.

15
16
17 397

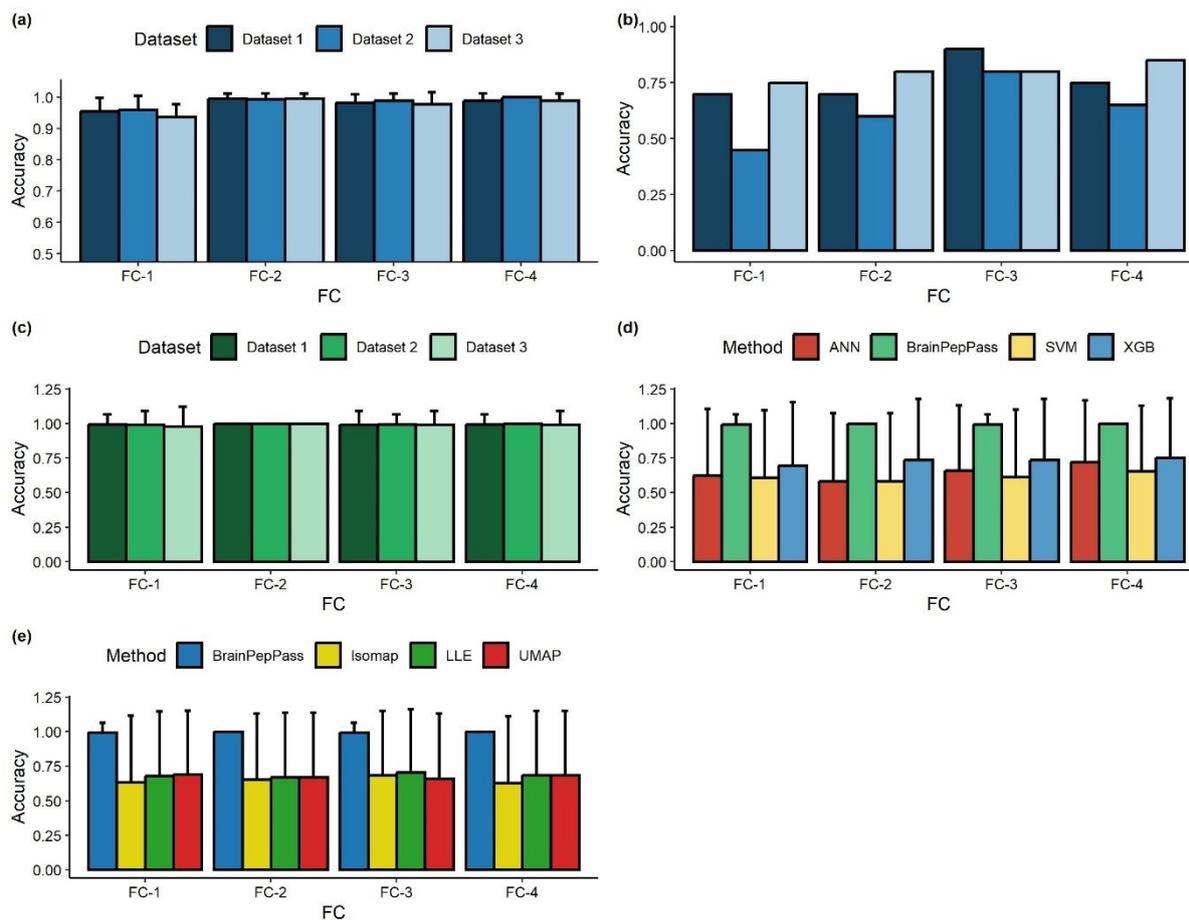
20 398 Cross-validation and Independent Test Analysis

21
22
23
24 399 We evaluated the predictive capacity of BrainPepPass based on the accuracy of 10-fold cross-
25
26
27
28 400 validation using the training portion of each dataset. This metric was applied to three dataset
29
30
31 401 samples and four FCs, and 72 simulations were performed for different values of the sLE
32
33
34 402 gamma parameter: 0.01, 0.02, 0.05, 0.1, 0.2, and 0.5. The best models were selected based on
35
36
37
38 403 the highest accuracy values in the cross-validation for a fixed gamma, which was determined
39
40
41 404 by filtering among all simulations (Figure 5a). The results demonstrate the contribution of each
42
43
44
45 405 group of descriptors in predicting B3PPs using the proposed ML-based framework.
46
47
48 406 BrainPepPass achieved values greater than 93% of average accuracy for all FCs. FC-1
49
50
51 407 exhibited the worst performance, with average accuracy values between 93.6% and 96%,
52
53
54
55 408 whereas FC-2, which comprised the largest number of features, achieved an accuracy of 99.4%
56
57
58 409 for the three datasets. FC-3 obtained values between 97.68% and 98.86%, whereas the FC-4

merged both FC-1 and FC-3 descriptors, obtaining accuracy values ranging from 98.8% to

100%.

412



413

414 **Figure 5.** Accuracy achieved by BrainPepPass in each FC. (a) 10-fold cross-validation. (b)

415 Independent test. (c) LOOCV. (d) LOOCV comparison with ANN, SVM, and XGB models.

416 (e) LOOCV comparison with frameworks based on LLE, Isomap, and UMAP manifold

417 algorithms.

1
2
3
4 418 The ANOVA test was applied to the accuracy values obtained for each fold of the 10-fold
5
6
7 419 cross-validation performed on the three datasets, 1, 2, and 3, FC-2 and FC-4. The ANOVA test
8
9
10 420 showed no statistically significant difference between the three datasets, yielding p -values of
11
12
13 421 0.526, 0.331, and 0.541 for datasets 1, 2, and 3, respectively. However, from a computational
14
15
16
17 422 perspective, a significant difference exists between the models, as BrainPepPass trained with
18
19
20 423 FC-2 requires the calculation of 749 descriptors, whereas the framework based on FC-4
21
22
23
24 424 requires only 19.

25
26
27 425 We also examined the predictive performance of our ML-based framework by applying
28
29
30 426 external validation on peptides that were not part of the cross-validation analysis (Figure 5b).
31
32
33
34 427 The accuracy outcomes obtained by the proposed tool for each feature composition indicate
35
36
37 428 that the feature distribution between the training and test data in each of the three datasets may
38
39
40 429 have been different. This is particularly evident when the performances of FC-2 and FC-4 are
41
42
43
44 430 compared with the performance of FC-3. The ten descriptors selected from Mordred
45
46
47 431 demonstrated superior predictive performance, achieving values ranging from 80% to 90% in
48
49
50 432 predicting which peptides can penetrate the BBB. The FC-4 model achieved an accuracy of
51
52
53
54 433 85% for one of the datasets. Other performance metrics for the best BrainPepPass models by
55
56
57 434 FC were also calculated, as shown in Table 1. The FC-3 model also yielded high F1-score and
58
59
60

1
 2
 3
 4 435 Matthew's correlation coefficient (MCC) values, along with the maximum recall value for one
 5
 6
 7 436 of the datasets. The area under the receiver operating characteristic curve (ROC-AUC) values
 8
 9
 10 437 between 0.74 and 0.84 also indicate that BrainPepPass has a good ability to distinguish between
 11
 12
 13 438 the two classes (BBB+ and BBB-). These results indicate that framework can accurately predict
 14
 15
 16
 17 439 which peptides can penetrate the BBB among all relevant instances using as much the selected
 18
 19
 20 440 molecular descriptors grouped in FC-3, as the properties included in FC-4, which maintained
 21
 22
 23
 24 441 similar performance. Supplementary Table S7 provides the metric values obtained by
 25
 26
 27 442 BrainPepPass and their respective gamma values using the three datasets in the 10-fold cross-
 28
 29
 30 443 validation and independent tests, respectively.
 31
 32
 33
 34 444
 35
 36
 37 445 **Table 1.** Independent test analysis of the best BrainPepPass models by FC.

FC	Dataset	Accuracy	F1 Score	MCC	Precision	Recall	ROC-AUC
FC-1	Dataset 3	0.75	0.71	0.52	0.86	0.60	0.74
FC-2	Dataset 3	0.80	0.82	0.61	0.75	0.90	0.75
FC-3	Dataset 1	0.90	0.91	0.82	0.83	1.00	0.74
FC-4	Dataset 3	0.85	0.84	0.70	0.89	0.80	0.84

446

1
2
3
4 447 The findings from the two analyses demonstrate the efficacy of the proposed ML-based
5
6
7 448 framework in accurately predicting B3PPs, with cross-validation accuracy values exceeding
8
9
10 449 90% and values between 75% and 90% for the external validation set. Furthermore, this study
11
12
13 450 highlights the contribution of the descriptors evaluated in terms of their association with BBB
14
15
16
17 451 permeability and their comparison with descriptors associated with the charge distribution of
18
19
20 452 the molecules. However, the independent test step involved a limited number of samples, with
21
22
23
24 453 each erroneous prediction causing 5% reduction in the accuracy of each model. Consequently,
25
26
27 454 determining the optimal BrainPepPass configuration was challenging. Therefore, this study
28
29
30 455 also employs the leave-one-out cross-validation (LOOCV) metric as a complementary analysis
31
32
33
34 456 to evaluate the proposed framework.

35
36
37 45738
39 458 LOOCV Analysis
40
41
4243 459
44

45
46 460 LOOCV is a model evaluation method similar to k-fold cross-validation, in which the testing
47
48
49 461 set contains only one sample ($k = 1$), and the remaining samples are used for training.⁶⁷ We
50
51
52 462 conducted an LOOCV evaluation using three complete datasets (consisting of the training and
53
54
55
56 463 testing subsets) for each FC. The results demonstrated that FC-2 enabled the BrainPepPass to
57
58
59
60

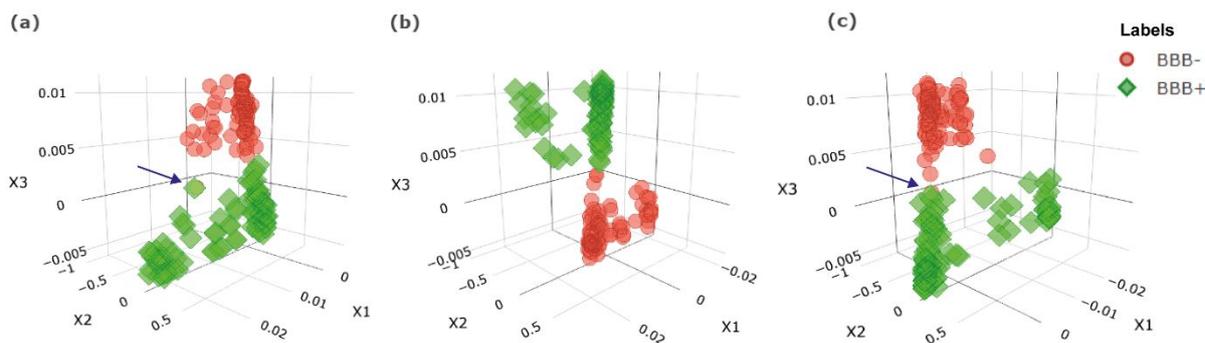
1
2
3
4 464 attain a mean accuracy with a peak value in all datasets, whereas FC-4 displayed comparable
5
6
7 465 efficacy only for Dataset 2. Datasets 1 and 3 achieved accuracy scores of 99% and 98%,
8
9
10 466 respectively. Supplementary Table S8 lists metric values obtained by BrainPepPass using the
11
12
13
14 467 three datasets in LOOCV.

15
16
17 468 A comparison of the results of LOOCV with those obtained in the 10-fold cross-validation
18
19
20 469 shows that the feature compositions that provided more information for predicting B3PPs were
21
22
23
24 470 FC-2 and FC-4, highlighting the importance of the molecular descriptors of both FCs in
25
26
27 471 differentiating the two classes of peptides. The outcomes from the LOOCV of the datasets
28
29
30 472 belonging to these two FCs were analyzed through a pairwise comparison using an ANOVA
31
32
33
34 473 test. The results indicate no statistically significant difference between the means of Datasets
35
36
37 474 1 and 3, with p -values of 0.318 and 0.157, respectively. Upon comparing the performance of
38
39
40
41 475 the most effective models in LOOCV with that achieved in an independent test, FC-3 and FC-4
42
43
44 476 descriptors ranked among the highest in their ability to predict B3PPs. Specifically,
45
46
47 477 BrainPepPass, based on FC-3, exhibited only a single misclassification in LOOCV and two
48
49
50
51 478 misclassifications in the independent test. In contrast, the model based on FC-4 achieved
52
53
54 479 satisfactory classification in LOOCV but failed to correctly classify the three molecules in the
55
56
57 480 external validation. Although FC-2 obtained the third-highest accuracy value in the
58
59

1
2
3
4 481 independent test, it outperformed FC-3 by achieving the maximum classification value in the
5
6
7 482 LOOCV experiment.
8
9

10 483 According to the three evaluation methods employed in the proposed ML-based framework,
11
12
13 484 FC-4 predicted B3PPs with the highest accuracy. This descriptor group employed a less
14
15
16
17 485 complex model consisting of 19 descriptors in contrast to FC-2, which also displayed high
18
19
20 486 accuracy values. The success of FC-4 can be attributed to the efficacy sLE algorithm in
21
22
23 487 reducing the dimensionality of the molecular descriptors. Figure 6 shows the projection of the
24
25
26
27 488 molecular descriptors belonging to this feature composition in a 3D space after dimensionality
28
29
30 489 reduction was performed during the pattern learning phase of the proposed framework. Our
31
32
33
34 490 observations revealed that Dataset 1 exhibits an overlap between two peptides belonging to
35
36
37 491 different classes (see the blue arrow in Figure 6a), whereas Dataset 3 displayed an overlap
38
39
40 492 between at least three peptides from distinct classes (see the blue arrow in Figure 6c). This
41
42
43
44 493 pattern is consistent with the results shown in Figure 5c. Additionally, the 3D projections of
45
46
47 494 FC-4 reveal the potential for differentiation of BBB+ and BBB- peptides, besides clustering
48
49
50 495 both classes, through the integration of molecular descriptors investigated in FC-1 and those
51
52
53
54 496 selected from Mordred.
55
56

57 497
58
59
60



498

499 **Figure 6.** Dimensionality reduction result of BrainPepPass in pattern learning stage for FC-4.

500 (a) Dataset 1. (b) Dataset 2. (c) Dataset 3.

501

502 Performance Comparison with other ML Algorithms

503 We compared the performance of BrainPepPass with SVM, ANN, and XGB models using
504 the same three evaluation methods as used previously. We also compared the proposed ML-
505 based framework with the same architecture shown in Figure 2 using the LLE, Isomap, and
506 UMAP algorithms. The results of the optimal models for 10-fold cross-validation were
507 computed for all combinations of FCs and datasets. The results of the best models are presented
508 in Table 2. Supplementary Table S9 presents the search range and best hyperparameters
509 achieved by LLE, Isomap, and UMAP in this analysis.

510

1
2
3
4 511 **Table 2.** Cross-validation analysis for the best BrainPepPass, ANN, SVM, XGB, LLE, Isomap,
5
6
7 512 and UMAP models by each FC.
8
9

Method	FC-1	FC-2	FC-3	FC-4
BrainPepPass	0.93 ± 0.04	0.99 ± 0.01	0.98 ± 0.02	0.98 ± 0.02
ANN	0.58 ± 0.14	0.57 ± 0.12	0.60 ± 0.14	0.61 ± 0.15
SVM	0.55 ± 0.12	0.59 ± 0.16	0.58 ± 0.09	0.55 ± 0.12
XGB	0.55 ± 0.11	0.60 ± 0.10	0.61 ± 0.11	0.59 ± 0.05
LLE	0.55 ± 0.11	0.61 ± 0.13	0.46 ± 0.14	0.51 ± 0.10
Isomap	0.45 ± 0.09	0.44 ± 0.08	0.48 ± 0.14	0.53 ± 0.10
UMAP	0.48 ± 0.16	0.52 ± 0.09	0.46 ± 0.13	0.54 ± 0.13

10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31 513
32
33
34 514 The results of the cross-validation analysis revealed that BrainPepPass surpassed the average
35
36
37 515 accuracy of the other ML algorithms for all feature compositions. These classifiers could not
38
39
40
41 516 achieve an accuracy higher than 61% even for FCs with a reduced number of molecular
42
43
44 517 descriptors. The frameworks based on LLE, Isomap, and UMAP achieved significantly lower
45
46
47 518 average accuracy results, with values between 0.44 and 0.61, when compared to BrainPepPass.
48
49
50
51 519 This discrepancy in performance can be attributed to the capacity of each technique to capture
52
53
54 520 the nonlinear correlation between the descriptors and permeability classes. Our observations
55
56
57 521 indicate that the sLE algorithm incorporated in the BrainPepPass effectively discriminates the
58
59
60

1
2
3
4 522 peptides that can cross the BBB from those that cannot, learning the nonlinear correlations
5
6
7 523 between molecular descriptors and peptide labels, even better than the other three manifold DR
8
9
10 524 algorithms employed in this study, contributing to the overall high performance of
11
12
13
14 525 BrainPepPass.

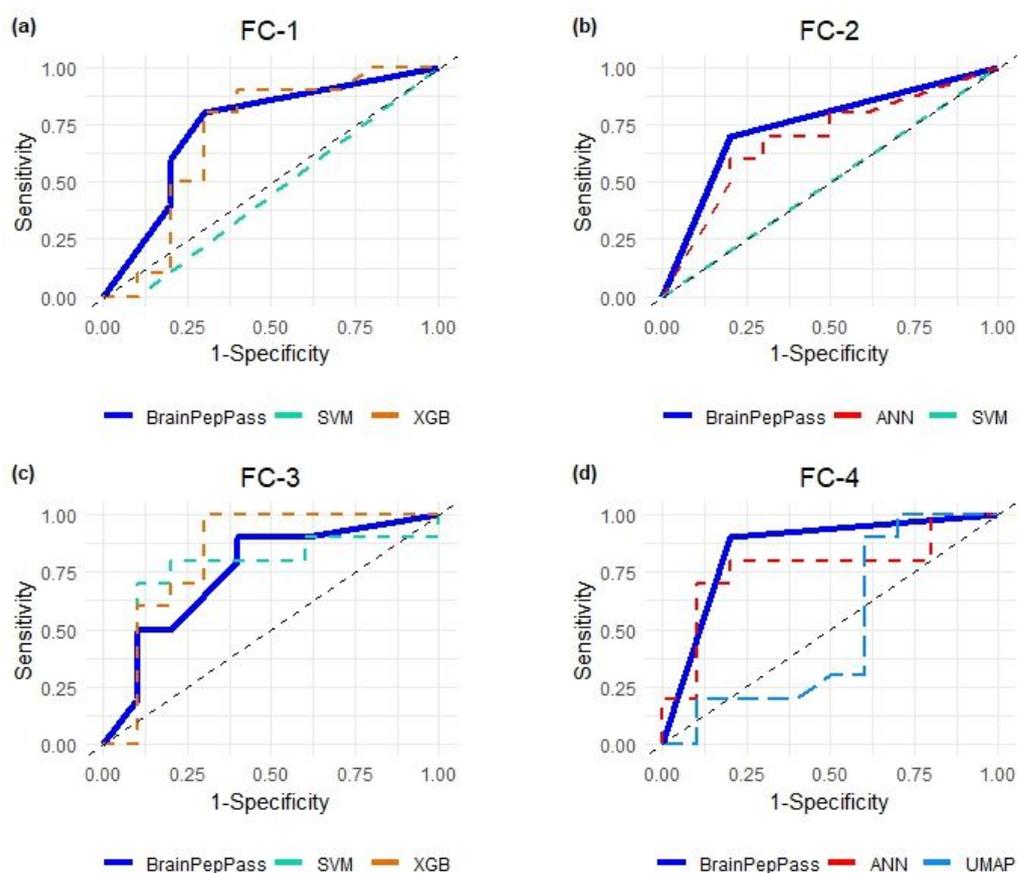
15
16
17 526 We also applied an independent test to the other algorithms and compared them with the
18
19
20 527 BrainPepPass (see Table 3). The results demonstrated that the proposed tool outperformed the
21
22
23
24 528 ML models in terms of accuracy and other evaluation metrics for almost all FCs in the B3PP
25
26
27 529 prediction. Among the state-of-the-art algorithms and other frameworks, the ANN model
28
29
30 530 achieved the highest accuracy for FC-3 (80%); however, it did not surpass the performance of
31
32
33
34 531 the proposed ML-based framework for the same FC (90%). Furthermore, the F1-score, MCC,
35
36
37 532 precision, recall and ROC-AUC metrics indicated the exceptional performance of
38
39
40 533 BrainPepPass, achieving higher values than the other techniques in most scenarios. A
41
42
43
44 534 comparison of the results obtained by the proposed framework with those of other models using
45
46
47 535 LLE, Isomap, and UMAP shows that in no scenario did these models surpass the performance
48
49
50 536 of BrainPepPass for any of the metrics, thereby corroborating the results achieved in the cross-
51
52
53
54 537 validation analysis. The performance metric values obtained by ANN, SVM, XGB, LLE,
55
56
57
58
59
60

1
2
3
4 538 Isomap, and UMAP using the three datasets in the 10-fold cross-validation and independent
5
6
7 539 tests are provided in Supplementary Tables S10, S11, S12, S13, S14, and S15.
8
9

10 540 Similarly, we evaluated the predictive capacity of all ML models using LOOCV and
11
12
13 541 compared their performance with that of BrainPepPass (see Figures 5d and 5e). Our findings
14
15
16
17 542 indicate that the classifiers failed to surpass the predictive capacity of the proposed ML-based
18
19
20 543 framework for all the FCs. Among the ML models, XGB achieved the highest average accuracy
21
22
23
24 544 value of 75.26% for FC-4, which was lower than the value achieved by BrainPepPass with the
25
26
27 545 same feature composition. The average accuracy values for the ANN and SVM ranged between
28
29
30 546 58.25% and 72.16% across different FCs. The results achieved by the frameworks using
31
32
33
34 547 manifold DR algorithms were also unable to surpass the predictive capacity of BrainPepPass.
35
36
37 548 The framework using LLE showed the best performance with an average accuracy of 70.62%
38
39
40 549 for FC-3, whereas Isomap and UMAP achieved values between 62.89% and 69.07% for all
41
42
43
44 550 FCs. The performance values obtained by ANN, SVM, XGB, LLE, Isomap, and UMAP models
45
46
47 551 using the three datasets in LOOCV analysis are provided in Supplementary Tables S16, S17,
48
49
50 552 S18, S19, S20, and S21.
51
52
53

54 553 The findings indicated in Table 3 were corroborated by evaluating the independent test based
55
56
57 554 on the results of the ROC curve shown in Figure 7. The achieved ROC-AUC value for
58
59
60

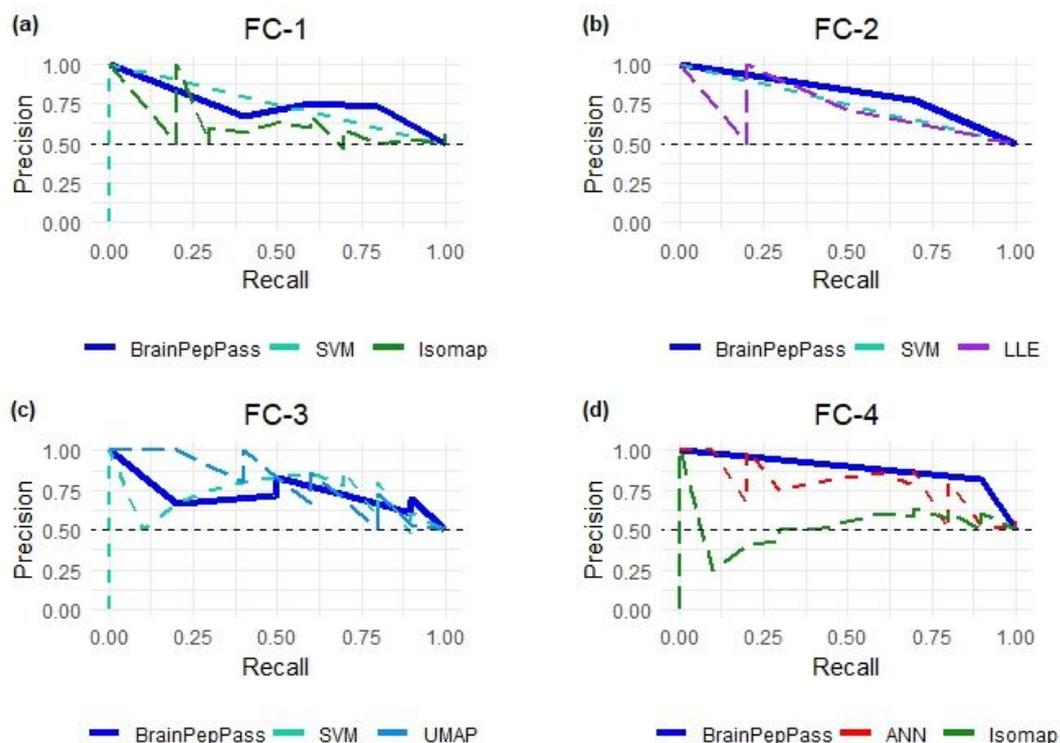
1
2
3
4 555 BrainPepPass for FC-1, FC-2, and FC-3 remained within a narrow range between 0.74 and
5
6
7 556 0.75, whereas for FC-4, it was 0.84, surpassing the other models. The XGB algorithm
8
9
10 557 demonstrated the second-best performance in this analysis, achieving an ROC-AUC value of
11
12
13 558 0.83 for FC-3. The ROC curves for all models are provided in Supplementary Figure S1.
14
15
16
17 559



560
561 **Figure 7.** ROC curves comparison among the best BrainPepPass and the best and worst model
562 by each FC in the independent test. (a) FC-1. (b) FC-2. (c) FC-3. (d) FC-4.

1
2
3
4 563 The difference in the performance of the proposed BrainPepPass for different FCs was
5
6
7 564 assessed from the corresponding precision-recall curve (see Figure 8). The proposed
8
9
10 565 framework achieved the highest average precision (AP) score of 0.66 for FC-1. The AP score
11
12
13 566 for FC-2, FC-3, and FC-4 were 0.69, 0.70, and 0.78, respectively. The AP values for all FCs
14
15
16
17 567 when the ANN, SVM, and XGB algorithms were used ranged from 0.5 to 0.8, whereas they
18
19
20 568 were between 0.51 and 0.82 for the manifold-based algorithms. These results indicate that,
21
22
23
24 569 despite the strong performance of BrainPepPass, the model using the UMAP algorithm
25
26
27 570 demonstrates a greater balance between precision and recall when employing FC-3. The
28
29
30 571 precision-recall curves for all models are provided in Supplementary Figure S2.

31
32
33
34 572
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60



573

574 **Figure 8.** Precision-recall curves comparison among the best BrainPepPass and the best and
 575 worst model by each FC in the independent test. (a) FC-1. (b) FC-2. (c) FC-3. (d) FC-4.

576

577 **Table 3.** Independent test analysis for the best BrainPepPass, ANN, SVM, XGB, LLE, Isomap,
 578 and UMAP models by each FC.

FC-1	Method	Accuracy	F1-score	MCC	Precision	Recall	ROC-AUC
	BrainPepPass	0.75	0.70	0.52	0.85	0.75	0.74
	ANN	0.55	0.52	0.10	0.55	0.55	0.55
	SVM	0.65	0.58	0.31	0.71	0.65	0.45
	XGB	0.60	0.50	0.21	0.66	0.60	0.70

LLE	0.65	0.63	0.30	0.66	0.60	0.57
Isomap	0.60	0.60	0.20	0.60	0.60	0.60
UMAP	0.75	0.70	0.52	0.85	0.60	0.45
<hr/>						
FC-2						
<hr/>						
BrainPepPass	0.80	0.81	0.61	0.75	0.80	0.75
ANN	0.65	0.58	0.31	0.71	0.65	0.69
SVM	0.55	0.52	0.10	0.55	0.55	0.50
XGB	0.70	0.70	0.40	0.70	0.70	0.63
LLE	0.65	0.58	0.31	0.71	0.50	0.64
Isomap	0.60	0.42	0.25	0.75	0.30	0.58
UMAP	0.70	0.70	0.40	0.70	0.70	0.44
<hr/>						
FC-3						
<hr/>						
BrainPepPass	0.90	0.90	0.81	0.83	0.90	0.75
ANN	0.80	0.80	0.60	0.80	0.80	0.77
SVM	0.70	0.66	0.40	0.75	0.70	0.75
XGB	0.70	0.70	0.40	0.70	0.70	0.83
LLE	0.75	0.73	0.50	0.77	0.70	0.71
Isomap	0.70	0.72	0.40	0.66	0.80	0.76
UMAP	0.75	0.73	0.50	0.77	0.70	0.76
<hr/>						
FC-4						
<hr/>						
BrainPepPass	0.85	0.84	0.70	0.88	0.85	0.84
ANN	0.65	0.69	0.31	0.61	0.65	0.77
SVM	0.65	0.58	0.31	0.71	0.65	0.55
XGB	0.70	0.75	0.43	0.64	0.70	0.72
LLE	0.60	0.60	0.20	0.60	0.60	0.53
Isomap	0.60	0.63	0.20	0.58	0.70	0.55

UMAP	0.60	0.55	0.20	0.62	0.50	0.50
------	------	------	------	------	------	------

579 It is noteworthy to compare the performance of the BrainPepPass with previously developed
580 techniques for predicting B3PPs. While some ML-based tools, such as BBPpred⁶⁸, B3Pred,⁶⁹
581 BBPpredict,³⁷ and SCMB3PP⁷⁰ have been developed to predict the BBB permeability of
582 peptides using ML algorithms trained with properties extracted from the primary structure of
583 natural peptides encoded in FASTA format, the proposed ML-based framework presented
584 herein employs a distinct approach by incorporating the 3D structure of these molecules
585 encoded in MOL format. Additionally, most peptides used for training and testing
586 BrainPepPass contain chemical modifications, which further distinguishes our tool from those
587 that focus on natural peptides.

588 We conducted a comparative analysis between BrainPepPass and the BBBPpred,
589 BBBPpredict, and SCMB3PP algorithms, which are available for public and free use. To assess
590 the performance of the proposed model against other tools in an independent test, we selected
591 the version based on FC-4 and trained with Dataset 2, which achieved the best performance in
592 LOOCV analyses. We used 17 natural BBB+ peptides extracted from Brainpeps to compare
593 the ML models performance, none of these molecules were utilized in any of the previously
594 described training or independent testing steps for the selected BrainPepPass version. This

1
2
3
4 595 dataset was balanced with 17 natural BBB- peptides randomly extracted from the test dataset
5
6
7 596 of the SCMB3PP tool, resulting in 34 structures for this analysis. We also developed a
8
9
10 597 BrainPepPass model with FC-4, named BrainPepPass-N, which was exclusively trained using
11
12
13 598 natural peptides collected from the same dataset that was used to train SCMB3PP model. Table
14
15
16
17 599 4 presents the values achieved by all the algorithms based on the key metrics. The peptide
18
19
20 600 sequences used in this analysis are listed in Supplementary Table S22.
21
22

601

602

30 603 **Table 4.** Analysis of independent test comparing BrainPepPass and BrainPepPass-N with
31
32
33 604 BBPpred, BBPpredict, and SCMB3PP algorithms using natural peptides.
34
35
36

Algorithm	Accuracy	F1-score	MCC	Precision	Recall
BrainPepPass	0.52	0.55	0.06	0.55	0.55
BrainPepPass-N	0.97	1.0	0.94	1.0	1.0
BBPpred	0.64	0.71	0.33	0.60	0.88
BBPpredict	0.55	0.66	0.15	0.53	0.88
SCMB3PP	0.91	0.90	0.82	0.93	0.88

605

1
2
3
4 606 According to the results presented in Table 4, BrainPepPass-N achieved the best outcomes,
5
6
7 607 attaining an accuracy of approximately 97%, along with values exceeding 94% for the other
8
9
10 608 metrics. This indicates that the proposed method, trained only on natural peptides and utilizing
11
12
13 609 molecular descriptors from FC-4, can predict the permeability of natural peptides across the
14
15
16
17 610 BBB with greater accuracy than that of the other tools. It is also important to highlight that the
18
19
20 611 BrainPepPass model that was not exclusively trained on natural peptides failed to outperform
21
22
23 612 the other tools. This could be attributed to the underfitting of this model with respect to natural
24
25
26
27 613 peptide data, considering that it was predominantly trained on structures featuring chemical
28
29
30 614 modifications.

31
32
33
34 615 Therefore, based on the results presented in this study, BrainPepPass exhibits exceptional
35
36
37 616 performance in predicting peptide penetration across the BBB, surpassing existing ML
38
39
40 617 classifiers in this area of research. BrainPepPass achieved average accuracy values exceeding
41
42
43 618 93% in the 10-fold cross-validation and between 75% and 90% in the independent test, with
44
45
46
47 619 average accuracy values ranging between 99.48% and 100% according to LOOCV. For the
48
49
50 620 FC-4 model, which exhibited a positive relationship between efficiency and complexity,
51
52
53 621 average accuracy values of 99.21%, 75%, and 99.48% were achieved in cross-validation,
54
55
56
57 622 independent testing, and LOOCV, respectively, across all three datasets. Although
58
59
60

1
2
3
4 623 BrainPepPass based on FC-4 did not achieve the best performance in predicting some natural
5
6
7 624 peptides, BrainPepPass-N version attained an accuracy of 97% in the same test. These
8
9
10 625 outcomes demonstrate that the proposed tool has impressive predictive capabilities in
11
12
13
14 626 determining whether natural or chemically modified peptides can penetrate the BBB, based on
15
16
17 627 the molecular descriptors examined.
18
19

20 628

21
22
23 629 CONCLUSION
24
25

26
27 630

28
29 631 Predicting the ability of natural and chemically modified peptides to penetrate the BBB is a
30
31
32
33 632 significant challenge in computational and medicinal chemistry. The development of an
34
35
36 633 efficient computational tool to perform this task requires solving the following problems: (1)
37
38
39 634 obtaining experimentally validated data that include natural and chemically modified peptides;
40
41
42
43 635 (2) performing an exploratory analysis of the correlation between several molecular descriptors
44
45
46 636 related to BBB permeability; and (3) training a robust and powerful ML-based model to learn
47
48
49 637 the non-linear pattern between peptide descriptors and permeability classes. However, public
50
51
52
53 638 information on the pharmacokinetic properties of peptides that could be used as a reference for
54
55
56
57
58
59
60

1
2
3
4 639 experimental studies is scarce, making generating large datasets that facilitates the training of
5
6
7 640 the algorithms difficult.
8
9

10 641 Our predictive model relies on an algorithmic architecture that reduces the various
11
12
13 642 dimensions of molecular descriptors derived from peptides to three. By leveraging the ability
14
15
16
17 643 of the sLE technique to cluster and segregate samples into their respective classes, our model
18
19
20 644 incorporates a robust preprocessing step, thereby streamlining the prediction process of B3PPs.
21
22
23 645 Moreover, our ML-based framework offers the additional advantage of processing information
24
25
26
27 646 extracted from peptide structures in the MOL format, which encodes chemical modifications
28
29
30 647 and cyclic chains in the molecular structure, thereby achieving a novel breakthrough in this
31
32
33 648 field of research and improving the exploration of increasingly complex structures within this
34
35
36
37 649 molecular class. Additionally, our investigation highlights the correlation between several
38
39
40 650 molecular descriptors and BBB permeability, specifically emphasizing the role of charge
41
42
43
44 651 distribution properties in the ability of peptides to permeate through the BBB.
45
46

47 652 Another advantage our of study was the improved performance of BrainPepPass relative to
48
49
50 653 other machine learning models in predicting peptide penetration. The predictive capability of
51
52
53
54 654 our tool in all applied tests surpassed that of other machine-learning classifiers as well as that
55
56
57 655 of the same framework but with other manifold DR algorithms. Furthermore, the proposed
58
59
60

1
2
3
4 656 framework demonstrated good performance compared with other available tools for predicting
5
6
7 657 natural B3PPs. This proves the ability of our tool in assisting the virtual screening of new
8
9
10 658 peptides that penetrate the BBB, thereby contributing to the discovery and development of new
11
12
13
14 659 bioactive molecules capable of reaching the CNS.

15
16
17 66018
19
20 661 ASSOCIATED CONTENT21
22
23
24 662 **Data Availability Statement**

25
26
27
28 663 The BrainPepPass tool is available in a GitHub repository, which can be accessed at:
29
30
31 664 <https://github.com/ewerton-cristhian/BrainPepPass>. This repository contains information
32
33
34
35 665 about the online versions of the BrainPepPass available for users, a user manual with
36
37
38 666 instructions on how to use the tools, and the ML models used in the framework.

39
40
41
42 667 The source code of the BrainPepPass in Python language to execute the best model can be
43
44
45 668 accessed at <https://figshare.com/s/18d704599c397f54b3ac>. The dataset of peptide structures
46
47
48
49 669 can be accessed at <https://figshare.com/s/f8ae1e2f6e4b2170807f>. The scripts used to generate
50
51
52 670 and evaluate the BrainPepPass and other ML models in the present work is available at
53
54
55
56 671 <https://figshare.com/s/8bc7ab7b424e04f680e0>.

1
2
3
4 6725
6 673 **Supporting Information**
78
9 674 The Supporting Information is available free of charge at JCIM web site.
10
11
1213
14 675 List of peptides used; Table S1, list of molecular descriptors in FC-1; Table S2, list of15
16
17 676 molecular descriptors in FC-2; Table S3, list of molecular descriptors in FC-3; Table S4,18
19
20 677 hyperparameters employed in grid search for training XGBr in BrainPepPass; Table S5,21
22
23 678 hyperparameters employed in grid search for training XGBc in BrainPepPass; Table S6, results24
25
26 679 reached in cross-validation and independent test by BrainPepPass for all FCs and datasets;27
28
29 680 Table S7, results reached in LOOCV BrainPepPass for all FCs and datasets; Table S8,30
31
32 681 hyperparameters employed in grid search for training LLE, Isomap, and UMAP; Table S9,33
34
35 682 results reached in cross-validation and independent test by the ANN for all FCs and datasets;36
37
38 683 Table S10, results reached in cross-validation and independent test by the SVM for all FCs and39
40
41 684 datasets; Table S11, results reached in cross-validation and independent test by the XGB for42
43
44 685 all FCs and datasets; Table S12, results reached in cross-validation and independent test by the45
46
47 686 framework based on LLE for all FCs and datasets; Table S13, results reached in cross-48
49
50 687 validation and independent test by the framework based on Isomap for all FCs and datasets;51
52
53 688 Table S14, results reached in cross-validation and independent test by the framework based on
54
55
56
57
58
59
60

1
2
3
4 689 UMAP for all FCs and datasets; Table S15, results reached in LOOCV by the ANN for all FCs
5
6
7 690 and datasets; Table S16, results reached in LOOCV by the SVM for all FCs and datasets; Table
8
9
10 691 S17, results reached in LOOCV by the XGB for all FCs and datasets; Table S18, results reached
11
12
13 692 in LOOCV by the framework based on LLE for all FCs and datasets; Table S19, results reached
14
15
16
17 693 in LOOCV by the framework based on Isomap for all FCs and datasets; Table S20, results
18
19
20 694 reached in LOOCV by the framework based on UMAP for all FCs and datasets; Table S21, list
21
22
23
24 695 of natural peptide sequences used to compare BrainPepPass with other online tools; Table S22,
25
26
27 696 ROC curves obtained by the best BrainPepPass, ANN, SVM, XGB, LLE, Isomap, and UMAP
28
29
30 697 models by each FC in independent test; Figure S1, Precision-recall curves obtained by the best
31
32
33
34 698 BrainPepPass, ANN, SVM, XGB, LLE, Isomap, and UMAP models by each FC in independent
35
36
37 699 test; Figure S2.

40
41 700

42
43
44 701 AUTHOR INFORMATION

45
46
47 702 **Corresponding Authors**

48
49
50
51 703 **Bart De Spiegeleer** – Drug Quality and Registration (DruQuaR) Group,

52
53
54
55 704 Faculty of Pharmaceutical Sciences, Ghent University, B-9000 Ghent, Belgium;

1
2
3
4 705 orcid.org/0000-0001-6794-3108; Phone: +32 9 2648100; Email: Bart.DeSpiegeleer@ugent.be
5
6
7

8 706 **Kauê Santana da Costa**- Laboratório de Simulação Computacional, Campos Marechal
9

10 707 Rondon, Instituto de Biodiversidade, Universidade Federal do Oeste do Pará, 68040-255,
11
12

13
14 708 Santarém, Pará, Brasil. E-mail: kaue.costa@ufopa.ed.br.
15
16

17
18 709 **Claudio de Souza de Sales Júnior** - Laboratório de Inteligência Computacional e Pesquisa
19

20
21 710 Operacional, Campos Belém, Instituto de Tecnologia, Universidade Federal do Pará, 66075-
22
23

24
25 711 110, Belém, Pará, Brasil. E-mail: cssj@ufpa.br.
26
27

28
29 712
30

31 713 **Authors**
32
33

34
35 714 **Ewerton Cristhian Lima de Oliveira** - Laboratório de Inteligência Computacional e Pesquisa
36
37

38 715 Operacional, Campos Belém, Instituto de Tecnologia, Universidade Federal do Pará, 66075-
39
40

41
42 716 110, Belém, Pará, Brasil; Instituto Tecnológico Vale, 66055-090, Belém, Pará, Brasil;
43
44

45 717 orcid.org/0000-0002-2338-7178.
46
47

48
49 718 **Hannah Hirmz** - Drug Quality and Registration (DruQuaR) Group, Faculty of Pharmaceutical
50
51

52
53 719 Sciences, Ghent University, B-9000 Ghent, Belgium; orcid.org/0000-0002-7446-0267.
54
55
56
57
58
59
60

- 1
2
3
4 720 **Evelien Wynendaele** - Drug Quality and Registration (DruQuaR) Group, Faculty of
5
6
7 721 Pharmaceutical Sciences, Ghent University, B-9000 Ghent, Belgium; orcid.org/0000-0003-
8
9
10 722 0969-7580.
11
12
13
14 723 **Juliana Auzier Seixas Feio** - Laboratório de Inteligência Computacional e Pesquisa
15
16
17 724 Operacional, Campos Belém, Instituto de Tecnologia, Universidade Federal do Pará, 66075-
18
19
20
21 725 110, Belém, Pará, Brasil; orcid.org/0009-0009-9781-5911
22
23
24
25 726 **Igor Matheus Souza Moreira** - Laboratório de Inteligência Computacional e Pesquisa
26
27
28 727 Operacional, Campos Belém, Instituto de Tecnologia, Universidade Federal do Pará, 66075-
29
30
31
32 728 110, Belém, Pará, Brasil; orcid.org/0000-0002-8145-9395.
33
34
35
36 729 **Anderson H. Lima** - Laboratório de Planejamento e Desenvolvimento de Fármacos, Instituto
37
38
39 730 de Ciências Exatas e Naturais, Universidade Federal do Pará, 66075-110, Belém, Pará, Brasil;
40
41
42
43 731 orcid.org/0000-0002-8451-9912.
44
45
46
47 732 **Kauê Santana** - Laboratório de Simulação Computacional, Campos Marechal Rondon, Instituto
48
49
50 733 de Biodiversidade, Universidade Federal do Oeste do Pará, 68040-255, Santarém, Pará, Brasil.
51
52
53
54 734 orcid.org/0000-0002-2735-8016.
55
56
57
58 735 **Bart De Spiegeleer** – Drug Quality and Registration (DruQuaR) Group,
59
60

1
2
3
4 736 Faculty of Pharmaceutical Sciences, Ghent University, B-9000 Ghent, Belgium;
5
6
7

8 737 orcid.org/0000-0001-6794-3108.
9
10
11

12 738 **Claudio de Souza de Sales Junior** - Laboratório de Inteligência Computacional e Pesquisa
13
14

15 739 Operacional, Campos Belém, Instituto de Tecnologia, Universidade Federal do Pará, 66075-
16
17

18 740 110, Belém, Pará, Brasil; orcid.org/0000-0002-2735-1383.
19
20
21

22
23 741 **Notes**
24
25

26 742 The authors declare no competing financial interest.
27
28
29

30 743 **Author Contributions**
31
32
33

34 744 The manuscript was written through contributions of all authors. All authors have given
35
36

37 745 approval to the final version of the manuscript.
38
39
40

41 746 **ACKNOWLEDGEMENTS**
42
43
44

45
46 747 The authors are grateful to Conselho Nacional de Desenvolvimento Científico e Tecnológico
47
48

49 748 (CNPq), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Pró-reitoria
50
51

52 749 de Pesquisa da UFPA (PROPESP/UFPA), and Instituto Tecnológico Vale (ITV) for their
53
54

55
56 750 financial and technical support.
57
58
59
60

751

752 REFERENCES

753

- 754 (1) Geldenhuys, W. J.; Mohammad, A. S.; Adkins, C. E.; Lockman, P. R. Molecular
755 Determinants of Blood-Brain Barrier Permeation. *Ther Deliv* **2015**, *6* (8), 961–971.
756 <https://doi.org/10.4155/tde.15.32>.
- 757 (2) Oller-Salvia, B.; Sánchez-Navarro, M.; Giralt, E.; Teixidó, M. Blood–Brain Barrier
758 Shuttle Peptides: An Emerging Paradigm for Brain Delivery. *Chem Soc Rev* **2016**, *45*
759 (17), 4690–4707. <https://doi.org/10.1039/C6CS00076B>.
- 760 (3) Díaz-Perlas, C.; Oller-Salvia, B.; Sánchez-Navarro, M.; Teixidó, M.; Giralt, E.
761 Branched BBB-Shuttle Peptides: Chemoselective Modification of Proteins to Enhance
762 Blood–Brain Barrier Transport. *Chem Sci* **2018**, *9* (44), 8409–8415.
763 <https://doi.org/10.1039/C8SC02415D>.
- 764 (4) de Oliveira, E. C. L.; da Costa, K. S.; Taube, P. S.; Lima, A. H.; Junior, C. de S. de S.
765 Biological Membrane-Penetrating Peptides: Computational Prediction and
766 Applications. *Front Cell Infect Microbiol* **2022**, *12*.
767 <https://doi.org/10.3389/fcimb.2022.838259>.
- 768 (5) Kastin, A. J.; Nissen, C.; Schally, A. V.; Coy, D. H. Blood-Brain Barrier, Half-Time
769 Disappearance, and Brain Distribution for Labeled Enkephalin and a Potent Analog.
770 *Brain Res Bull* **1976**, *1* (6), 583–589. [https://doi.org/10.1016/0361-9230\(76\)90085-X](https://doi.org/10.1016/0361-9230(76)90085-X).
- 771 (6) Pelletier, G.; Labrie, F.; Kastin, A. J.; Schally, A. V. Radioautographic Localization of
772 Radioactivity in Rat Brain after Intracarotid Injection of ^{125}I - α -Melanocyte-
773 Stimulating Hormone. *Pharmacol Biochem Behav* **1975**, *3* (4), 671–674.
774 [https://doi.org/10.1016/0091-3057\(75\)90190-2](https://doi.org/10.1016/0091-3057(75)90190-2).
- 775 (7) Kastin, A. J.; Nissen, C.; Nikolics, K.; Medzihradzky, K.; Coy, D. H.; Teplan, I.;
776 Schally, A. V. Distribution of 3H - α -MSH in Rat Brain. *Brain Res Bull* **1976**, *1* (1), 19–
777 26. [https://doi.org/10.1016/0361-9230\(76\)90045-9](https://doi.org/10.1016/0361-9230(76)90045-9).

- 1
2
3
4 778 (8) Banks, W. A. Peptides and the Blood–Brain Barrier. *Peptides (N.Y.)* **2015**, *72*, 16–19.
5 779 <https://doi.org/10.1016/j.peptides.2015.03.010>.
6
7
8 780 (9) Wang, D.; Chen, F.; Han, Z.; Yin, Z.; Ge, X.; Lei, P. Relationship Between Amyloid- β
9 781 Deposition and Blood–Brain Barrier Dysfunction in Alzheimer’s Disease. *Front Cell*
10 782 *Neurosci* **2021**, *15*. <https://doi.org/10.3389/fncel.2021.695479>.
11
12
13
14 783 (10) Baskin, D. G.; Stein, L. J.; Ikeda, H.; Woods, S. C.; Figlewicz, D. P.; Porte, D.;
15 784 Greenwood, M. R. C.; Dorsa, D. M. Genetically Obese Zucker Rats Have Abnormally
16 785 Low Brain Insulin Content. *Life Sci* **1985**, *36* (7), 627–633.
17 786 [https://doi.org/10.1016/0024-3205\(85\)90166-3](https://doi.org/10.1016/0024-3205(85)90166-3).
18
19
20
21 787 (11) Kaiyala, K. J.; Prigeon, R. L.; Kahn, S. E.; Woods, S. C.; Schwartz, M. W. Obesity
22 788 Induced by a High-Fat Diet Is Associated with Reduced Brain Insulin Transport in Dogs.
23 789 *Diabetes* **2000**, *49* (9), 1525–1533. <https://doi.org/10.2337/diabetes.49.9.1525>.
24
25
26
27 790 (12) Urayama, A.; Banks, W. A. Starvation and Triglycerides Reverse the Obesity-Induced
28 791 Impairment of Insulin Transport at the Blood-Brain Barrier. *Endocrinology* **2008**, *149*
29 792 (7), 3592–3597. <https://doi.org/10.1210/en.2008-0008>.
30
31
32
33 793 (13) Banks, W. A.; Jaspan, J. B.; Kastin, A. J. Effect of Diabetes Mellitus on the Permeability
34 794 of the Blood-Brain Barrier to Insulin. *Peptides (N.Y.)* **1997**, *18* (10), 1577–1584.
35 795 [https://doi.org/10.1016/S0196-9781\(97\)00238-6](https://doi.org/10.1016/S0196-9781(97)00238-6).
36
37
38
39 796 (14) Stalmans, S.; Bracke, N.; Wynendaele, E.; Gevaert, B.; Peremans, K.; Burvenich, C.;
40 797 Polis, I.; De Spiegeleer, B. Cell-Penetrating Peptides Selectively Cross the Blood-Brain
41 798 Barrier in Vivo. *PLoS One* **2015**, *10* (10), e0139652.
42 799 <https://doi.org/10.1371/journal.pone.0139652>.
43
44
45
46 800 (15) Zhou, X.; Smith, Q. R.; Liu, X. Brain Penetrating Peptides and Peptide–Drug Conjugates
47 801 to Overcome the Blood–Brain Barrier and Target CNS Diseases. *Wiley Interdiscip Rev*
48 802 *Nanomed Nanobiotechnol* **2021**, *13* (4). <https://doi.org/10.1002/wnan.1695>.
49
50
51
52 803 (16) He, S.; Zhiti, A.; Barba-Bon, A.; Hennig, A.; Nau, W. M. Real-Time Parallel Artificial
53 804 Membrane Permeability Assay Based on Supramolecular Fluorescent Artificial
54 805 Receptors. *Front Chem* **2020**, *8*. <https://doi.org/10.3389/fchem.2020.597927>.
55
56
57
58
59
60

- 1
2
3
4 806 (17) Radan, M.; Djikic, T.; Obradovic, D.; Nikolic, K. Application of in Vitro PAMPA
5 807 Technique and in Silico Computational Methods for Blood-Brain Barrier Permeability
6 808 Prediction of Novel CNS Drug Candidates. *European Journal of Pharmaceutical*
7 809 *Sciences* **2022**, *168*, 106056. <https://doi.org/10.1016/j.ejps.2021.106056>.
- 10
11 810 (18) Bagchi, S.; Chhibber, T.; Lahooti, B.; Verma, A.; Borse, V.; Jayant, R. D. In-Vitro
12 811 Blood-Brain Barrier Models for Drug Screening and Permeation Studies: An Overview.
13 812 *Drug Des Devel Ther* **2019**, *13*, 3591–3605. <https://doi.org/10.2147/DDDT.S218708>.
- 16
17 813 (19) Williams-Medina, A.; Deblock, M.; Janigro, D. In Vitro Models of the Blood–Brain
18 814 Barrier: Tools in Translational Medicine. *Front Med Technol* **2021**, *2*.
19 815 <https://doi.org/10.3389/fmedt.2020.623950>.
- 22
23 816 (20) Santana, K.; do Nascimento, L. D.; Lima e Lima, A.; Damasceno, V.; Nahum, C.; Braga,
24 817 R. C.; Lameira, J. Applications of Virtual Screening in Bioprospecting: Facts, Shifts,
25 818 and Perspectives to Explore the Chemo-Structural Diversity of Natural Products. *Front*
26 819 *Chem* **2021**, *9*. <https://doi.org/10.3389/fchem.2021.662688>.
- 30
31 820 (21) Vastag, M.; Keseru, G. M. Current in Vitro and in Silico Models of Blood-Brain Barrier
32 821 Penetration: A Practical View. *Curr Opin Drug Discov Devel* **2009**, *12*(1), 115–124.
- 34
35 822 (22) Martin, I. Prediction of Blood-Brain Barrier Penetration: Are We Missing the Point?
36 823 *Drug Discov Today* **2004**, *9* (4), 161–162. [https://doi.org/10.1016/S1359-](https://doi.org/10.1016/S1359-6446(03)02961-1)
37 824 [6446\(03\)02961-1](https://doi.org/10.1016/S1359-6446(03)02961-1).
- 40
41 825 (23) Bao, W.; Yang, B. Protein Acetylation Sites with Complex-Valued Polynomial Model.
42 826 *Frontiers (Boulder)* **2024**, *18*(3).
- 44
45 827 (24) Bao, W.; Cui, Q.; Chen, B.; Yang, B. Phage_UniR_LGBM: Phage Virion Proteins
46 828 Classification with UniRep Features and LightGBM Model. *Comput Math Methods*
47 829 *Med* **2022**, *2022*, 1–8. <https://doi.org/10.1155/2022/9470683>.
- 50
51 830 (25) de Oliveira, E. C. L.; da Costa, K. S.; Taube, P. S.; Lima, A. H.; Junior, C. de S. de S.
52 831 Biological Membrane-Penetrating Peptides: Computational Prediction and
53 832 Applications. *Front Cell Infect Microbiol* **2022**, *12*.
54 833 <https://doi.org/10.3389/fcimb.2022.838259>.

- 1
2
3
4 834 (26) Shoombuatong, W.; Schaduangrat, N.; Pratiwi, R.; Nantasenamat, C. THPep: A
5 835 Machine Learning-Based Approach for Predicting Tumor Homing Peptides. *Comput*
6 836 *Biol Chem* **2019**, *80*, 441–451. <https://doi.org/10.1016/j.compbiolchem.2019.05.008>.
- 8
9 837 (27) Mishra, A.; Kabir, M. W. U.; Hoque, M. T. DiSBPred: A Machine Learning Based
10 838 Approach for Disulfide Bond Prediction. *Comput Biol Chem* **2021**, *91*, 107436.
11 839 <https://doi.org/10.1016/j.compbiolchem.2021.107436>.
- 14
15 840 (28) Bao, W.; Gu, Y.; Chen, B.; Yu, H. Golgi_DF: Golgi Proteins Classification with Deep
16 841 Forest. *Front Neurosci* **2023**, *17*. <https://doi.org/10.3389/fnins.2023.1197824>.
- 18
19 842 (29) Stalmans, S.; Gevaert, B.; Wynendaele, E.; Nielandt, J.; De Tre, G.; Peremans, K.;
20 843 Burvenich, C.; De Spiegeleer, B. Classification of Peptides According to Their Blood-
21 844 Brain Barrier Influx. *Protein Pept Lett* **2015**, *22* (9), 768–775.
22 845 <https://doi.org/10.2174/0929866522666150622101223>.
- 26
27 846 (30) Ecker, G.; Noe, C. In Silico Prediction Models for Blood-Brain Barrier Permeation. *Curr*
28 847 *Med Chem* **2012**, *11* (12), 1617–1528. <https://doi.org/10.2174/0929867043365071>.
- 30
31 848 (31) Abbott, N. J.; Patabendige, A. A. K.; Dolman, D. E. M.; Yusof, S. R.; Begley, D. J.
32 849 Structure and Function of the Blood–Brain Barrier. *Neurobiol Dis* **2010**, *37*(1), 13–25.
33 850 <https://doi.org/10.1016/j.nbd.2009.07.030>.
- 36
37 851 (32) Mensch, J.; Oyarzabal, J.; Mackie, C.; Augustijns, P. In Vivo, in Vitro and in Silico
38 852 Methods for Small Molecule Transfer across the BBB. *J Pharm Sci* **2009**, *98*(12), 4429–
39 853 4468. <https://doi.org/10.1002/jps.21745>.
- 42
43 854 (33) Konovalov, D. A.; Coomans, D.; Deconinck, E.; Heyden, Y. Vander. Benchmarking of
44 855 QSAR Models for Blood-Brain Barrier Permeation. *J Chem Inf Model* **2007**, *47* (4),
45 856 1648–1656. <https://doi.org/10.1021/ci700100f>.
- 48
49 857 (34) Bendels, S.; Kansy, M.; Wagner, B.; Huwyler, J. In Silico Prediction of Brain and CSF
50 858 Permeation of Small Molecules Using PLS Regression Models. *Eur J Med Chem* **2008**,
51 859 *43* (8), 1581–1592. <https://doi.org/10.1016/j.ejmech.2007.11.011>.
- 54
55 860 (35) Faramarzi, S.; Kim, M. T.; Volpe, D. A.; Cross, K. P.; Chakravarti, S.; Stavitskaya, L.
56 861 Development of QSAR Models to Predict Blood-Brain Barrier Permeability. *Front*
57 862 *Pharmacol* **2022**, *13*. <https://doi.org/10.3389/fphar.2022.1040838>.

- 1
2
3
4 863 (36) Liu, X.; Tu, M.; Kelly, R. S.; Chen, C.; Smith, B. J. Development of a Computational
5 864 Approach to Predict Blood-Brain Barrier Permeability. *Drug Metabolism and*
6 865 *Disposition* **2004**, *32* (1), 132–139. <https://doi.org/10.1124/dmd.32.1.132>.
7
8
9 866 (37) Chen, X.; Zhang, Q.; Li, B.; Lu, C.; Yang, S.; Long, J.; He, B.; Chen, H.; Huang, J.
10 867 BBPpredict: A Web Service for Identifying Blood-Brain Barrier Penetrating Peptides.
11 868 *Front Genet* **2022**, *13*. <https://doi.org/10.3389/fgene.2022.845747>.
12
13
14
15 869 (38) Dai, R.; Zhang, W.; Tang, W.; Wynendaele, E.; Zhu, Q.; Bin, Y.; De Spiegeleer, B.;
16 870 Xia, J. BBPpred: Sequence-Based Prediction of Blood-Brain Barrier Peptides with
17 871 Feature Representation Learning and Logistic Regression. *J Chem Inf Model* **2021**, *61*
18 872 (1), 525–534. <https://doi.org/10.1021/acs.jcim.0c01115>.
19
20
21
22
23 873 (39) Zou, H. Identifying Blood-Brain Barrier Peptides by Using Amino Acids
24 874 Physicochemical Properties and Features Fusion Method. *Peptide Science* **2022**, *114*(2).
25 875 <https://doi.org/10.1002/pep2.24247>.
26
27
28
29 876 (40) Van Dorpe, S.; Bronselaer, A.; Nielandt, J.; Stalmans, S.; Wynendaele, E.; Audenaert,
30 877 K.; Van De Wiele, C.; Burvenich, C.; Peremans, K.; Hsuchou, H.; De Tré, G.; De
31 878 Spiegeleer, B. Brainpeps: The Blood-Brain Barrier Peptide Database. *Brain Struct Funct*
32 879 **2012**, *217*(3), 687–718. <https://doi.org/10.1007/s00429-011-0375-0>.
33
34
35
36 880 (41) Gjedde, A. High- and Low-Affinity Transport of D-Glucose from Blood to Brain. *J*
37 881 *Neurochem* **1981**, *36* (4), 1463–1471. [https://doi.org/10.1111/j.1471-](https://doi.org/10.1111/j.1471-4159.1981.tb00587.x)
38 882 [4159.1981.tb00587.x](https://doi.org/10.1111/j.1471-4159.1981.tb00587.x).
39
40
41
42 883 (42) Poth, A. G.; Chiu, F. C. K.; Stalmans, S.; Hamilton, B. R.; Huang, Y. H.; Shackelford,
43 884 D. M.; Patil, R.; Le, T. T.; Kan, M. W.; Durek, T.; Wynendaele, E.; De Spiegeleer, B.;
44 885 Powell, A. K.; Venter, D. J.; Clark, R. J.; Charman, S. A.; Craik, D. J. Effects of
45 886 Backbone Cyclization on the Pharmacokinetics and Drug Efficiency of the Orally Active
46 887 Analgesic Conotoxin CVc1.1. *Med Drug Discov* **2021**, *10*, 100087.
47 888 <https://doi.org/10.1016/j.medidd.2021.100087>.
48
49
50
51
52
53 889 (43) Van Dorpe, S.; Bronselaer, A.; Nielandt, J.; Stalmans, S.; Wynendaele, E.; Audenaert,
54 890 K.; Van De Wiele, C.; Burvenich, C.; Peremans, K.; Hsuchou, H.; De Tré, G.; De
55 891 Spiegeleer, B. Brainpeps: The Blood-Brain Barrier Peptide Database. *Brain Struct Funct*
56 892 **2012**, *217*(3), 687–718. <https://doi.org/10.1007/s00429-011-0375-0>.
57
58
59
60

- 1
2
3
4 893 (44) Janssens, Y.; Debunne, N.; De Spiegeleer, A.; Wynendaele, E.; Planas, M.; Feliu, L.;
5 894 Quarta, A.; Claes, C.; Van Dam, D.; De Deyn, P. P.; Ponsaerts, P.; Blurton-Jones, M.;
6 895 De Spiegeleer, B. PapRIV, a BV-2 Microglial Cell Activating Quorum Sensing Peptide.
7 896 *Sci Rep* **2021**, *11* (1), 10723. <https://doi.org/10.1038/s41598-021-90030-y>.
- 10
11 897 (45) Abbruscato, T. J.; Williams, S. A.; Misicka, A.; Lipkowski, A. W.; Hruby, V. J.; Davis,
12 898 T. P. Blood-to-Central Nervous System Entry and Stability of Biphalin, a Unique
13 899 Double-Enkephalin Analog, and Its Halogenated Derivatives. *Journal of Pharmacology*
14 900 *and Experimental Therapeutics* **1996**, *276* (3), 1049–1057.
- 17
18 901 (46) Di, L.; Kerns, E. H.; Fan, K.; McConnell, O. J.; Carter, G. T. High Throughput Artificial
19 902 Membrane Permeability Assay for Blood-Brain Barrier. *Eur J Med Chem* **2003**, *38* (3),
20 903 223–232. [https://doi.org/10.1016/S0223-5234\(03\)00012-6](https://doi.org/10.1016/S0223-5234(03)00012-6).
- 23
24 904 (47) Zorbaz, T.; Braïki, A.; Maraković, N.; Renou, J.; de la Mora, E.; Maček Hrvat, N.;
25 905 Katalinić, M.; Silman, I.; Sussman, J. L.; Mercey, G.; Gomez, C.; Mougeot, R.; Pérez,
26 906 B.; Baati, R.; Nachon, F.; Weik, M.; Jean, L.; Kovarik, Z.; Renard, P. Y. Potent 3-
27 907 Hydroxy-2-Pyridine Aldoxime Reactivators of Organophosphate-Inhibited
28 908 Cholinesterases with Predicted Blood–Brain Barrier Penetration. *Chemistry - A*
29 909 *European Journal* **2018**, *24* (38), 9675–9691. <https://doi.org/10.1002/chem.201801394>.
- 32
33 910 (48) Rossi, M.; Freschi, M.; De Camargo Nascente, L.; Salerno, A.; De Melo Viana Teixeira,
34 911 S.; Nachon, F.; Chantegreil, F.; Soukup, O.; Prchal, L.; Malaguti, M.; Bergamini, C.;
35 912 Bartolini, M.; Angeloni, C.; Hrelia, S.; Soares Romeiro, L. A.; Bolognesi, M. L.
36 913 Sustainable Drug Discovery of Multi-Target-Directed Ligands for Alzheimer’s Disease.
37 914 *J Med Chem* **2021**, *64* (8), 4972–4990. <https://doi.org/10.1021/acs.jmedchem.1c00048>.
- 40
41 915 (49) Felegyi-Tóth, C. A.; Tóth, Z.; Garádi, Z.; Boldizsár, I.; Nedves, A. N.; Simon, A.;
42 916 Felegyi, K.; Alberti, Á.; Riethmüller, E. Membrane Permeability and Aqueous Stability
43 917 Study of Linear and Cyclic Diarylheptanoids from *Corylus Maxima*. *Pharmaceutics*
44 918 **2022**, *14* (6), 1250. <https://doi.org/10.3390/pharmaceutics14061250>.
- 47
48 919 (50) Simon, A.; Darcsi, A.; Kéry, Á.; Riethmüller, E. Blood-Brain Barrier Permeability
49 920 Study of Ginger Constituents. *J Pharm Biomed Anal* **2020**, *177*, 112820.
50 921 <https://doi.org/10.1016/j.jpba.2019.112820>.

- 1
2
3
4 922 (51) Garberg, P.; Ball, M.; Borg, N.; Cecchelli, R.; Fenart, L.; Hurst, R. D.; Lindmark, T.;
5 923 Mabondzo, A.; Nilsson, J. E.; Raub, T. J.; Stanimirovic, D.; Terasaki, T.; Öberg, J. O.;
6 924 Österberg, T. In Vitro Models for the Blood-Brain Barrier. *Toxicology in Vitro* **2005**,
7 925 *19*(3), 299–334. <https://doi.org/10.1016/j.tiv.2004.06.011>.
- 10
11 926 (52) Yoon, C. H.; Kim, S. J.; Shin, B. S.; Lee, K. C.; Yoo, S. D. Rapid Screening of Blood-
12 927 Brain Barrier Penetration of Drugs Using the Immobilized Artificial Membrane
13 928 Phosphatidylcholine Column Chromatography. *J Biomol Screen* **2006**, *11* (1), 13–20.
14 929 <https://doi.org/10.1177/1087057105281656>.
- 18
19 930 (53) Dichiaro, M.; Amata, B.; Turnaturi, R.; Marrazzo, A.; Amata, E. Tuning Properties for
20 931 Blood-Brain Barrier Permeation: A Statistics-Based Analysis. *ACS Chem Neurosci*
21 932 **2020**, *11* (1), 34–44. <https://doi.org/10.1021/acscemneuro.9b00541>.
- 24
25 933 (54) Mikitsh, J. L.; Chacko, A. M. Pathways for Small Molecule Delivery to the Central
26 934 Nervous System across the Blood-Brain Barrier. *Perspect Medicin Chem* **2014**, *6* (6),
27 935 11–24. <https://doi.org/10.4137/PMc.s13384>.
- 30
31 936 (55) Fong, C. W. Permeability of the Blood–Brain Barrier: Molecular Mechanism of
32 937 Transport of Drugs and Physiologically Important Compounds. *Journal of Membrane*
33 938 *Biology* **2015**, *248* (4), 651–669. <https://doi.org/10.1007/s00232-015-9778-9>.
- 36
37 939 (56) Lovering, F.; Bikker, J.; Humblet, C. Escape from Flatland: Increasing Saturation as an
38 940 Approach to Improving Clinical Success. *J Med Chem* **2009**, *52* (21), 6752–6756.
39 941 <https://doi.org/10.1021/jm901241e>.
- 42
43 942 (57) Lipinski, C. A. C. A. C. A.; Lombardo, F.; Dominy, B. W. B. W. B. W. B. W. B. W.;
44 943 Feeney, P. J. P. J.; Lombardo, F.; Dominy, B. W. B. W. B. W. B. W. B. W.; Feeney, P.
45 944 J. P. J. Experimental and Computational Approaches to Estimate Solubility and
46 945 Permeability in Drug Discovery and Development Settings. *Adv Drug Deliv Rev* **1997**,
47 946 *23*(1–3), 3–25. [https://doi.org/10.1016/S0169-409X\(96\)00423-1](https://doi.org/10.1016/S0169-409X(96)00423-1).
- 51
52 947 (58) Matsson, P.; Doak, B. C.; Over, B.; Kihlberg, J. Cell Permeability beyond the Rule of
53 948 *5*. *Adv Drug Deliv Rev* **2016**, *101*, 42–61. <https://doi.org/10.1016/j.addr.2016.03.013>.
- 54
55
56
57
58
59
60

- 1
2
3
4 949 (59) Doak, B. C.; Over, B.; Giordanetto, F.; Kihlberg, J. Oral Druggable Space beyond the
5 950 Rule of 5: Insights from Drugs and Clinical Candidates. *Chem Biol* **2014**, *21* (9), 1115–
6 951 1142. <https://doi.org/10.1016/j.chembiol.2014.08.013>.
- 7
8
9 952 (60) Moriwaki, H.; Tian, Y. S.; Kawashita, N.; Takagi, T. Mordred: A Molecular Descriptor
10 953 Calculator. *J Cheminform* **2018**, *10*(1), 4. <https://doi.org/10.1186/s13321-018-0258-y>.
- 11
12
13 954 (61) Raducanu, B.; Dornaika, F. A Supervised Non-Linear Dimensionality Reduction
14 955 Approach for Manifold Learning. *Pattern Recognit* **2012**, *45* (6), 2432–2444.
15 956 <https://doi.org/10.1016/j.patcog.2011.12.006>.
- 16
17
18 957 (62) Gálvez, J.; Garcia, R.; Salabert, M. T.; Soler, R. Charge Indexes. New Topological
19 958 Descriptors. *J Chem Inf Comput Sci* **1994**, *34* (3), 520–525.
20 959 <https://doi.org/10.1021/ci00019a008>.
- 21
22
23 960 (63) Reddy, A. S.; Kumar, S.; Garg, R. Hybrid-Genetic Algorithm Based Descriptor
24 961 Optimization and QSAR Models for Predicting the Biological Activity of Tipranavir
25 962 Analogs for HIV Protease Inhibition. *J Mol Graph Model* **2010**, *28* (8), 852–862.
26 963 <https://doi.org/10.1016/j.jm gm.2010.03.005>.
- 27
28
29 964 (64) Todeschini, R.; Consonni, V. *Handbook of Molecular Descriptors; Methods and*
30 965 *Principles in Medicinal Chemistry; Wiley, 2000; Vol. 11.*
31 966 <https://doi.org/10.1002/9783527613106>.
- 32
33
34 967 (65) Bayat, Z.; Movaffagh, J.; Noruzi, S. Development of a Computational Approach to
35 968 Predict Blood-Brain Permeability on Anti-Viral Nucleoside Analogues. *Russian Journal*
36 969 *of Physical Chemistry A* **2011**, *85* (11), 1923–1930.
37 970 <https://doi.org/10.1134/S0036024411110021>.
- 38
39
40 971 (66) Ghose, A. K.; Viswanadhan, V. N.; Wendoloski, J. J. A Knowledge-Based Approach in
41 972 Designing Combinatorial or Medicinal Chemistry Libraries for Drug Discovery. 1. A
42 973 Qualitative and Quantitative Characterization of Known Drug Databases. *J Comb Chem*
43 974 **1999**, *1* (1), 55–68. <https://doi.org/10.1021/cc9800071>.
- 44
45
46 975 (67) Sohil, F.; Sohali, M. U.; Shabbir, J. *An Introduction to Statistical Learning with*
47 976 *Applications in R; 2022; Vol. 6.* <https://doi.org/10.1080/24754269.2021.1980261>.
- 48
49
50
51
52
53
54
55
56
57
58
59
60

- 1
2
3
4 977 (68) Dai, R.; Zhang, W.; Tang, W.; Wynendaele, E.; Zhu, Q.; Bin, Y.; De Spiegeleer, B.;
5 978 Xia, J. BBPpred: Sequence-Based Prediction of Blood-Brain Barrier Peptides with
6
7 979 Feature Representation Learning and Logistic Regression. *J Chem Inf Model* **2021**, *61*
8
9 980 (1), 525–534. <https://doi.org/10.1021/acs.jcim.0c01115>.
- 10
11 981 (69) Kumar, V.; Patiyal, S.; Dhall, A.; Sharma, N.; Raghava, G. P. S. B3pred: A Random-
12
13 982 Forest-Based Method for Predicting and Designing Blood–Brain Barrier Penetrating
14
15 983 Peptides. *Pharmaceutics* **2021**, *13* (8), 1237.
16 984 <https://doi.org/10.3390/pharmaceutics13081237>.
- 17
18
19 985 (70) Charoenkwan, P.; Chumnanpuen, P.; Schaduangrat, N.; Lio', P.; Moni, M. A.;
20 986 Shoombuatong, W. Improved Prediction and Characterization of Blood-Brain Barrier
21
22 987 Penetrating Peptides Using Estimated Propensity Scores of Dipeptides. *J Comput Aided*
23
24 988 *Mol Des* **2022**, *36* (11), 781–796. <https://doi.org/10.1007/s10822-022-00476-z>.

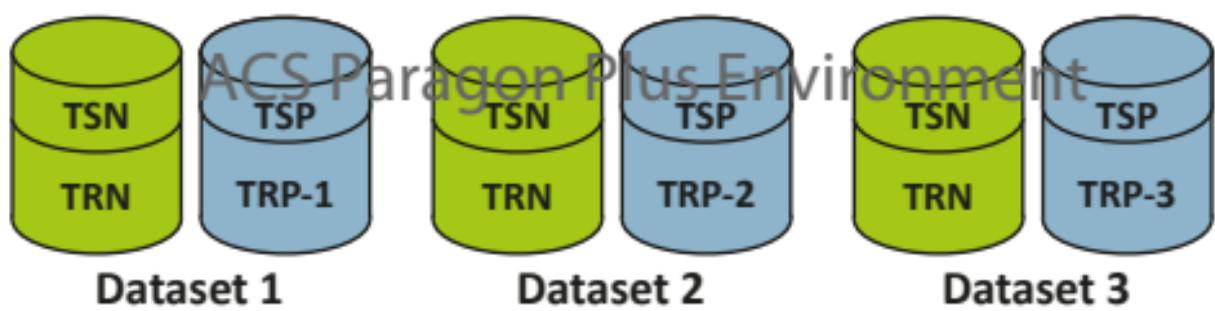
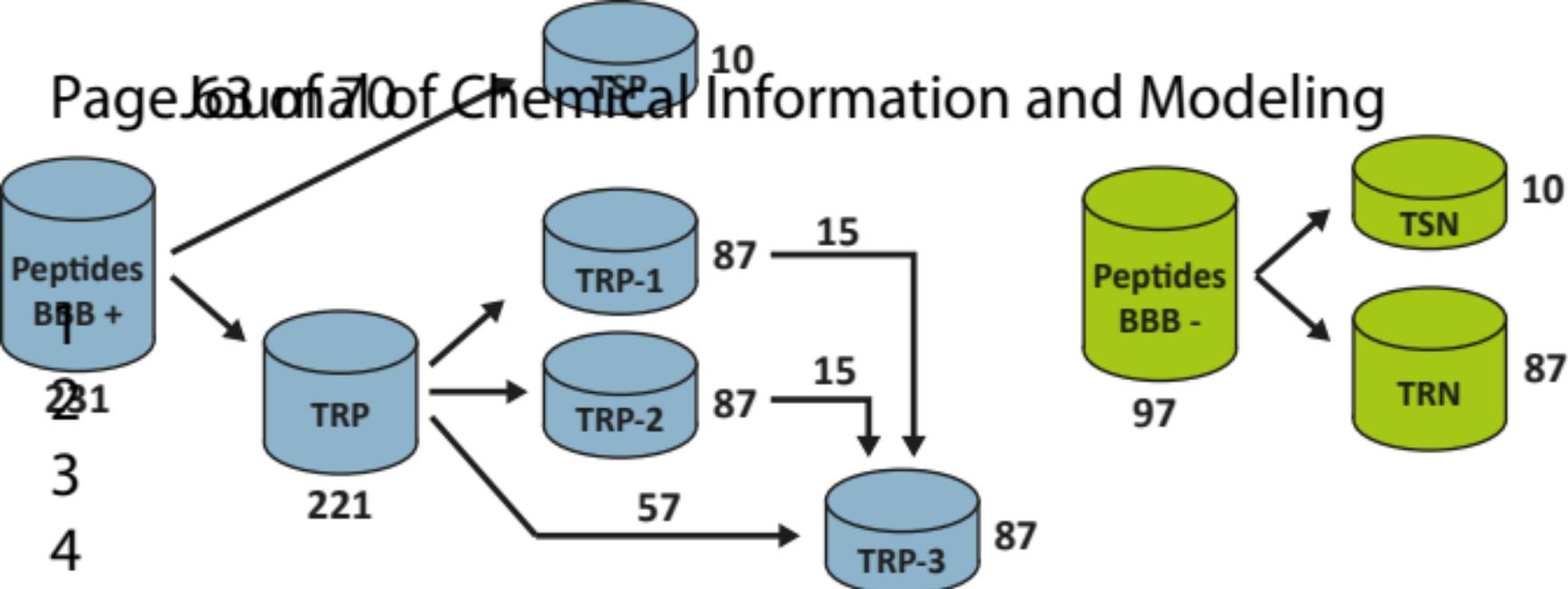
989

990

991

992

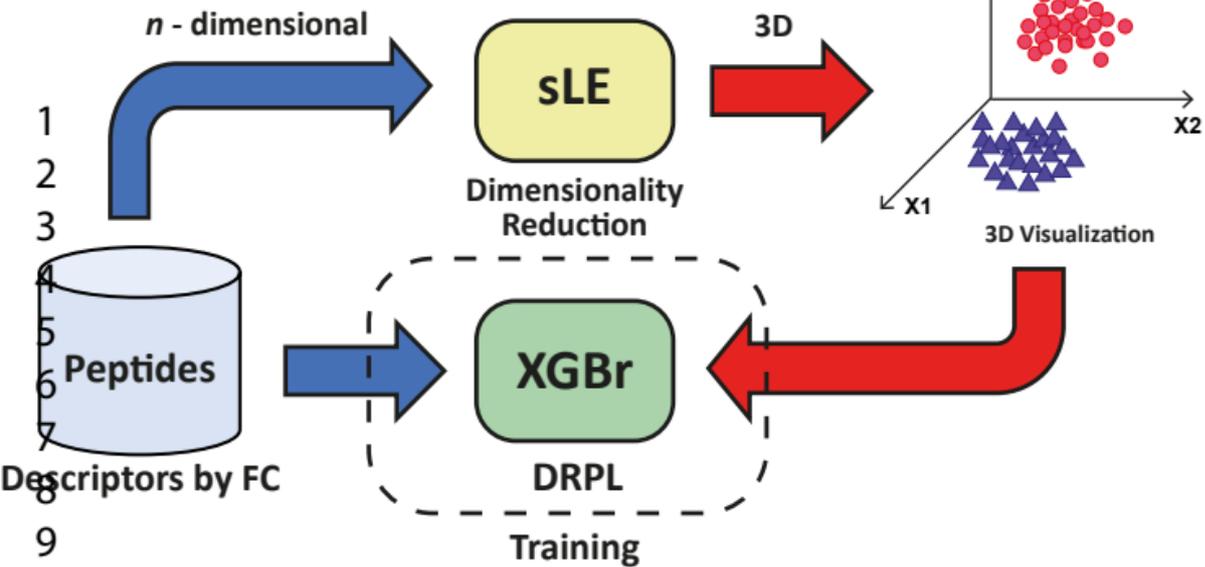
993



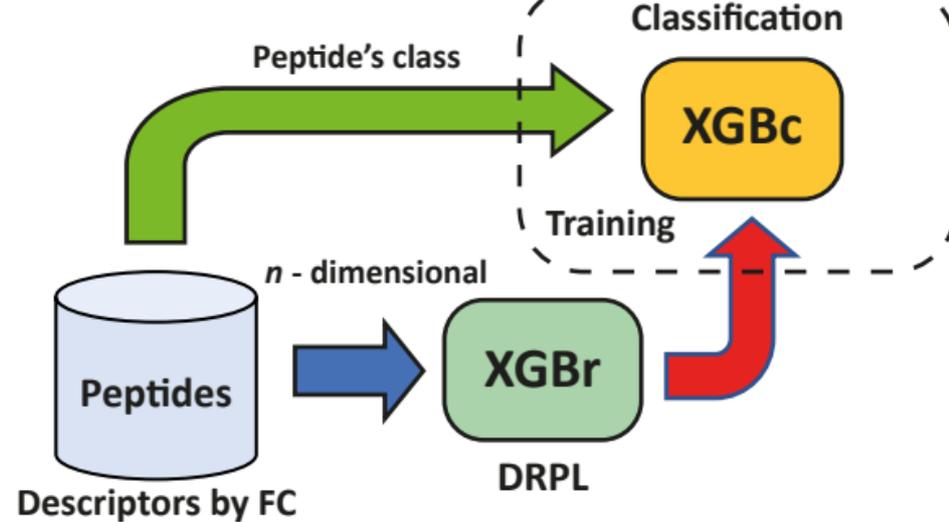
2
3
4
5
6
7
8
9

ACS Paragon Plus Environment

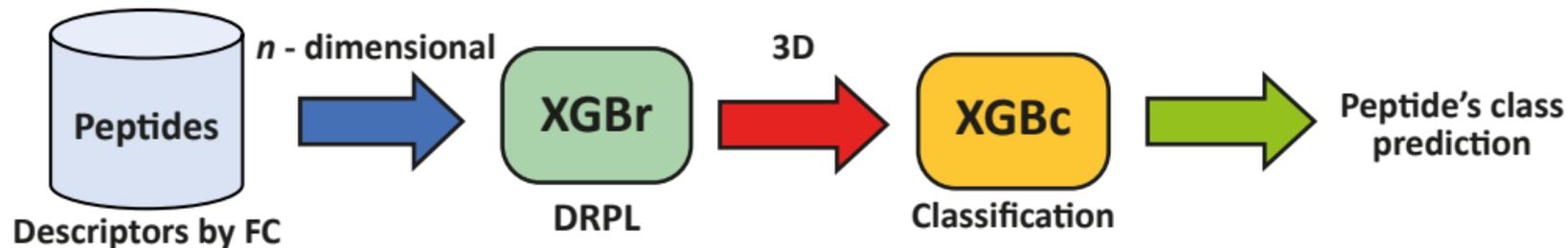
(a)



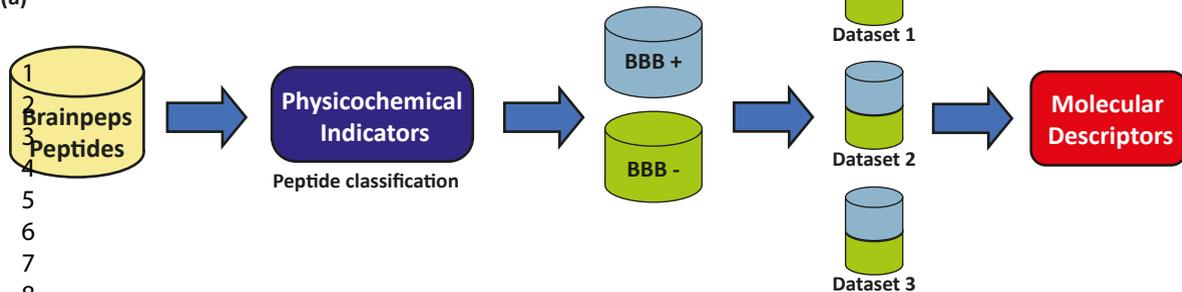
(b)



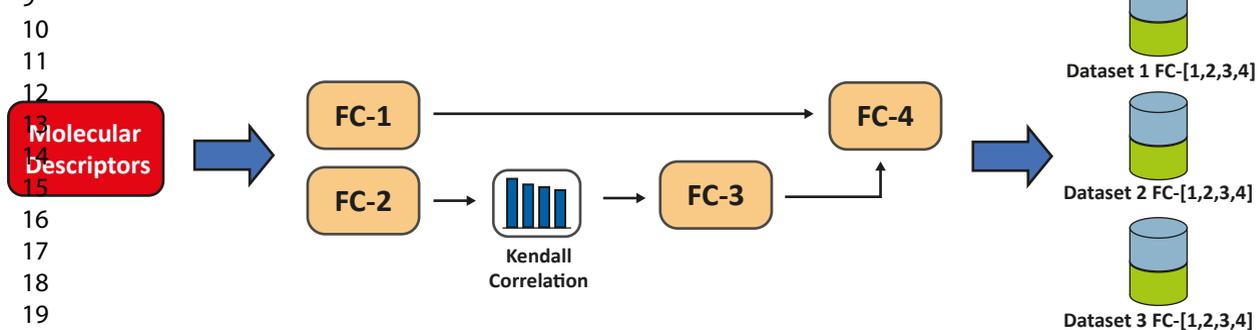
(c)



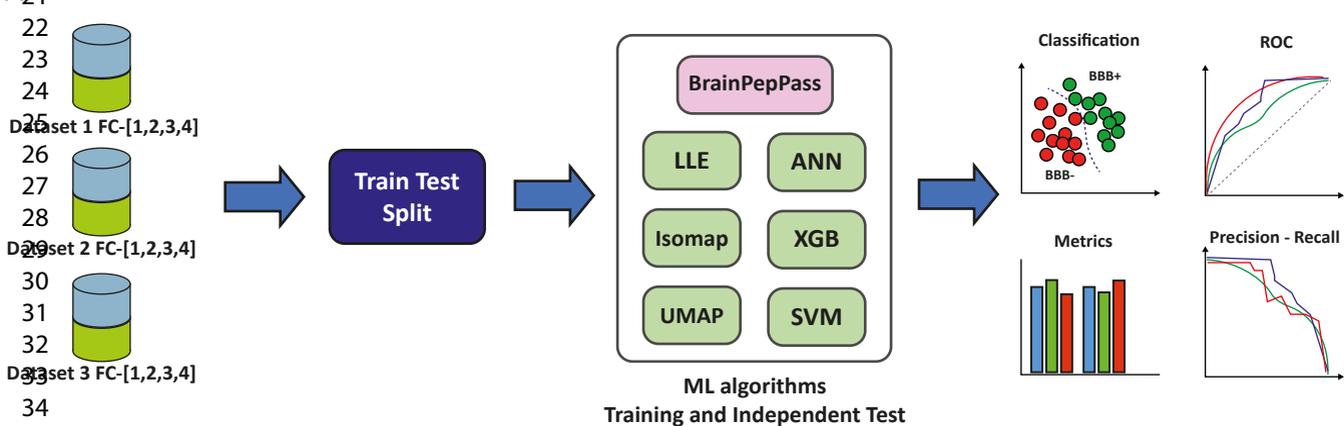
(a)



(b)

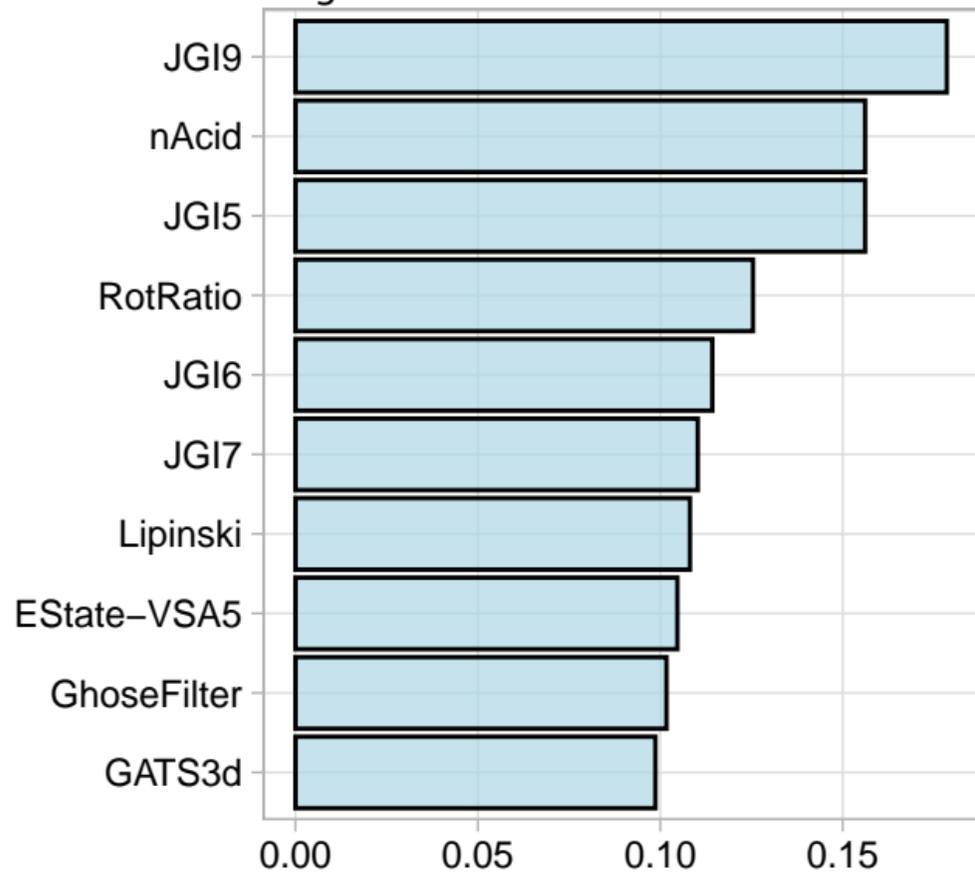
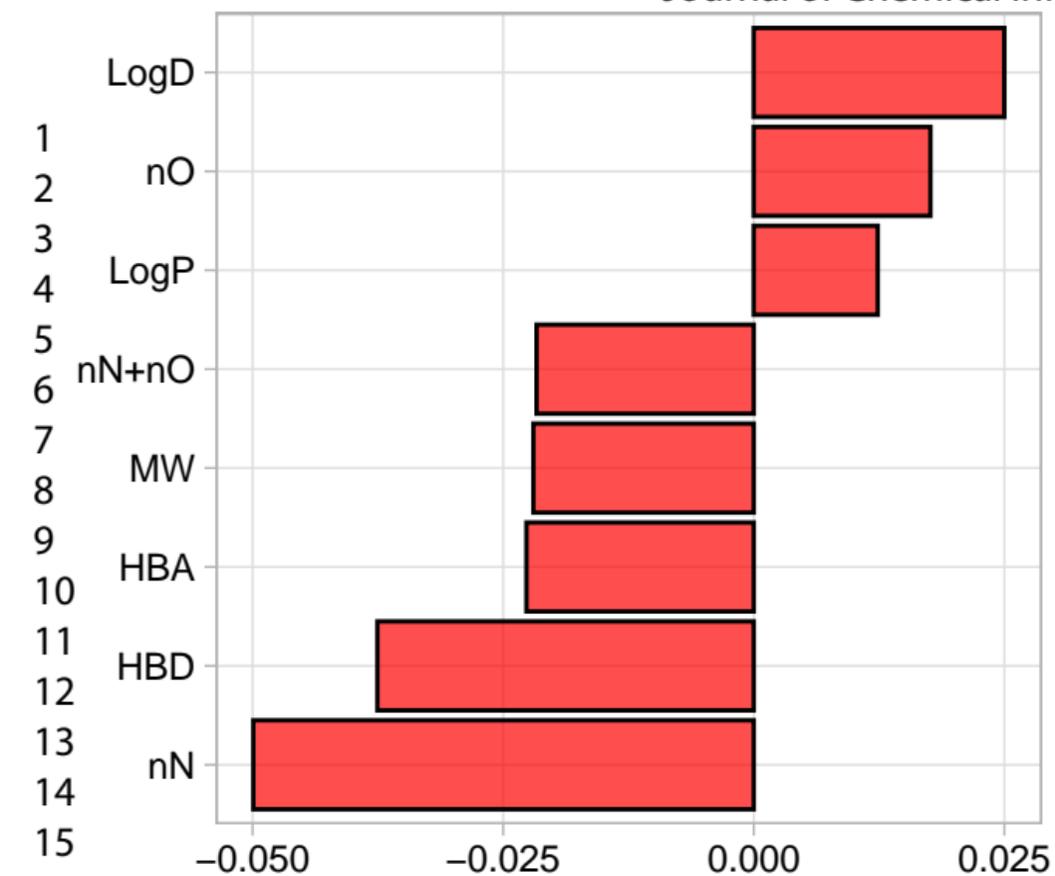


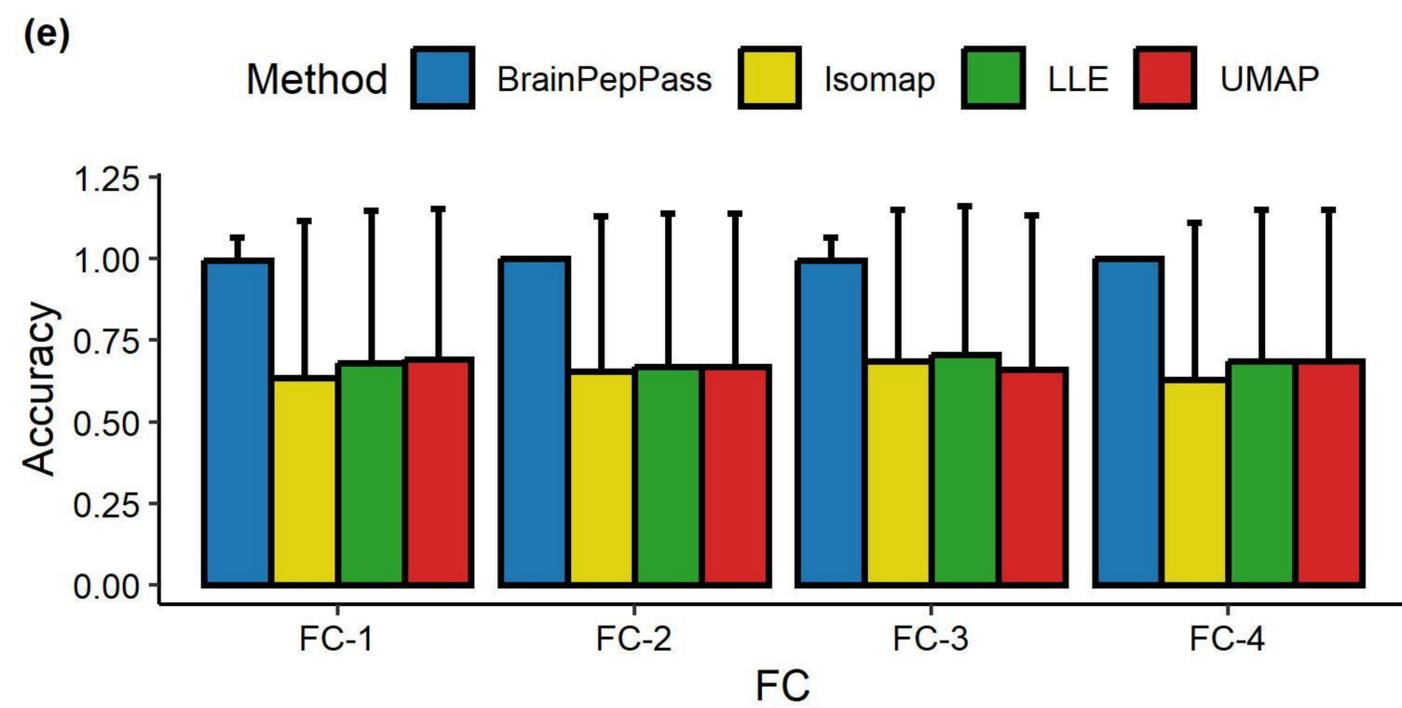
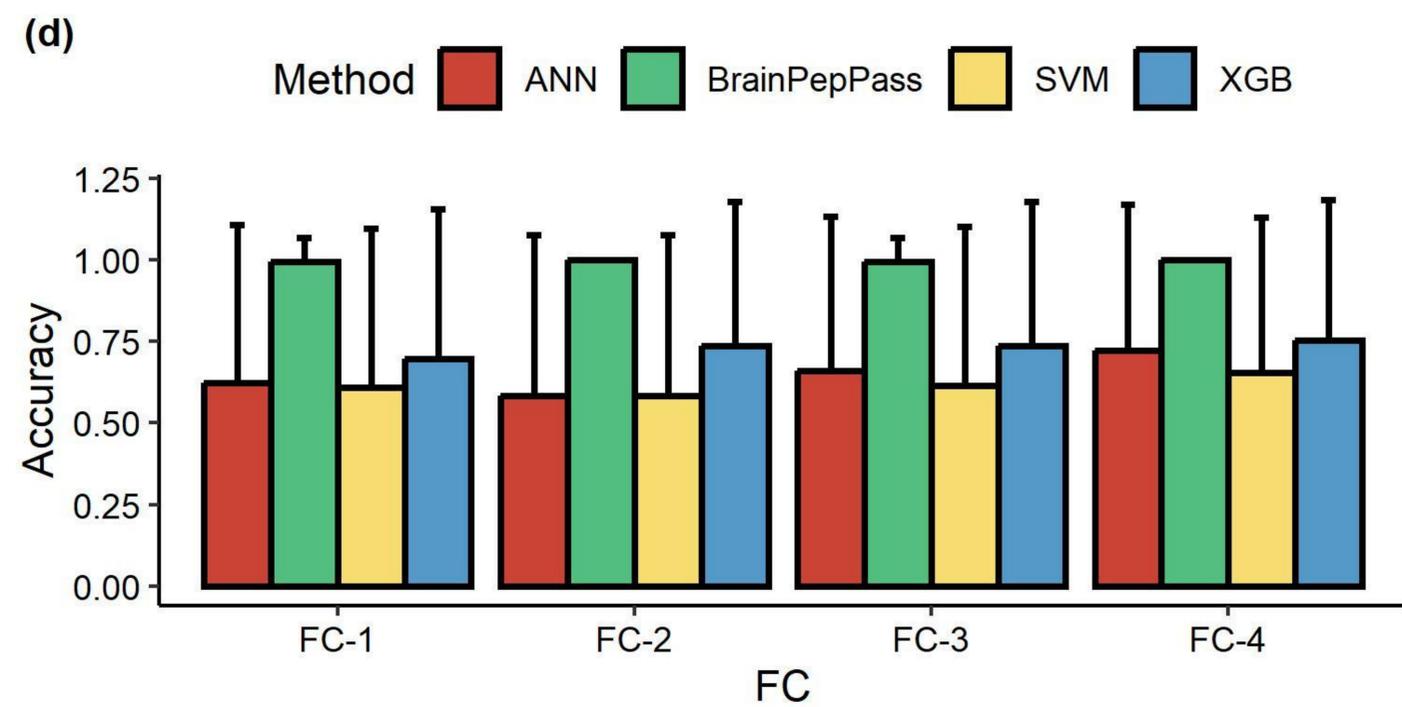
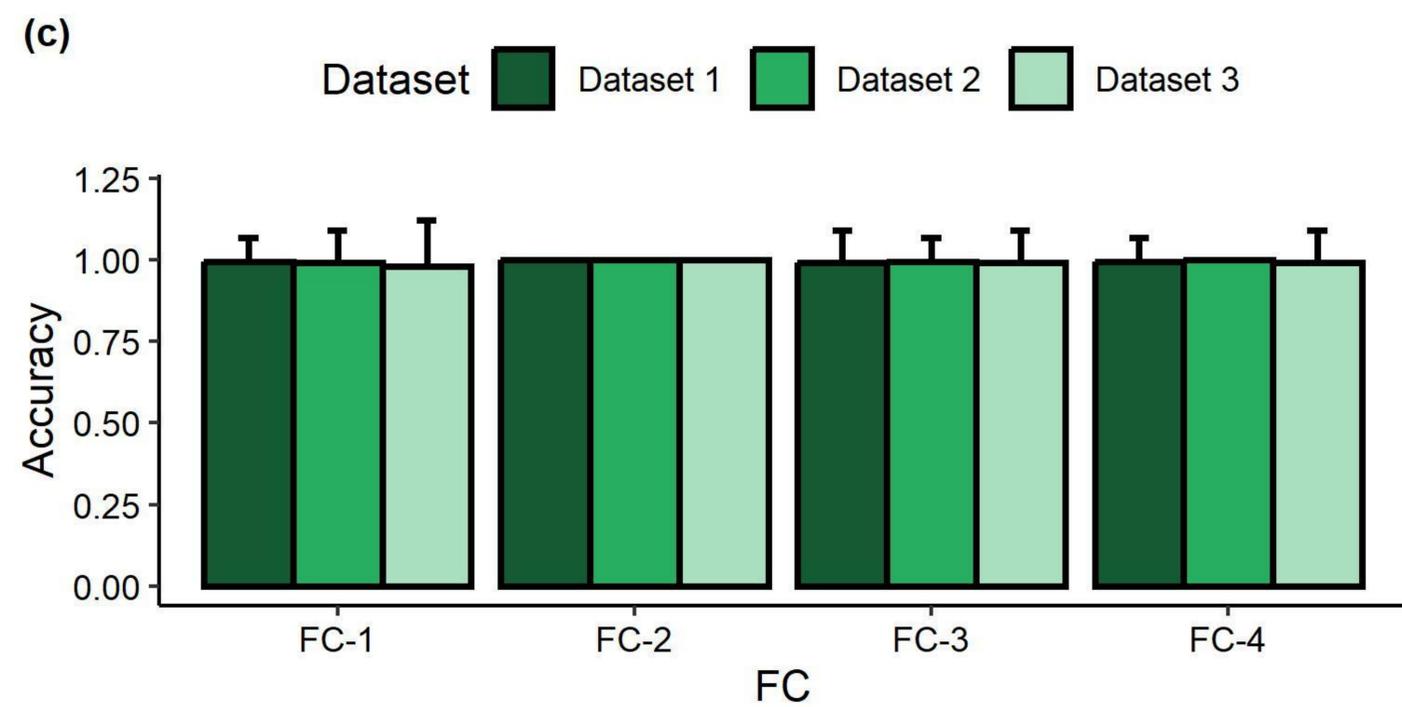
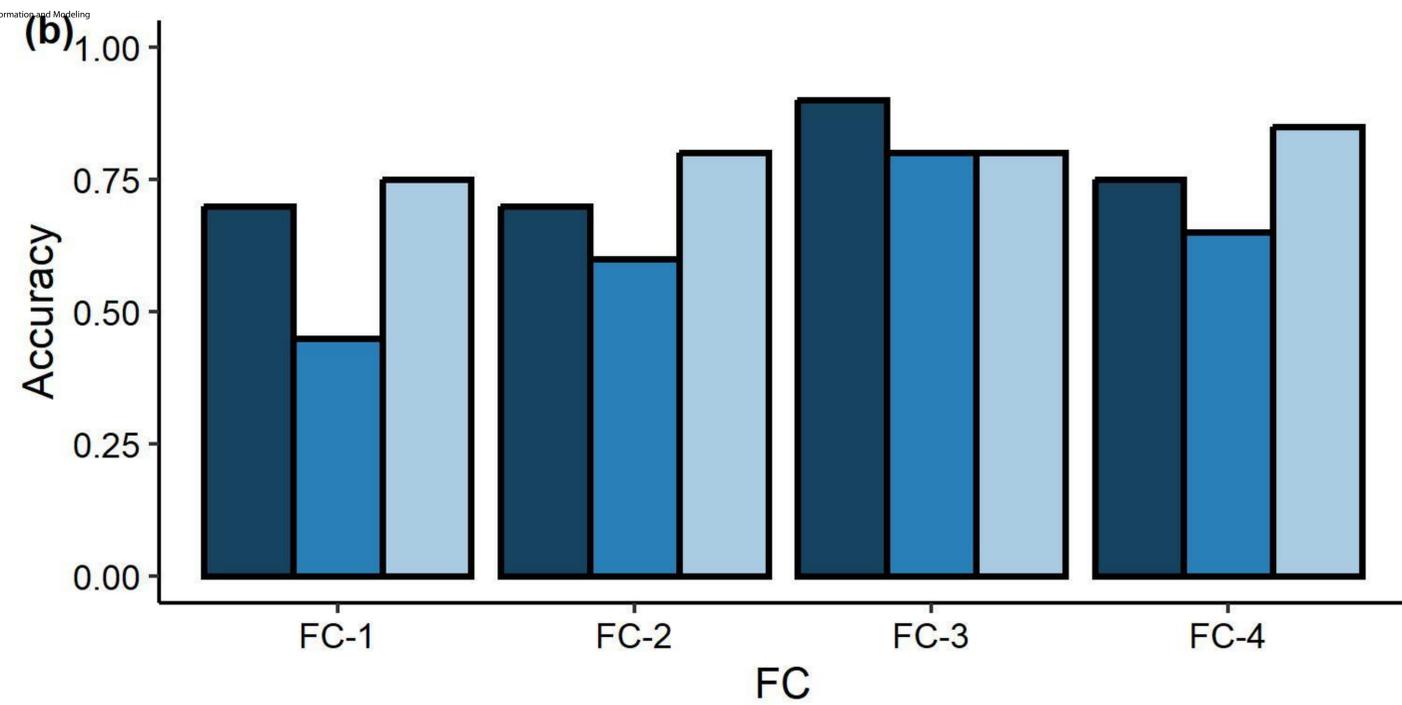
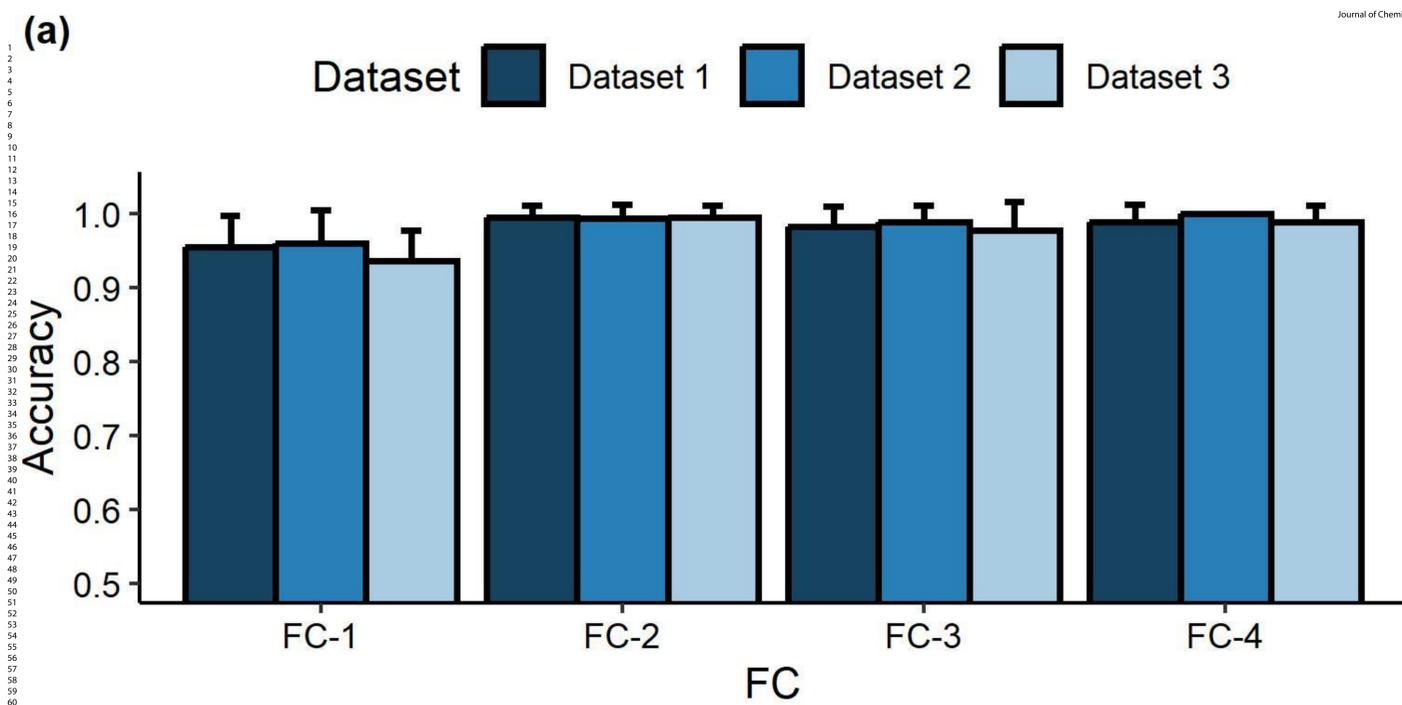
(c)

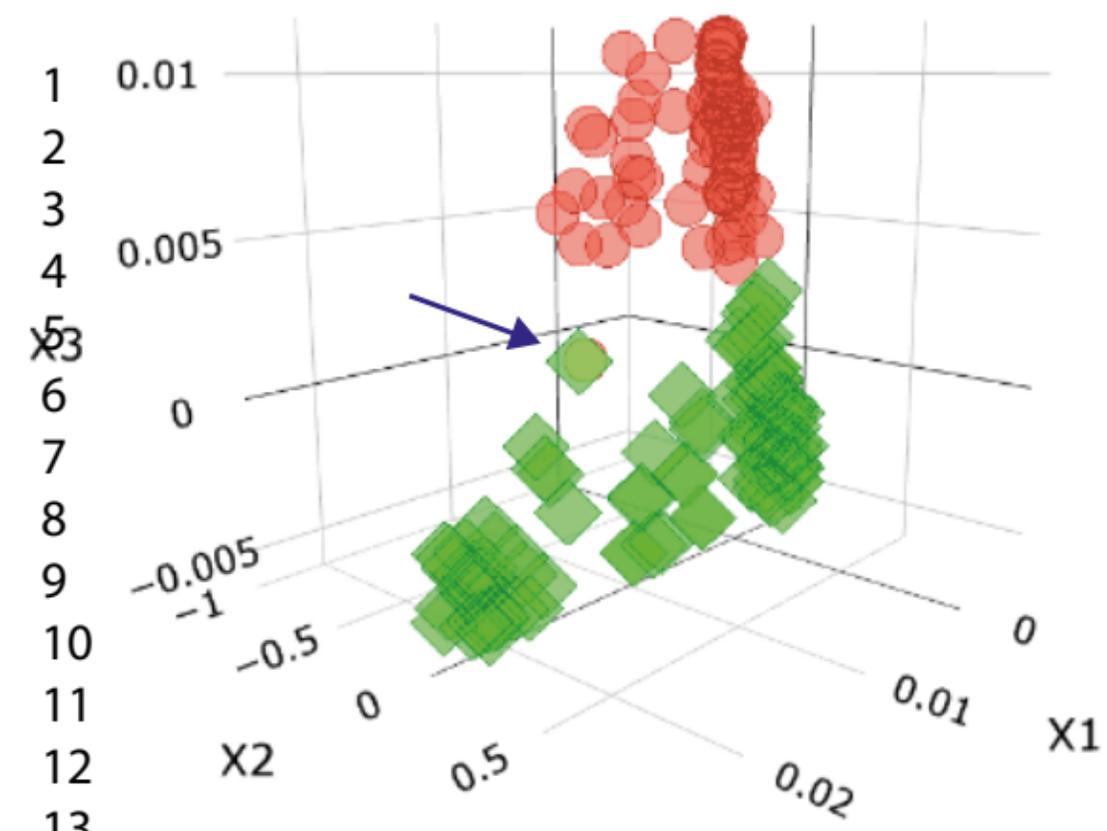


(b)

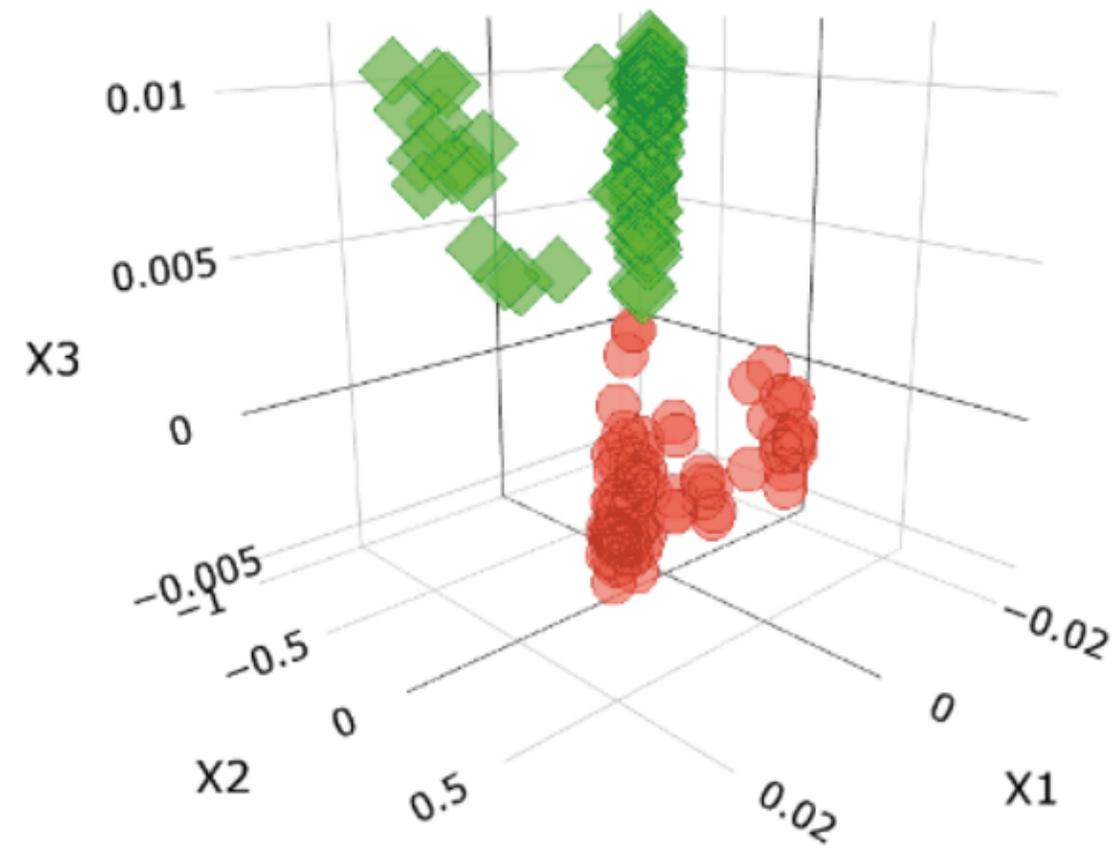
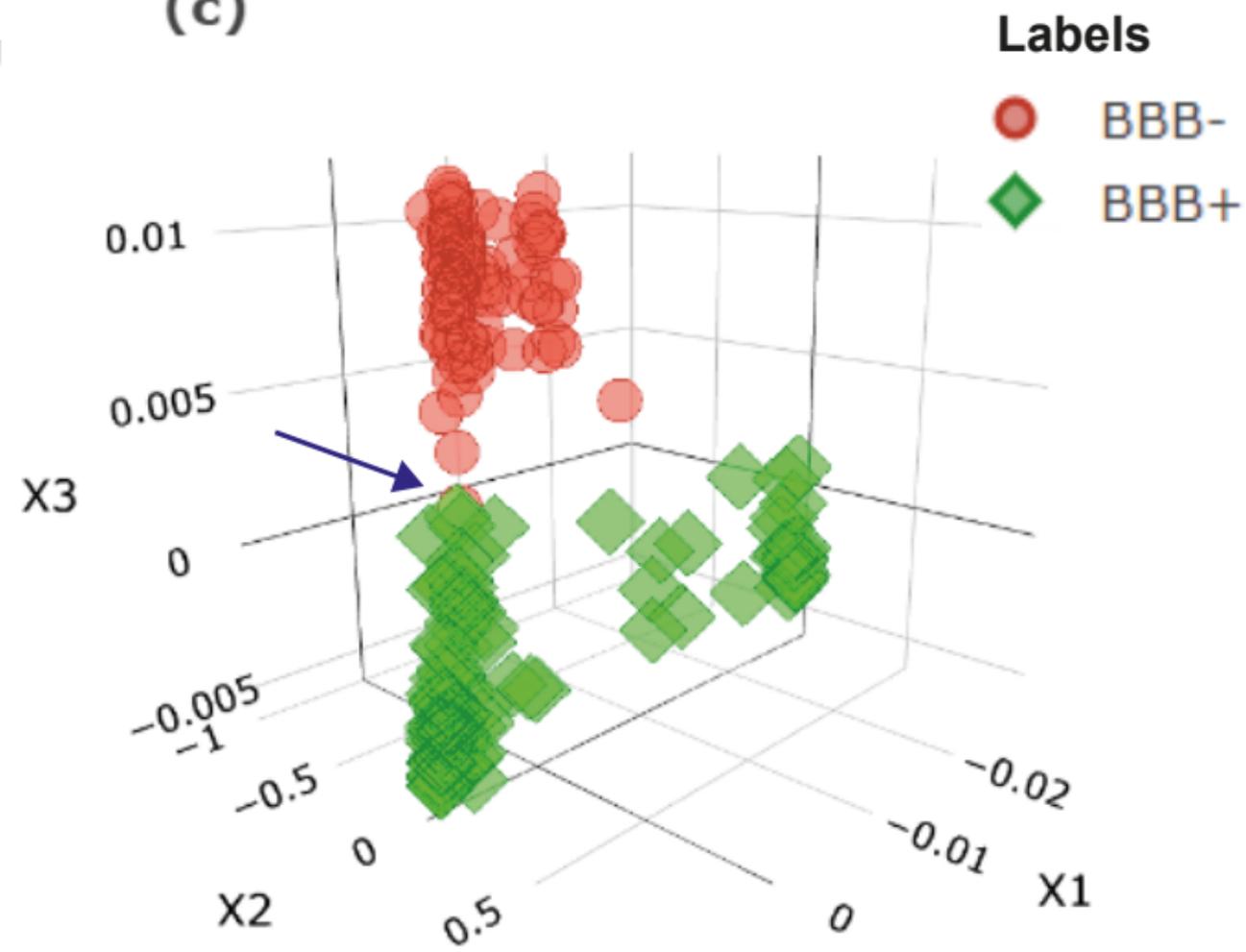
Journal of Chemical Information and Modeling

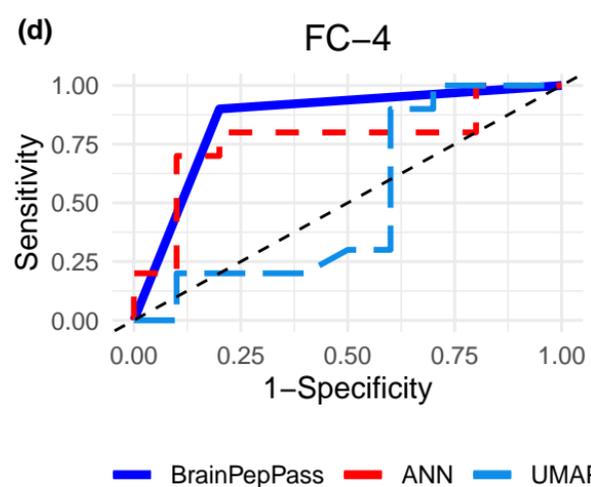
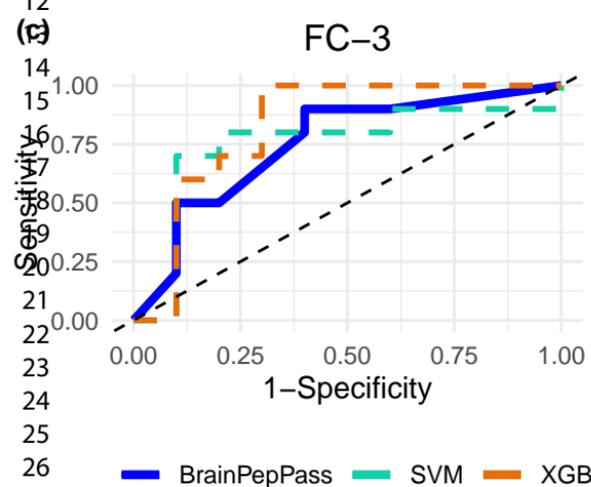
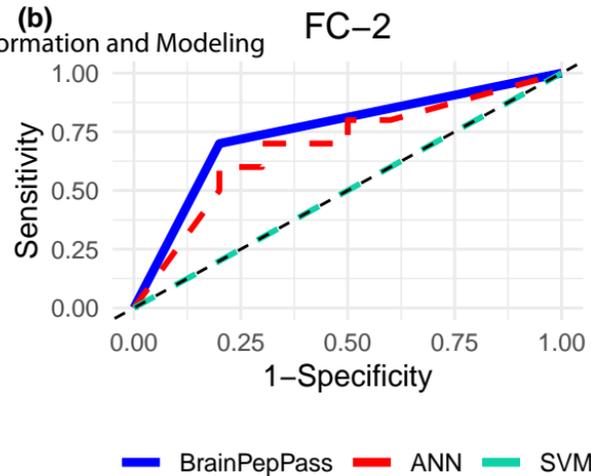
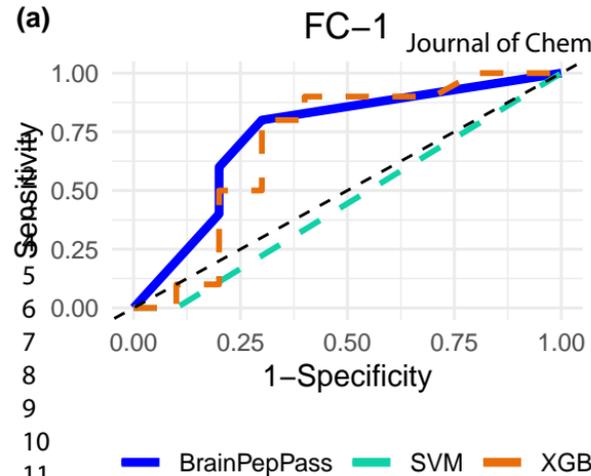
**(a)**



(a)**(b)**

Journal of Chemical Information and Modeling

**(c)**



5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28

