

1 ALLELOPATHY IN MACROALGAE: ECOLOGICAL PRINCIPLES, RESEARCH
2 OPPORTUNITIES AND PITFALLS REVIEWED

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12 Abstract

13 Macroalgae are a vast source of bioactive metabolites, some of which are produced as a defence
14 against competing and/or fouling species, i.e., allelochemicals. As both the occurrence of
15 allelopathy in seaweed, and its underlying mechanisms, are understudied, we summarised the
16 current knowledge on this phenomenon, as well as to elucidate opportunities and challenges in
17 this emerging field. We mark out which biotic and environmental factors govern the stability
18 and durability of allelopathic interactions, and which of them might confound conclusions on
19 the absence/presence of allelopathy in macroalgae. We compiled information of the taxonomic
20 position of 138 putative allelopathic species of brown, green, and red seaweed, as well as the
21 identity of the compounds they produce. Additionally, we characterise which physiological
22 processes are likely targeted by aquatic allelochemicals. In summary, this work updates our
23 understanding of both the occurrence of allelopathy in seaweed on a global scale as their

24 allelochemicals affecting competitors, bringing forth recommendations for future research on
25 this topic.

26 Keywords: allelopathy; allelochemicals; chemical defence; biodiscovery marine ecology;
27 species interactions; marine ecology; antifouling

28 1. Introduction

29 In the euphotic zone, competition among sessile photoautotrophic organisms for space, light
30 and nutrients is particularly fierce and acute (Chadwick and Morrow, 2011; Dayton, 1971).
31 Competition has been identified as a pivotal process shaping the structure and functioning of
32 communities and ecosystems, as well has impacted the ecology and evolution of species
33 (Chadwick and Morrow, 2011; Dayton, 1971; Sousa, 1979). In coral reefs for example, coral-
34 macroalgal competition can be a significant factor of the biodiversity, structure, and functioning
35 of the benthic community (e.g., McCook et al., 2001; Miller, 1998). Moreover, while competing
36 for space, seaweeds themselves become a substrate for colonisation by other organisms, ranging
37 from prokaryotes to animals, whose species richness is also known to climax in the photic zone
38 (da Gama et al., 2014). Surface ‘clotting’ by fouling organisms is undesirable due to its potential
39 to reduce algal fitness, by (1) tissue damage by mechanical anchoring and thallus penetration,
40 (2) affecting nutrient and gas exchanges that take place through the mostly undifferentiated
41 thallus surface, (3) affecting photosynthetic activity, and (4) affecting reproduction as the
42 reproductive stages from macroalgae, such as gametes and different types of spores, are usually
43 released from the seaweed’s surface (da Gama et al., 2014; Gonzalez & Goff, 1989). Indirect
44 effects include an increased drag with consequent tissue loss during extreme weather events
45 and advanced tissue damage due to organisms grazing on the epibionts (the so called ‘shared
46 doom’ effect, proposed by Dixon et al., 1981). As a consequence, some macroalgal taxa have
47 evolved to respond to herbivory and competition by synthesizing “a pool of defensive

48 metabolites that can vary among individuals within a population, but also temporally and/or
49 spatially” (e.g., Andras et al., 2012; Pezolesi et al., 2021; Rasher and Hay, 2010; Rasher et al.,
50 2012). Allelopathy, as initially introduced by Molisch (1937), was defined as “the biochemical
51 interaction, both stimulatory and inhibitory, between primary producers and/or
52 microorganisms” (Table 1). The term itself stems from the Greek *allelon* (of one another) and
53 *pathos* (suffering), mainly focusing on the deleterious effects from plants (Harlin and Rice,
54 1987). There has been considerable discussion of the concept and definition of allelopathy in
55 the past (Muller, 1969; IAS, 1996; Rice, 1984). Nowadays, in literature, allelopathy is
56 predominantly regarded to be a distinct strategy to either eliminate or to deter coexisting
57 competitors and/or predators (e.g., Budzałek et al., 2021; Gomes et al., 2017; Śliwińska-
58 Wilczewska et al., 2021). In this review, we will interpret the term allelopathy in the light of
59 the negative effect(s) of bioactive compounds secreted by macroalgae to target other
60 macroalgae/other competing organisms.

61 In aquatic ecosystems, so far, allelopathy has been investigated more (profoundly) in freshwater
62 habitats compared to the marine environment (Ben Gharbia et al., 2017; Erhard, 2006; Macías
63 et al., 2008). Nevertheless, as of late, research on allelopathy in macroalgae has increased
64 profoundly. Therefore, Budzałek et al. (2021) reviewed allelopathy in macroalgae and its
65 repercussions for coexisting animals, mainly focusing on herbivory, but animal competitors
66 were included as well. Budzałek et al. (2021) mainly focused on listing the allelopathic
67 macroalgae species, their distribution, and the affected animal species, excluding other
68 potentially affected evolutionary groups. While their review touched upon the identity of some
69 of the allelochemicals, a profound review on these compounds, their mode of action, as well as
70 their production, is currently missing. Hence, the aim of the current article is to put forward the
71 ecological roles and chemical diversity of (putative) allelochemicals produced/secreted by
72 macroalgae in marine environments that affect the full range of aquatic competitors (but

73 excluding herbivory). We aimed to (1) characterize the diversity of (putative) allelopathic
 74 macroalgae and their target organisms, (2) outline the temporal and geographical distribution
 75 of publications studying allelopathy in seaweed, (3) summarize the incentives/relevance of
 76 studying allelopathy in seaweed, including their economic potential, (4) describe the commonly
 77 used methods for the study of macroalgal allelopathy, as well as their limitations, (5) compile
 78 information on the identity of the putative allelopathic compounds and their potential mode of
 79 action, and (6) to obtain a better understanding of the influence of biotic and abiotic parameters
 80 influencing allelopathy production and/or sensitivity.

81 Table 1: Terminology and definitions used throughout the text

Biochemical compound	A carbon-based compound found in living organisms
Bioactive compound:	A substance with a biological activity or with the ability to modify or regulate one or more metabolic processes in a living organism, tissue or cell
Algicidal compound	A compound killing or preventing the growth of algae
Allelochemical	Bioactive compound, produced by either plants, macro/microalgae, bacteria, or fungi that impact the growth, physiology and/or development of other organisms
Allelopathic compound	Synonym of allelochemical
Allelopathy	Any process in which bioactive metabolites secreted by an organism affect the development of another species
Antifoulant	A substance that prevents, reduces, or eliminates biofouling
Epibiont	An organism that lives on the surface of another organism
Basibiont	An organism that is host to an epibiont.

82

83 2. Methods

84 *Literature search*

85 In accordance with the Preferred Reporting Items for Systematic reviews and Meta-Analyses
 86 (PRISMA) recommendations (Moher et al., 2009), we searched two major databases to collect
 87 relevant peer reviewed papers. We searched both Google Scholar and Web of Science using all
 88 possible combinations of the search terms “allelopathy”, “allelochemicals”, “antifouling”,
 89 “seaweed”, “macroalgae”, “Rhodophyta”, “Chlorophyta”, “Phaeophyta”, and “Phaeophyceae”

90 from 29 June to 06 July 2022. We used the search criteria “all topics” to search Web of Science,
91 returning in total 203 different articles (101 were deemed relevant and collected) and “in title”
92 was used to search Google Scholar, returning 540 articles (39 additional relevant papers were
93 collected, that were not yet found in the WoS database). We also scouted the literature that was
94 cited in the acquired articles for appropriate publications (27 additional articles collected). To
95 be included as relevant in our analyses, each article must have been an original, peer reviewed
96 research study, reporting used methodology as well as the data analysis. Studies were only taken
97 into consideration if the authors physically exposed a target organism to macroalgae in any
98 form and if that target organism experienced a significant negative growth/metabolic impact.
99 Studies that cannot fully claim allelopathy, due to incomplete investigations, or lack of control
100 over confounding factors, were included in the meta-analysis. In such cases, the responsible
101 compounds are referred to as either putative allelopathic or simply bioactive in this review.
102 Studies that reported chemicals produced by macroalgae as a predator defence were excluded,
103 as this is out of the scope of this review. We also excluded publications that examined the
104 effects of allelochemicals on non-marine target organisms (i.e., terrestrial or freshwater
105 species), as this is out of the scope of this review. In contrast, studies focusing on effects on
106 pelagic target taxa (i.e., phytoplankton) were not excluded as allelopathic benthic-pelagic
107 interactions due to competition for light and/or nutrients can take place, especially in coastal
108 areas. The applied search criteria yielded a total assembly of 167 articles, which we screened
109 for the following information: publication year, study location, the allelopathic and target taxa,
110 the used exposure method(s), used bioassay(s), the mitigation of potential confounding
111 variables, the identity of the allelochemicals, and the used methodology to isolate and identify
112 the chemical compound.

113 *Data analysis*

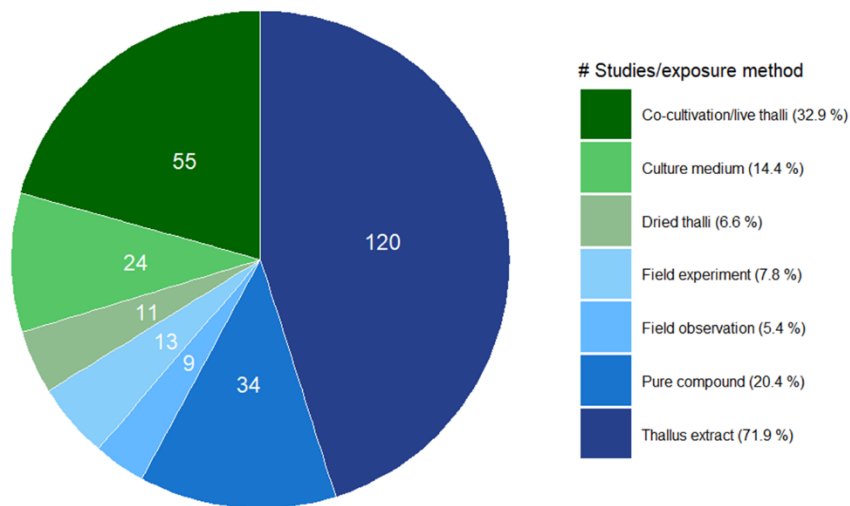
114 All retrieved studies were summarized and relevant data (described above) was collected in one
115 data table (Table S1). Figures were produced using the package ggplot2 (Wickham, 2009)
116 in R v4.1.1 (R Core Team, 2017).

117 3. Results

118 In total, 167 research articles were collected, of which majority of the studies (n =80, 47.9 %)
119 reported putative allelopathy present in at least one red seaweed species, followed by 74 brown
120 macroalgae and 61 green macroalgae studies (44.3 % and 36.5 %, respectively; Table S1, S2).
121 More statistics about country of origin, tested macroalgae and target taxa can be found in
122 supportive information (Fig. S1-S4). The results sections is outlined as follows: we first discuss
123 which methods have been used to investigate allelopathy in macroalgae. Subsequently, we
124 discuss the production of allelochemicals and how the environment can influence their
125 production or effectiveness. Finally, we focus on the identity of allelochemicals and discuss
126 their mode of action, prior to highlighting the different incentives to study this phenomenon.

127 3.1. Commonly used practices to study allelopathy

128 Several research methods are required to identify and confirm the presence of allelopathy in
129 macroalgae and their impact on competitors. We distinguish seven different methods for
130 observing macroalgal allelopathy, ranging from field experiments to exposure to thalli extracts
131 (Fig. 1).



132

133 Figure 1: Pie chart of absolute number of studies using a certain investigation method. Relative numbers (percentages) are
 134 added to the legend.

- 135 - Field observations relate to observing the co-occurrence (or absence) of the competitors in
 136 the natural environment. For coral-algae interactions, studies focus on contact zones
 137 between the coral and the seaweed thallus, and whether bleaching has occurred or not (e.g.,
 138 Rickert et al., 2015, 2016; Slattery and Lesser 2014; Vermeij et al., 2011). Field observation
 139 studies, on their own, are not sufficient to demonstrate the presence of allelopathy, as
 140 several confounding variables might be at play (cf. more profound discussion in section
 141 3.2).
- 142 - A second group of studies have used field experiments to demonstrate allelopathy (found
 143 in 7.9 % of the collected studies). For example, investigating coral-seaweed interactions,
 144 several studies have manipulated contact between coral and macroalgae in the field and
 145 investigated bleaching and mortality rates after exposing the corals to either live macroalgal
 146 tissue (e.g., Bonaldo and Hay, 2014) or their extracts, embedded in phytigel disks (e.g.,
 147 Rasher and Hay, 2010; Slattery and Lesser, 2014). In the recruitment plate method, the
 148 impact of seaweed on other organisms is studied by observing the settlement and survival
 149 rate of target taxa on specially placed grids or substrates, positioned in the field.

- 150 - Third, 55 publications (33.3 %) have used co-culturing experiments or microcosms to
151 identify allelopathy by focusing on effects on the target organism after exposure to live
152 thalli of the potential allelopathic species (e.g., in Ben Gharbia et al., 2017; Svensson et al.,
153 2013; Wang et al., 2016, 2017b). Similarly, co-cultivation on its own is usually not
154 sufficient to demonstrate allelopathy due to confounding variables that might be
155 responsible for the observed effect (see discussion in section 3.2).
- 156 - Some studies choose to work with dried thalli rather than live thalli, hereby avoiding some
157 of the confounding variables mentioned in section 3.2 (6.7 % of total studies; Table S1).
- 158 - The fifth method includes investigating the effect of the macroalgae culture medium
159 (filtrate) on the target organism (applied in 14.5 % of the collected studies). Here, the
160 macroalgae are cultured for a well-defined time period in the culture medium that,
161 subsequently, will be used for exposure, after correcting the nutrient concentrations and
162 filtering (for example with an autoclaved membrane filter) to avoid bacterial contamination
163 (e.g. Wang et al., 2006). The culture medium is added at the beginning of the experiment
164 (initial addition) or added intermittently (semi-continuous addition), when part of the
165 culture medium is removed daily and replaced with an equal volume of macroalgal medium
166 filtrate, re-enriched with nutrients (e.g. Wang et al., 2007). The latter method ensures the
167 effectivity of an allelochemical, as it controls for potential degradation of the compound,
168 as its half-life depends on its chemical structure.
- 169 - In the most-used method (exploited in 71.5 % of the studies analysed), the effect of
170 macroalgal extracts (of either fresh or dried thalli or thallus powder) on the growth,
171 survival, physiology and/or development of the competitors are analysed (e.g., Fu et al.,
172 2022; Pansch et al., 2009; Zhang et al., 2021; Table S1). Depending on the test organism,
173 these extracts can be tested either dissolved in the test medium, in phytagel disks, in

174 medium in a petri dish. While, ideally, thallus surface extracts should be tested, most
175 studies have used whole cell extracts to test for effects on the tested target organism.

176 - Finally, 20.6 % of the studied have tested pure isolated compounds (from extracts) for its
177 allelopathic properties (Table S1).

178 While most of the collected studies in this review have illustrated the inhibiting activity of a
179 compound through one or more of the methods described above, few studies have
180 demonstrated that the inhibitory activity is ecologically relevant, i.e., that the algal metabolites
181 are present at the algal surface, or released in the surrounding water at concentrations that are
182 deterrent to naturally occurring foulers (e.g., Dworjanyn et al., 2006; Schmitt et al., 1995). For
183 example, a majority of studies have been testing crude seaweed extracts rather than surface
184 extracts or have not considered confounding factors (as elaborated on in section 3.2). These
185 studies imply the presence of allelochemicals but overlook other factors, implying to be careful
186 in attributing allelopathic properties; they could be merely bioactive. In most cases, more
187 research is thus required to truly confirm the presence of allelopathy. Only a quarter of studies
188 (42) addressed the challenges mentioned above in their designs. Hence, we decided to not cover
189 the less relevant studies in terms of ecology in this review, they are only considered in the
190 statistical summaries in the supplementary information and in the discussion of the chemical
191 identity of the allelochemicals.

192 3.2. Confounding variables and other challenges studying allelopathy in macroalgae

193 Observing allelopathy among aquatic organisms in a natural setting provides the best evidence
194 of these negative interactions effectively occurring in nature. Yet, investigating allelopathy in
195 the field is often a challenge, as several environmental factors can mask allelopathic effects
196 (Legrand et al., 2003). Hence, several studies attempted to characterise allelopathic interactions
197 in co-cultivation experiments, in a controlled system (i.e., controlled light and illumination

198 period), where confounding variables can be actively controlled. If the identity of the
199 allelochemical is not known, reductions in growth, or reduced physiological status, could be
200 attributable to multiple factors. Below, we briefly discuss these confounding variables and we
201 provide several recommendations and suggestions to take these factors into account in future
202 studies.

203 First, in co-cultivation experiments, a decline in macroalgae/microalgae growth can be
204 anticipated, due to significant nutrient consumption and competition, as seaweed are known to
205 consume a substantial amount of nutrients (Bambaranda et al., 2019; Neori et al., 2004; Patil
206 et al., 2020). By actively monitoring the nutrient flux during the experiment, studies obtained
207 a better insight in the nutrient flow and whether nutrients became depleted during the
208 experiment or not (e.g., Accoroni et al., 2015; Nan et al., 2014; Table S1). Other studies (e.g.,
209 Nan et al., 2014; Patil et al., 2020) tackled this problem by periodically replacing part of the
210 culture medium with an equal amount of fresh medium.

211 Second, shading by (floating) macroalgal thalli may affect the growth of other
212 photoautotrophic species (White and Shurin, 2011). Several studies (e.g., Ma et al., 2017; Nan
213 et al., 2014; Patil et al., 2020; Table S1) mention that light limitation was unlikely in their
214 experiment as the macroalgae were always at the container's bottom, with the light source
215 above the flasks. Additionally, Tang et al. (2011) applied a PAR sensor to measure and confirm
216 that light intensity did not fluctuate significantly among the different treatments.

217 Third, studies that investigate the contact between the seaweed surface and the contact
218 organism (such as corals), should apply a plastic model or another algal mimic to control for
219 shading, abrasion, fractures, and the presence of a cable tie/other objects to connect the
220 organisms (e.g., Andras et al., 2012; Del Monaco et al., 2017; Rasher et al., 2011; Slattery and
221 Lesser, 2014; Table S1).

222 Furthermore, macroalgae can compete with co-occurring macro/microalgae through pH
223 alteration (Green-Gavrielidis et al., 2018; Sylvers and Gobler, 2021). Seaweed may increase
224 the pH of the test medium in which it is cultivated, by rapid photosynthesis and, thus, reducing
225 pCO₂, making it unsuitable for other organisms to grow (Hansen, 2002; Patil et al., 2020;
226 Sylvers and Gobler, 2021). For instance, *Ulva intestinalis* has been shown to increase the pH
227 of rock pools to above 10, a pH level where other macroalgae can no longer utilize the enzyme
228 carbonic anhydrase to convert HCO₃⁻ to CO₂ for use in photosynthesis, hence becoming carbon
229 limited (Björk et al., 2004). Hence, it is crucial that pH is regularly monitored in co-cultivation
230 studies (e.g., in Green-Gavrielidis et al., 2018; Jin et al., 2003; Nan et al., 2014; Sylvers and
231 Gobler, 2021; Tang et al., 2011; Table S1) to ensure the observed effects are not the result of
232 unsuitable pH values of the test media. Few studies actively maintained pH until termination
233 of the experiment by adjusting the pH accordingly (e.g., Accoroni et al., 2015; Benitt et al.,
234 2022). In their experiments, Patil et al. (2020) describe how they provided a continuous supply
235 of air to the culture medium via bubbling to counterbalance the observed pH increase.

236 Finally, several publications have also demonstrated that high levels of epiphytic and biofilm
237 forming bacteria can grow on the surface of the seaweed thalli and that some of these species
238 have growth-inhibiting and algicidal properties as well (Imai et al., 2002, 2006; Inaba et al.,
239 2017, 2020; Mayali and Azam, 2004). To prevent strong impact from these bacteria, several
240 studies suggested/took precautionary measures, such as liberal rinsing of the thalli with sterile
241 seawater or physically wiping of the macroalgae, yet the most suitable option is the inclusion
242 of an antibiotic (mixture) in a pre-treatment step (e.g., Benitt et al., 2002; Denboh et al., 1997;
243 Jeong et al., 2000; Jin et al., 2003; Tang et al., 2011; Table S1). Antibiotics that have been
244 applied in the investigated studies include penicillin, streptomycin, chloramphenicol,
245 ampicillin, kanamycin, polymixin, neomycin or a combination/ mixture of some of these. Tang

246 et al. (2011) performed periodic DAPI-staining of cultures to confirm the absence of bacteria
247 in their cultures.

248 If the results in co-cultivation systems cannot be entirely attributed to the effects of nutrient
249 depletion, pH elevations, light limitation, or algicidal bacteria, much of the inhibiting activity
250 observed is usually attributed to allelopathic metabolites secreted by macroalgae (e.g., in Patil
251 et al., 2020). Ideally, such findings should be confirmed by including other experiments and/or
252 the identification of the responsible compound(s). Finally, field studies should confirm whether
253 the observed interactions occur in a natural setting under the influence of natural processes,
254 such as advection, mixing of the water column, turbulence, etc., or if they only take place in
255 controlled conditions, i.e., in stagnant water in the lab.

256 3.3. Sites of production, transport and storage of allelochemicals in macroalgae

257 *Excretion or contact bound allelopathy?*

258 One important reason for our limited apprehension of allelopathy involving macroalgae is
259 related to the scarcity of knowledge on the localization of allelochemicals on/in seaweeds and
260 the potential processes that transport these compounds to the thallus surface. Bioactive
261 compounds may have various ecological functions (e.g., simultaneously functioning as an anti-
262 herbivore and an antifouling agent, as for example found in Schmitt et al., 1995), however in
263 some instances they exert a distinct functional role, and in those instances they may need to be
264 differentially distributed on/in macroalgae, according to their role. Herbivory deterrence can be
265 achieved by storing compounds within the seaweed, as herbivores encounter these metabolites
266 upon biting and ingesting entire thalli (Lane et al., 2009; Schmitt et al., 1995). However, in
267 order to be effective as defensive compounds, it would be more efficient if they are deployed
268 on the thallus surface, as this is the interface where interacting organisms, such as settling
269 larvae, come into contact (Andras et al., 2012; Nylund et al., 2007; Lane et al., 2009; Schmitt

270 et al., 1995; Steinberg and de Nys, 2002). Based on reviewed literature, two main types of
271 allelopathic mechanisms in macroalgae are suggested. First, seaweed can target competitors
272 and/or epiphytes by direct contact (i.e., through surface-associated compounds). The chemistry
273 of allelochemicals is most likely hydrophobic in nature, and thus non-polar, with the purpose
274 of maximizing its concentrations at/near host surfaces and avoiding swift dissolution (A Abdul
275 Malik et al., 2020; Steinberg and de Nys, 2002). The dominance of fatty acids/lipids as
276 allelochemicals (see below) makes sense in an evolutionary and energetic context given the
277 potential of the marine environment to dilute water-soluble compounds (Rasher and Hay,
278 2010). Secondly, macroalgae could produce a sufficient amount of polar bioactive metabolites,
279 which is subsequently excreted into the water and reaches target taxa with an effective, active
280 concentration (A Abdul Malik et al., 2020; Gross, 2003). Compared to terrestrial plants,
281 seaweed have thalli without stomata, with less tight cell connections, and with a reduced (or
282 even absent) cuticle (Hutchinson, 1975), likely facilitating a direct release of hydrophilic,
283 water-soluble metabolites into seawater. However, both dilution and short half-life of these type
284 of allelopathic agents are major challenges, hence requiring greater concentrations of
285 macroalgal allelochemicals (Fong et al., 2019). Solely based on polar properties, it is
286 challenging to ascertain whether a single metabolite could exert an allelopathic function at the
287 macroalgal interface (A Abdul Malik et al., 2020). Physicochemical interactions between the
288 allelochemical and the seaweed surface (e.g., through ionic interactions or hydrogen bonds)
289 could promote or restrict its diffusion into the environment, as suggested by A Abdul Malik et
290 al. (2020). The storage location, the site(s) of production, as well as the disposition among
291 external and/or internal tissues of most allelochemicals has not been established yet, but a
292 couple of bioactive compounds have been determined to specifically be present on the thallus
293 surface of seaweed (e.g., Andras et al., 2012; Dworjanyn et al., 1999; Lane et al., 2009; Schmitt
294 et al., 1995). For instance, Schmitt et al. (1995) demonstrated that larvae of the bryozoan *Bugula*

295 *neritina* were not affected by water from near the brown macroalga *Dictyota menstrualis* and
296 did not avoid contacting *Dictyota*, relative to another substrate. However, the larvae would not
297 settle on *Dictyota* following contact. These findings suggested the presence of a deterrent
298 hydrophobic/lipophilic metabolite located on the thallus surface of *Dictyota*, i.e., the
299 compounds pachydictyol A and dictyol E, that were not released into the water. Furanones from
300 the rhodophyte *Delisea pulchra* were also quantified at the algal surface and studied as
301 antifoulants towards a range of target organisms (Dworjanyn et al., 1999, 2006). Finally, Rasher
302 and Hay (2010) found lipophilic extracts from a number of seaweed species, including the
303 brown macroalga *Lobophora variegata*, caused significant coral bleaching, while this was not
304 the case for the hydrophilic metabolites extracted from the, respectively, green and red
305 macroalga *Chlorodesmis fastigata* and *Galaxaura filamentosa*. Up to now, all demonstrated
306 allelopathic interactions between seaweed and corals depend on the transfer of compounds via
307 direct contact (via tissue bound allelochemicals), rather than via transmission across seawater
308 (de Nys et al., 1991; Rasher and Hay, 2010). This suggests that contact generated effects, rather
309 than proximity alone, and that their allelopathic agents are lipid soluble rather than hydrophilic
310 in nature.

311 Together, these findings seem to confirm the importance of direct contact, which would be
312 advantageous for lipophilic allelopathic compounds. Yet, in several studies (e.g., Jin et al.,
313 2005; Nan et al., 2008; Nelson et al., 2003; Patil et al., 2020; Wang et al., 2007, 2012; Xu et al.,
314 2012), isolated live thalli (co-cultivation), growth medium filtrates, dried thalli, and aqueous
315 extracts of thalli all dwindled the growth of harmful algal blooms, suggesting that direct contact
316 is not always required to cause the observed inhibitory effects. The possibility of growth
317 inhibition by direct contact with the seaweed specimen itself can be ruled out by carrying out
318 isolation co-culture assays (e.g., Jin et al., 2003).

319 *Storage and transport processes*

320 Whether secreted or surface bound, currently little is known as well about the
321 mechanisms/processes related to storage and transport to the seaweed's thallus surface. In
322 seaweed, bioactive metabolites are potentially stored in internal structures inside the thallus and
323 not distributed to its surface, are potentially made in inner glands but secreted into the exterior
324 via a connection pore, or can be allocated to the thallus surface via an unknown route (Andras
325 et al., 2012; de Nys et al., 1998; Dworjanyn et al., 1999; Lane et al., 2009; Paul et al., 2006).
326 Bioactive compounds that come to the surface and that have been observed to act as
327 allelochemicals, have been found to be stored within several types of specialised structures
328 associated with the thallus surface: secretory trichomes; epidermal gland cells (or vesicle cells),
329 in which a storage vesicle preoccupies almost the entire cellular space (Dworjanyn et al., 1999;
330 Steinberg and de Nys, 2002), physodes, i.e. membrane-bound vesicles from brown macroalgae
331 (Schoenwaelder, 2002), and *corps en cerise* or "cherry bodies" (CC), which are spherical,
332 reniform to claviform, refractile intracellular inclusions mostly occurring in the cortex of red
333 algae species of the genus *Laurencia* (Feldmann and Feldmann 1950; Salgado et al., 2008;
334 Young et al., 1980). For instance, *Laurencia dendroidea* was shown to store halogenated
335 metabolites in the *corps en cerise* and to transport these through vesicular traffic to the
336 seaweed's surface in response to epiphytic bacteria (Salgado et al., 2008, Paradas et al., 2010).
337 The production and transfer of most bioactive metabolites remain unclear however. For
338 example, Paul et al. (2006) reported about the presence of stalk-like structures in *Asparagopsis*
339 *armata* and argued that these could function as a possible mechanism to transport
340 allelochemicals to the surface of the thalli. The red alga *Bonnemaisonia asparagoides* has
341 multitudinous specialized gland cells on its thallus's exterior, in which it likely stores its
342 antibacterial metabolites to release them to the thallus surface, as suggested by Nylund et al.
343 (2010).

344 Reis et al. (2013) were able to demonstrate that both microfilaments and microtubules play a
345 pivotal part in the vesicle transport from the *corps en cerise* to the periphery of the cells for
346 subsequent exocytosis of the secondary metabolites in *Laurencia dendroidea*, as well as the
347 anchoring and positioning of the CC's themselves to the cell's periphery. Via time-lapse video
348 microscopy, exocytosis of these vesicles in the cortical cells of *L. obtusa* through the plasma
349 membrane to the inner cell wall was observed by Salgado et al. (2008). They argued that
350 exocytosis transfers the bioactive metabolites to the seaweed's exterior and that fluctuations in
351 this process might result in varying concentrations of the metabolites on the seaweed's exterior.
352 Next to allowing the regulation of metabolite exudation, another explication for the energetic
353 investment in storage room (e.g., the *corps in cerise* in *Laurencia*, the physodes of brown
354 seaweeds, the gland cells in *Delisea pulchra*) and the transport mechanisms into the apoplast,
355 might be the autotoxicity of allelochemicals, as compartmentalization and transport can abate
356 the concentration of the metabolites in the cytosol, lessening cytotoxicity (Shitan, 2016).

357 *The necessity to differentiate between whole tissue extracts and thallus surface extracts*

358 Most of the investigated studies used crude-extracts of entire macroalgae to screen directly for
359 allelopathic activity (Table S1). The use of crude extracts of fresh thalli or dried powder
360 theoretically might have an advantage over the direct application of fresh tissue, as
361 concentrations can be controlled for as well as eliminating the discussed confounding variables
362 (section 3.2). In contrast, few studies have quantified the naturally occurring concentrations of
363 allelochemicals on seaweed surfaces or have investigated the variation between surface-
364 associated metabolites and those held within the seaweed's tissue. Yet, examining the
365 localization, storage as well as the exudation mechanisms of allelochemicals is required to
366 enhance our apprehension of the ecological significance of these metabolites (Nylund et al.,
367 2007), especially from the exposure point of view. Although the crude extract procedure
368 provides an insight in the broad-spectrum activity of bioactive compounds, the natural transport

369 mechanisms, as well as the responsiveness of ecologically relevant species to the secreted
370 bioactive metabolites remain mostly undisclosed (Dworjanyn et al., 2006; Nylund et al., 2007).
371 The whole thallus extraction approach risks to overestimate the defence capability of the studied
372 macroalga, and the absence or presence of spatial allocation could significantly change the
373 interaction outcome between competitors (Andras et al., 2012; Nylund et al., 2007; Steinberg
374 and de Nys, 2002). If metabolites occurring within macroalgae are extracted and tested as if
375 they were present on the macroalgae, then their true ecological effects are likely misrepresented
376 (Longo and Hay, 2017). Few studies have explicitly tested this, and concluded that the
377 inhibiting effects of whole thallus extracts were inadequate predictors of the inhibiting abilities
378 of the tested macroalgae. For instance, elatol concentrations, obtained from total (lipophilic)
379 tissue extracts of the rhodophyte *Laurencia obtusa* did suppress herbivorous and fouling
380 activity significantly, while surface concentrations did not (da Gama et al., 2002; Pereira et al.,
381 2003; Sudatti et al., 2008). Nylund and Pavia (2003) investigated the effects of lipophilic crude
382 extracts obtained from four red macroalgae species on the settlement behaviour of cyprid larvae
383 of the co-existing barnacle *Balanus improvisus* using different approaches. First, they found
384 that larval settlement was strongly inhibited at presumably ecologically relevant surface
385 concentrations, based on surface area/dry weight ratio estimations, for all four different seaweed
386 species. These findings were then compared with a settlement preference experiment, in which
387 was tested whether the cyprid larvae rather settled on algal surfaces or on control surfaces. The
388 thallus surface of only one of the four tested seaweed species, *Chondrus crispus*, significantly
389 hampered settlement in the preference experiment, while in one other species there seemed to
390 be a settlement stimulation. Nylund and Pavia (2003) argued that these contradicting results
391 likely are the consequence from the extraction of internal metabolites, that usually do not occur
392 on the thallus surface. Longo and Hay (2017) performed several field experiments to assess the
393 impacts of (1) lipid-soluble extracts from seaweed surfaces of multiple species, next to (2) total

394 lipid-soluble extracts from both internal and external seaweed tissues on the coral *Pocillopora*
395 *verrucosa* and its symbionts. Extracts of the brown seaweed *Dictyota bartayresiana*, the green
396 macroalga *Chlorodesmis fastigiata* and the red macroalgae *Amansia rhodantha* and
397 *Asparagopsis taxiformis*, were found to suppress coral photochemical efficiency (Longo and
398 Hay, 2017). However, they also found that the total lipid-soluble extracts of these species were
399 not more toxic compared to the surface-only extracts, even though the concentrations of the
400 total lipid-soluble extracts exceeded the surface-only extracts notably. Together, these findings
401 suggest that bioassays with entire tissue extracts can be biologically meaningful but are also
402 less ecologically relevant compared to the surface extracts. According to Paul et al. (2006), the
403 delivery of a compound to the macroalgal surface and the inhibiting efficiency of the metabolite
404 on the thallus designate important criteria that differentiate a natural antifoulant from potential
405 bioactive metabolites. Hence, we recommend that future bioassays are preferably performed
406 with the more simple, less concentrated seaweed surface extracts, and thus more relevant in
407 terms of ecology. Several studies have already demonstrated allelopathy based on testing with
408 relevant surface concentrations of the putative allelopathic agent. For example, surface-
409 extracted compounds derived from the brown macroalgae *Fucus vesiculosus* and *F. serratus*
410 were shown to deter the settlement of several micro- and macrofoulers (Rickert et al., 2015,
411 2016), whereas surface extracts generated from *Gracilaria vermiculophylla* were found to deter
412 epiphytic, filamentous red macroalgae (*Ceramium* spp.) as well as benthic diatoms (Wang et
413 al., 2017b).

414 There is to date no standardised method for testing ecologically relevant concentrations of
415 surface compounds against fouling activity or any other type of competition. Most studies have
416 employed the “swab” or “hexane dip” method to investigate undamaged biotic surfaces (de Nys
417 et al., 1998; Nylund et al., 2005, 2007, 2010; Schmitt et al., 1995). The “hexane dipping”
418 method, developed by de Nys et al. (1998), allows a quantitative extraction of hydrophobic

419 metabolites from the surface of the thalli without (visually) disrupting cells by dipping the algae
420 in hexane for 5 to 40 s at room temperature. Several studies (e.g., A Abdul Malik et al., 2020;
421 de Nys et al., 1998; Nylund et al., 2007), have used epifluorescence microscopy to clearly
422 distinguish intact cells from lysed cortical cells of the macroalgae thalli after exposing (dipping)
423 them to different solvents, such as dichloromethane, methanol, n-hexane, and
424 dichloromethane/methanol (1:1), without any staining. In the study by Othmani et al. (2016),
425 several dyes were tested to confirm the viability of cells from the brown alga *Taonia atomaria*
426 after applying the ‘dipping method’ described above. More rigorous extraction methods using
427 either other solvents, or longer extraction times in hexane (> 50 s), were found to cause
428 significant visible cell damage (A Abdul Malik et al., 2020; de Nys et al., 1998; Nylund et al.,
429 2007). However, while these procedures confirm whether the cell membrane remains intact
430 after extraction or swabbing, they do not confirm whether or not a transfer of intracellular
431 metabolites occurred without a visible cell disruption (Andras et al., 2012). Moreover, the
432 dipping method only allows incomplete extractions of surface compounds, hence, likely
433 underestimating the true allelochemical concentration (Wang et al., 2017a). These techniques
434 have thus a limited ability to quantify metabolite distribution or concentration and are mostly
435 valuable for confirming whether the bioactive compounds occur on the thallus surface.

436 Advances in imaging mass spectrometry have made it possible to analyse the chemistry of
437 natural surfaces in their original state. Some studies have employed desorption electrospray
438 ionization mass spectrometry (DESI-MS) to localise and measure bioactive metabolites on the
439 thallus surfaces (Andras et al., 2012; Lane et al., 2009; Nyadong et al., 2009), illustrating the
440 potential of this technique to determine the existence and concentrations of allelopathic
441 metabolites on thallus surfaces. For example, Andras et al. (2012) used DESI-MS to visualise
442 and quantify the metabolite neurymenolide A on the surface of the red alga *Phacelocarpus*
443 *neurymenioides*, which was found to induce coral bleaching on natural colonies of *Porites rus*

444 in a field experiment. In conclusion, improving our comprehension of where compounds occur,
445 and in which concentrations and conditions, enlightens us about the metabolite's ecological
446 function and is critical for the design of ecological representational experiments (Andras et al.,
447 2012; de Nys et al., 1998; Lane et al., 2009).

448 3.4 Biotic and abiotic factors affecting allelopathic behaviour and effectivity

449 Abiotic conditions such as temperature, salinity, irradiance, UV radiation, pH, and nutrient
450 availability vary continuously in aquatic ecosystems and these fluctuations have a considerable
451 impact on the seaweed's growth, development and physiology (Gross, 2003; Jin et al., 2013).
452 The environment might influence allelopathic effectivity in at least three different ways: (1) the
453 production of the chemicals, (2) their bioavailability and (3) how they affect the target
454 competitor, which itself might be more sensitive as well to the allelopathic substance under
455 different circumstances (Gross, 2003).

456 Under conditions of environmental stress, a lower investment of resources and/or energy into
457 defence and increased investments into maintenance is predicted by the environmental stress
458 theory (Cronin, 2001; Pansch et al., 2009). The theory presumes that defences are costly, and
459 thus are actively reduced under resource limitation and environmental stress, yet these
460 assumptions have rarely been tested in experimental studies (Dworjany et al., 2006).
461 Noteworthy, Rasher and Hay (2014) found that competition with corals induced increased
462 allelochemical content in the seaweed *Galaxaura filamentosa*. Yet, they also observed the
463 seaweed experienced significantly reduced growth and increased palatability to herbivores
464 (because of reduced chemical defences). As they did not observe these patterns in non-
465 allelopathic species, it illustrates a complex coral-seaweed-herbivore interaction, with different
466 defense strategies against herbivores and competing corals. In terrestrial plants, there is strong
467 evidence that exposure to environmental stress induces the production of allelochemicals

468 (Muzell Trezzi et al., 2016). As chemical defense production can be adapted (regulated)
469 according to the seaweed's need, it could be assumed that the production would fluctuate
470 temporarily in temperate latitudes as both environmental factors, as well as biotic factors, such
471 as fouling pressure, undergo seasonal shifts (Saha and Wahl, 2013). Hence, macroalgal defence
472 intensity should thus either fluctuate (1) with abiotic factors controlling the energy resources,
473 (2) with defence demand, such as fouling pressure, or (3) with both (Rickert et al., 2015).
474 Despite common recognition of these impacts, data on allelopathic dynamics in marine
475 ecosystems are restricted (Table S1; Gross, 2003). Producing defensive metabolites can
476 fluctuate in several ways: seasonal or temporal, spatial and/or geographical, within populations
477 (density dependent), or even within different parts of the thallus. Hence, several studies, mainly
478 on temperate macroalgae, have recorded seasonal variability in antifouling activity (e.g., Hellio
479 et al., 2004; Rickert et al., 2015; Saha and Wahl, 2013). The robustness of spatio-temporal
480 patterns observed in the defence strength of several species, suggests that variation in
481 allelopathy strength is not random and likely regulated. If so, the mechanistic foundation for
482 such regulation, as well as its drivers remain to be identified and requires further research. In
483 this section, we provide and discuss, non-exhaustively, a few studies in which biotic and abiotic
484 factors have been suggested to affect either allelochemical production as well as sensitivity
485 towards the allelochemical.

486 *The influence of genetic factors*

487 Variation in bioactive metabolite content in different seaweed populations has been
488 documented in a number of studies (e.g., Koivikko et al., 2008; Pereira et al., 2004; Vallim et
489 al., 2005). These fluctuations might be interpreted as resulting from genetic differentiation
490 between populations and/or are the response to local changes in the environment, as described
491 above, possibly affecting allelochemical activity. For example, Plouguerné et al. (2010)
492 reported spatial variation of the antifouling activity of *Sargassum vulgare*, collected at different

493 sites along the coast of Rio de Janeiro, Brazil. Similarly, Saha and Wahl (2013) observed anti-
494 settlement activity of eight biofilm-forming bacteria after exposure to surface extracts of *F.*
495 *vesiculosus* and found a consistent and compelling difference between locations throughout the
496 year. In contrast, Jeong et al. (2000) found no regional and seasonal variation (collected
497 bimonthly from five different locations around Korea) in the algicidal activity of *Corallina*
498 *pilulifera*. This research area will likely progress with increased efforts in understanding the
499 genetic structure of macroalgae populations as well as the characterisation of biosynthetic
500 pathways, enzymes and genes in control of the production of the allelopathic agents.

501 *The influence of light intensity and irradiance*

502 The distribution pattern of light is rather complex in aquatic systems, with both low and high
503 light intensities limiting photosynthetic efficiency, either by insufficient energy input or by
504 photo-inhibition (Cade-Menun & Paytan, 2010). Light can impact allelopathic effectiveness at
505 different levels. First, light intensity is presumed to impact photosynthetic rate and, thus,
506 seaweed physiology, e.g., variation in protein and lipid content (e.g., Cade-Menun & Paytan,
507 2010; Jin et al., 2016). For example, assuming that fatty acids (mainly PUFAs) are the bioactive
508 compounds of several seaweed species (e.g., Alamsjah et al., 2005, 2008; Jin et al., 2016; Tang
509 and Gobler, 2011), an increased concentration of these fatty acids under lower/higher light
510 intensities may increase the biocidal effect. Second, photochemical reactions of the allelopathic
511 agents under high levels of irradiance might improve or decrease their effectivity (e.g., through
512 photodegradation for example; Jin et al., 2016). Finally, also the sensitivity of the target
513 organism might be altered after exposure to different levels of irradiance and allelochemical
514 concentrations. However, study-based evidence for these hypotheses is rare. For instance,
515 Pansch et al., (2009) did not find an effect of light reduction on the chemically mediated
516 defences of *Dictyota kunthii* on the bioactivity against mussels.

517 *The influence of temperature, pH and ocean acidification*

518 Temperature and pH are key factors affecting the growth and physiological activities of all
519 aquatic organisms. Similarly as to light intensity, temperature and pH changes might alter
520 allelopathic efficiency and/or sensitivity. Valenti et al. (2010) proposed that pH can change the
521 ionisation of metabolites, thereby influencing the lipophilicity, bioavailability, bioaccumulation
522 and aquatic toxicity of allelopathic compounds. For example, Del Monaco et al. (2017)
523 demonstrated that lipophilic, surface extracts of the brown seaweed *Dictyota cervicornis* were
524 more harming to the coral *Acropora intermedia* when competing under CO₂ levels predicted to
525 occur in 2050 and 2100, compared to current-day, ambient conditions. The extracts from two
526 other tested macroalgae species *Chlorodesmis fastigiata* and *Amansia glomerata* were not more
527 potent to the coral when grown under increased CO₂ conditions (Del Monaco et al., 2017).
528 Several studies reported that the growth inhibitory efficiency of the examined seaweed species
529 was modulated by temperature (e.g. Denboh et al., 1997; Jin et al., 2016; Wang and Tang, 2016;
530 Table S1). For instance, Denboh et al. (1997) reported a temperature dependent growth
531 inhibition of *Laminaria japonica* after exposure to rhodophytes *Corallina pilulifera* and
532 *Pneophyllum zostericolum*, especially at higher temperatures. In a series of multivariate
533 experiments, Jin et al. (2016) found that the growth inhibitory impact of *Ulva pertusa* on co-
534 cultivated *H. akashiwo* was most potent at the second highest temperature tested (25°C) with,
535 high irradiance (400 μmol m⁻² s⁻¹) and high alkalinity (pH = 10).

536 *Nutrient concentrations*

537 Many macroalgae are often restricted in their capacity to concentrate and store nitrogen
538 internally, depending on a continuous and prominent supply of nitrogen (Xu et al., 2012). *Ulva*
539 spp., for example have a high surface area/volume ratio and exhibit high rates of nutrient uptake
540 (Xu et al., 2012; Zhang et al., 2013). However, several species are capable of storing nutrients;
541 e.g., the genus *Gracilaria* requires only pulse fertilisation, as it is capable of storing nitrogen,
542 partly as the pigment phycoerythrin (Friedlander et al., 1996). Benitt et al. (2022) investigated

543 the co-effect of nutrient limitation and allelopathy in the inhibition of the blooming pelagophyte
544 microalga *Aureococcus anophagefferens* (Ochrophyta) by *Gracilaria* and *Dasysiphonia*. They
545 found that allelopathy effectivity depended upon nutrient conditions and that *A.*
546 *anophagefferens* was apparently more resistant to seaweed cultured in low nutrient
547 concentrations.

548 *Population densities*

549 More biomass in both natural and culture systems steers towards (more) nutrient limitation and,
550 hence, a fiercer competition (Gao et al., 2014). Multiple studies have indeed examined whether
551 higher macroalgal population densities/biomass impact allelopathic effectivity under fixed
552 light, nutrient and pH conditions (e.g. Lv et al., 2021; Patil et al., 2020; Tang et al., 2015; Wang
553 et al., 2013; Xu et al., 2012; Table S1). Benitt et al. (2022) described how red algae species
554 significantly reduced densities of the microalga *A. anophagefferens* in a dose-dependent way,
555 with the effectivity of each species depending mainly on both *A. anophagefferens* abundances
556 as well as the seaweed densities themselves. From the target point of view, Patil et al. (2020)
557 described how the inhibitory effects of the thalli of the red seaweed *Pyropia haitanensis* were
558 substantially reduced at higher initial microalgal cell densities of the bloom forming
559 *Skeletonema costatum* (Bacillariophyceae). Similarly, an attenuation in the inhibition of the
560 thalli of *Porphyra purpurea* on the growth of *A. anophagefferens* and *Karlodinium veneficum*
561 (Dinoflagellata) was noted after an elevation in initial microalgal cell abundance (Tang et al.,
562 2015).

563 *Tissue age and specialisation*

564 The age of the studied tissue can also have an impact on the concentration of the studied
565 allelochemical. For example, Andras et al., (2012) found that average concentrations of the
566 allelochemical neurymenolide A were more than double on older, bottom portions of the
567 sampled thalli compared to those of younger tissues in the blades of the rhodophyte

568 *Phacelocarpus neurymenioides*. This could hint that this macrolide is actively being
569 sequestered in elder parts to defend the basal portions of the thallus that anchor the macroalga.
570 Alternatively, it could suggest that the allelochemical accumulates over time in tissues, or that
571 it is selectively transported to older parts of the thallus, as these parts have encountered long-
572 term exposure to fouling organisms already (Andras et al., 2012). Additionally, Plouguerné et
573 al. (2012) observed tissue specialisation in *Sargassum vulgare* in regard to its antifouling
574 activity against the mussel *Perna perna*. They observed that polyphenol extracts from its
575 pneumatocysts (air vesicles, providing buoyancy) had the highest antifouling activity, followed
576 by the leaflets, while the reproducing structures, i.e. the receptacles, exhibited the least
577 antifouling activity.

578 *Predation*

579 Several studies have examined the impact of predation on the production of allelopathic
580 compounds or vice versa. Ensuing eight days of competing with the coral *Porites cylindrica*,
581 the macroalga *Galaxaura filamentosa* elevated allelochemical production and was almost as
582 double harmful to the coral (Rasher and Hay, 2014). However, owing to competition,
583 *Galaxaura* itself experienced a reduced growth and a softening of its anti-herbivore chemical
584 defence, followed by an increased palatability. Rasher and Hay (2014) concluded that
585 *Galaxaura* secreted different chemical compounds to oppose/resist competitors versus
586 herbivores. In some species, bioactive compounds are broadly defensive against a wide range
587 of both consumers and competitors, circumventing the energy costs of herbivore deterrence as
588 the same trait both provides herbivore deterrence/resistance, as well as offers competitive
589 advantages (Schmitt et al., 1995). For instance, Schmitt et al. (1995) were able to demonstrate
590 that diterpene alcohols, secreted by brown macroalga *Dictyota menstrualis*, are not only used
591 to defend itself from predation by sea urchins and herbivorous fishes, but also serve as
592 allelochemicals damaging the larvae of fouling organisms.

593 3.5. The identity of (putative) allelochemicals in macroalgae

594 Identification of the responsible allelochemical is a costly and time-consuming undertaking.
595 Yet, 32.9 % (n = 55) of the investigated and collected studies have described at least one
596 chemical, produced by either brown, green and red macroalgae, that significantly affects the
597 biological activity of one or more target organisms (Table S3). In total, bioactive compounds
598 have been identified in 57 different species so far, of which almost half of them brown
599 macroalgae (n = 26, 45.6 %). In comparison to Rhodophyta (n =20; 35.1 %) and Ochrophyta,
600 fewer compounds in Chlorophyta were described to date (n =11 species; 19.3 %), of which
601 more than half belonging to the genus *Ulva* (Table S3). Remarkably, only twelve of these 55
602 studies (21.8 %) have truly identified an allelochemical with inhibiting properties via realistic
603 exposure conditions (i.e., through ecologically relevant assays, as discussed profoundly below;
604 Table S3). Hence, much effort still lies in the characterization of allelopathic compounds in
605 macroalgae, as well as in distinguishing true allelochemicals from bioactive molecules.

606 According to their chemical structure, four major categories of algal synthesized bioactive
607 compounds can be distinguished: (1) carboxylic acids/fatty acids and derivatives; (2)
608 (poly)phenolic compounds; (3) terpenoid compounds; and one of its subgroups (4) sterols
609 (Table 1). Fatty acids and derivatives were the most retrieved group of putative allelochemicals
610 in our survey (59.6 %) in all three evolutionary lineages, including the regularly found
611 hexadecenoic acid, octadecanoic acid and eicosatetradecanoic acid (Table 1). The occurrence
612 of simple phenols, such as flavonoids, benzoic acids and derivatives has been observed in all
613 seaweed lineages, but brown macroalgae have been described to possess higher phenolic
614 contents relative to green and red macroalgae (Aina et al., 2022; Cotas et al., 2020). In the
615 searched literature, only terpenes/terpenoids with a growth inhibiting activity in brown and red
616 macroalgae have been described so far (Table S3), even though terpenoids are occurring in all
617 living organisms (because of their role in the respiration chain electron transport).

618 While it is important to investigate the structure and identity of \$ allelochemicals, we should
619 recall that allelopathic function might not depend exclusively on one key metabolite. Mixtures
620 of compounds could act synergistically or additively, even when the pure compounds
621 individually might not cause a visible effect (Blum, 1999).

622 3.6. The allelopathic mode of action

623 Although the inhibitory consequences of macroalgae extracts have been studied on a plethora
624 of species, not much is known about their mechanisms of inhibition in the target taxon.
625 Allelochemicals likely cause disturbances in range of physiological and developmental
626 processes in target organisms (Ben Gharbia et al., 2017). To understand their mode of action as
627 well as to establish the extent of harmful impacts on the neighbouring/target species, several
628 investigations must be conducted to elucidate the physiological mechanisms of allelopathy.

629 The inhibition of photosynthetic efficiency or functioning of competing primary producers is
630 considered a very common mode of action among many photosynthetic organisms.
631 Photosystem II (PSII), a membrane protein supercomplex executing the first reaction of the
632 photosynthetic pathway, is most often identified as being the main target (Ben Gharbia et al.,
633 2017; Gross, 2003). To examine photosynthetic energy conversion, the maximum quantum
634 yield of PSII, i.e., a proxy for the efficiency of excitation energy capture, can be used to confirm
635 the integrity and functioning of PSII and has been broadly used to analyse effects of
636 stressors/stress on the target organism's photosynthetic activity (e.g., Baker and Rosenqvist,
637 2004; Gross, 2003; Table S1). For example, exposure to thalli of *Ulva rigida* caused a
638 significant decrease in the maximum PSII quantum yield in several dinoflagellate species in a
639 dose-response manner (Ben Gharbia et al., 2017). Ye et al. (2014) found that dried thalli of
640 *Gracilaria lemaneiformis* mainly inhibited photosynthetic targets in *Scrippsiella trochoidea*
641 (Dinoflagellata) with a reduction in the quantities and sizes of antenna chlorophyll, a decrease

642 in photochemical efficacy of PSII, obstruction of the electron transport chain, in addition to
643 damage to the oxygen-evolving complex. None of the studies were able to describe the initiating
644 event in the inhibition of the photosynthetic pathway. Nevertheless, considering the crucial
645 importance of photosynthesis photoautotrophs, a reduction in efficiency of the photosynthetic
646 apparatus, induced by an allelochemical, is a strong indicator of its phytotoxicity (Raniello et
647 al., 2007; Ralph, 2000).

648 Some authors have argued that (microalgal) cells could be triggered to produce reactive oxygen
649 species (ROS) and trigger oxidative stress by stimulation of allelochemicals (El-Darier et al.,
650 2021; Qian et al., 2009; Sun et al., 2012, 2021b; Xu et al., 2020; Zhang et al., 2021).
651 Allelochemicals might potentially cause an increase in oxygen free radicals (Gill and Tuteja,
652 2010). These radicals are usually involved in membrane lipid peroxidation, degradation of
653 chloroplasts and in the reaction with several macromolecules, subsequently disturbing cell
654 growth and proliferation (Gill and Tuteja, 2010). To evaluate the effect of α -linolenic acid,
655 identified in crude *Sargassum fusiforme* extracts, on antioxidant enzymes, Sun et al. (2021b)
656 measured superoxide dismutase (SOD) and peroxidase (POD) activity, two common
657 antioxidant enzymes of HAB species *Heterosigma akashiwo*. Their analyses showed that SOD
658 and POD activity gradually decreased under high α -linolenic acid concentrations. Similarly, the
659 activities of four oxidoreductases, including SOD, showed an initial increasing trend of activity
660 and then decreasing in the diatom *Skeletonema costatum* after cultivation with *S. fusiforme*
661 extracts as well (Zhang et al., 2021). According to the authors, these results can be interpreted
662 that the stress caused by low extract concentrations initially increased ROS levels in diatoms,
663 as well as that antioxidant enzymes were not capable of eradicating the excessive amount of
664 oxidation products, which in turn caused damage to the enzyme system. In other words,
665 excessive amounts of free radicals cannot be eradicated by the antioxidant system, which
666 potentially affects the activity of its enzymes in turn (Gill and Tuteja, 2010; Zhang et al., 2021).

667 Another allelopathic mechanism in macroalgae that has been suggested is associated with the
668 disruption of the osmotic pressure regulation, mainly by studies that found changes in the shape
669 and/or motility of exposed (micro)algal cells. For example, phlorotannins from phaeophyte
670 *Ecklonia kurome* made the cells of three red tide HABs lose their motility and almost all
671 microalgae became round in shape before expanding and bursting (Nagayama et al., 2003).
672 Nagayama et al. (2003) argued that interactions between phlorotannins and microalgal channel
673 proteins in their membranes played a pertinent role in the algicidal actions of phlorotannins,
674 consequently disturbing the control of osmotic pressure. Wang et al. (2013) speculated as well
675 that the allelopathic mode of action through which *Ulva pertusa* or *U. prolifera* inhibited the
676 growth of the dinoflagellate *Prorocentrum donghaiense* might be associated to a disrupted
677 cellular control of its osmotic pressure, because motility and modality of the microalgal cells
678 were limited by the allelopathic compounds as well. Wu et al. (2006) described that fatty acids
679 caused rapid permeability changes in plasma membranes, which are related to K⁺ ion leakage,
680 and the resulting degradation of the organisation of the membrane. In the study of Oh et al.
681 (2009), cells of the affected organism first displayed swelling and then lysed during exposure
682 to the fatty acid EPA. Despite these finding based hypotheses, the mode of action of most
683 allelochemicals remains unclear. To enhance our understanding of underlying mechanisms of
684 action, further research is required.

685 3.7. Incentives and relevance of studying macroalgal allelopathy

686 The interest in allelopathy among aquatic organisms is growing for several reasons. Each of the
687 main incentives will be discussed briefly below.

688 *The role of allelopathy in shaping and structuring marine communities*

689 The character and intensity of competitive interactions between organisms influence the
690 functioning and structure of any community (Maggi et al., 2012; Nabivailo et al., 2014; Paine,

691 1990). Allelopathy among primary producers has been reported to have a considerable impact
692 on the structure and succession of an aquatic ecosystem (e.g., Getachew et al., 2017; Gross,
693 2003; Jin et al., 2016; Kakisawa et al., 1988; Kim et al., 2004; Green-Gavrielidis et al., 2018;
694 Slattery and Lesser, 2014). Macroalgal communities are usually dominated by species with high
695 competitive abilities under certain biotic and abiotic settings (Carpenter, 1990). This ability can
696 either be (1) rapidly adapting to the limitations in resources and exploiting them effectively,
697 making the resources inaccessible for their competitors (i.e., exploitative competition), or (2)
698 influencing the competitor's physiology through allelopathy (i.e., interference competition)
699 (Maggi et al., 2012; Paine, 1990). For example, coralline red algae were found to produce
700 metabolites causing maturity suppression in *Laminaria* (Denboh et al., 1997).

701 Additionally, allelopathic activity has currently been acknowledged as a mechanism in the
702 invasion of a species into a community (Inderjit et al., 2011). Callaway & Aschehoug (2000)
703 formulated the 'novel weapons hypothesis' (NWH). Their proposition projects that exotic
704 invaders will become successful if they bring unique bioactive compounds to their range
705 expansions to which the native, endemic species are not adapted. As the endemic taxa are,
706 consequently, out-competed through these allelopathic interactions, the invasive species is able
707 to establish and proliferate in the new environment, as described by Callaway & Aschehoug
708 (2000). Evidence for this hypothesis has been found in macroalgae as well. For example,
709 settlement of endemic brown, green and red macroalgal propagules, as well as settlement and
710 growth of benthic diatoms, was firmly inhibited on surfaces coated with 1,1,3,3-tetrabromo-2-
711 heptanone, an allelochemical isolated from the non-indigenous red algae *Bonnemaisonia*
712 *hamifera* (Svensson et al., 2013). Moreover, using field and laboratory experiments, *B.*
713 *hamifera* was found to actively transfer the allelochemical from its own thallus to its native host
714 algae at inhibitory concentrations (Svensson et al., 2013).

715 Currently, a large percentage of coral reefs globally are on a trajectory of decline, and some of
716 them are already severely degraded (De'ath et al., 2012; Del Monaco et al., 2017; Jackson et
717 al., 2014). One of the symptoms of this downturn is a reduced coverage of reef-building corals
718 and an increased occurrence and abundance of competing macroalgae (e.g., Bruno et al., 2009;
719 Hughes et al., 2010; Rogers and Miller, 2006). Further coral reef decline is expected due to
720 aggravated coral-seaweed competition as macroalgae proliferate and interactions with the
721 remaining corals increase in quantity and quality (Clements et al., 2020; Del Monaco et al.,
722 2017; Hughes et al., 2010). Evidence from both field and laboratory studies indicates that
723 several physical, chemical, and microbial processes can either directly or indirectly moderate
724 coral-seaweed competition, one of them being allelopathy (Clements et al., 2020; Del Monaco
725 et al., 2017). We refer to the review from Chadwick and Morrow (2011) who have described
726 all of these (in)direct mechanisms, both physical and chemical, in detail. Rasher and Hay (2010)
727 observed that 40 to 70 % of the investigated Caribbean and Pacific macroalgae species suppress
728 coral growth due to contact-mediated allelopathy. From literature, it appears that the
729 allelopathic character of macroalgae does not necessarily target a specific life stage of the
730 corals. For instance, the phaeophyte *Lobophora variegata* prevents settlement of larval *Porites*
731 *astreoides*, while coral larvae of *Acropora palifera* and *Stylophora pistillata* were not found to
732 metamorphose after surface contact with *Lobophora* spp. (Baird and Morse, 2004; Kuffner et
733 al., 2006). On the other hand, Rasher and Hay (2014) described a significant increase in coral
734 bleaching and a reduction of photosynthetic activity of zooxanthellae within 'adult' corals of
735 *Porites cylindrica* after exposure to hydrophobic extracts of the rhodophyte *Galaxaura*
736 *filamentosa*. Understanding the impacts of coral-macroalgal interactions on the resilience of
737 coral reefs, or the speed of their deterioration, is of crucial significance for conservation efforts
738 of reef ecosystems worldwide.

739 *Potential use of allelopathic agents as natural agrochemicals*

740 In agriculture, allelopathic compounds known to either suppress or eliminate competing plants
741 (crops) have gained specific interest, due to their prospect to be employed as selective natural
742 herbicides (Getachew et al., 2017; Vyvyan, 2002). Nowadays, this interest has partly shifted to
743 marine alternatives, like the development of antifoulants and biocides based on natural
744 products. For example, diterpenes extracted from the brown macroalgae *Dictyota menstrualis*
745 have been found to impede settlement and development of fouling bryozoans (Schmitt et al.,
746 1995). The bioactive compound 1,1,3,3-tetrabromo-2-heptanone, obtained from the rhodophyte
747 *Bonnemaisonia hamifera* through bioassay guided fractionation of hexane surface extracts, was
748 demonstrated to inhibit the growth, at naturally occurring concentrations, of several relevant
749 marine fouling bacteria, that were isolated from macroalgal surfaces (Nylund et al., 2008).
750 Alternatively, several authors have suggested to use the biocidal properties of some seaweed
751 species to steer towards the development and application of environment friendly and cost-
752 effective strategies to control ‘harmful algae blooms’ (HABs), the excessive growth of
753 deleterious micro- and macroalgae in response to coastal eutrophication (e.g., Nan et al., 2004;
754 Tang and Gobler, 2011; Wang et al., 2006; Table S1).

755 In contrast to many synthetic compounds, bioactive molecules that are produced by aquatic
756 organisms are both naturally degradable and show high efficiency at low concentrations (Nakai
757 et al., 1996, 2000; Jin & Dong, 2003; Jin et al., 2016). Moreover, indigenous allelopathic
758 species are relatively easy to collect at low cost and they are environmentally benign (Jin et al.,
759 2005; Tang and Gobler, 2011; Wang et al., 2012; Ye et al., 2014). Additionally, many
760 macroalgae taxa have a high nutrient assimilation capacity, hence, potentially simultaneously
761 reducing eutrophication levels of coastal ecosystems significantly, contributing to HAB
762 mitigation (Jin et al., 2005, 2016; Tang and Gobler, 2011; Ye et al., 2014). Both local/regional
763 control authorities as well as the International Maritime Organisation (IMO) legislation are

764 propulsing towards the design and implementation of natural, non-toxic antifoulants (Bazes et
765 al., 2009; Chambers et al., 2006).

766 *Aquaculture*

767 Allelochemicals of co-occurring seaweed might directly negatively impact the growth,
768 development and physiology of commercially important taxa, such as blue mussel *Mytilus*
769 *edulis* and oysters *Crassostrea gigas* and *C. virginica* (e.g., Green-Gavrielidis et al., 2018),
770 which is especially relevant in attempts to improve aquaculture yields is to enrich the culture
771 tanks with seaweed. For example, several macroalgae have already been combined into land-
772 based integrated multi-trophic aquaculture (IMTA) systems (e.g., Carl et al., 2014). On the
773 other hand, allelopathic seaweed species might negatively affect and even eliminate
774 competitors, foulers and grazers, co-occurring in the cultivation sites. For instance, *Saccharina*
775 *latissima* significantly lowered the saxitoxin accumulation in co-cultivated blue mussels by
776 significantly inhibiting the saxitoxin-producing *Alexandrium catenella* (Sylvers and Gobler.,
777 2021).

778 *Biodiscovery of allelochemicals*

779 In screening biologically active compounds derived from marine species, macroalgae have
780 received substantial attention as a natural source of metabolites with promising bioactive
781 properties. These metabolites have been observed to exhibit distinct biological activities, such
782 as cytotoxic, anthelmintic, antibiotic, antineoplastic, antioxidant, antibacterial, antifungal,
783 anti-inflammatory, and cholesterol-lowering activity (Lourenço-Lopes et al., 2020; Kabera et
784 al., 2014). More than 4,000 different metabolites have been extracted from mostly brown and
785 red macroalgae, but from some green seaweed species as well, in the last couple of decades,
786 and have been listed in the DNP, the Dictionary of Natural Products (Lourenço-Lopes et al.,
787 2020). Further characterisation of compounds with allelopathic activities in different species,
788 variation in their bioactivity, and a more profound comprehension of their mode of action, is

789 recommended to improve the ongoing examination on the potential use of allelochemicals as
790 pharmaceutical agents and bio-herbicides.

791 4. Conclusion

792 Macroalgae are a wealthy source of bioactive metabolites with allelopathic potential. To
793 discriminate bioactive metabolites from true allelochemicals it is crucial to verify whether (1)
794 the allelopathic properties are tested using ecologically relevant organisms; (2) the chemicals
795 are encountered by the target organism with reference to the macroalga (on its thallus's exterior
796 or in the surrounding seawater, but not stored inside the seaweed's blade); and (3) the putative
797 allelochemical is bioavailable and inhibits settlement/development/growth at the concentration
798 present at the seaweed surface/released into the water. Even though several macroalgal
799 compounds have been directly or potentially perceived as exhibiting allelochemical properties,
800 there is a clear necessity to improve and strengthen our knowledge on the identity of these
801 compounds, as well as their mode of action. Currently, photosynthesis inhibition in competing
802 photoautotrophs appears to be a frequently employed mechanism of action. Molecular
803 approaches can improve our knowledge on the identity of these compounds, as well as their
804 mode of action. In total, 138 macroalgae species have been described in literature that putatively
805 exhibit allelopathic properties, mostly belonging to a few dominating genera. The allelopathy
806 phenomenon in macroalgae should be further investigated considering several aspects from a
807 scientific and commercial point of view.

808

809

810 Declarations

811 **Ethical Approval**

812 Not applicable

813

814 **Competing interests**

815 The authors declare no competing interests.

816 **Conflict of interest**

817 The authors declare that they have no conflict of interest.

818 **Authors' contributions**

819 **Ilias Semmouri:** Data collection, data analysis, Writing - Original Draft, Review & Editing;

820 **Colin Janssen:** Supervision, Writing - Review & Editing; **Jana Asselman:** Supervision,

821 Writing - Review & Editing.

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825 **Availability of data and materials**

826 The datasets generated during and/or analysed during the current study are available from the
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828 **References**

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