

# **Characterization and comparative study on structural and physicochemical properties of buckwheat starch from 12 varieties**

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**Abstract:** Buckwheat is an important starch source because of its health benefits. In this study, buckwheat starches isolated from 12 varieties were analyzed based on the morphological, structural and physicochemical properties. The results showed that starch samples from different varieties had high purity with the total starch ranging from 91.29 to 95.11%, while showing significant differences in ash content (0.12-0.25%), protein content (0.26-0.34%) and amylose content (29.55-36.13%), respectively. All samples presented spherical and irregular shapes and typical A-type crystalline structure, but obvious differences in granule size distribution and relative crystallinity (26.37-35.21%) were observed among 12 varieties. Starch samples differed in lamellar structures, showing higher values of thickness of the samples with higher amylose content. In addition, buckwheat starches with higher amylose content showed higher values in light transmittance and rheological properties, while starch samples with lower amylose content obtained higher values in terms of water solubility, swelling power, pasting behaviors and thermal parameters. The principal component analysis and cluster analysis based on starch property parameters indicated that there were significant similarities and differences among 12 varieties, which might be related to the genotypes. This study would provide valuable information for the full use of buckwheat starch in food and non-food industries.

**Keywords:** Buckwheat; starch; structural properties; physicochemical properties

## 1 **1. Introduction**

2 Buckwheat is an annual dicotyledonous crop belonging to the genus *Fagopyrum*  
3 of the Polygonaceae family. It is widely recognized that buckwheat grain is rich in  
4 starch, protein, lipid, minerals, vitamins and dietary fiber (Ahmed, et al., 2014; Gao, et  
5 al., 2016). Buckwheat has also been considered as a source of herbal medicine for  
6 preventing and controlling the cardiovascular disease, obesity and cancer due to the  
7 high proportion of beneficial health components (phenolic compounds and phytosterols)  
8 (Kaur, Jha, Sabikhi, & Singh, 2014; Liu, Wang, Cao, Fan, & Wang, 2016). Recently,  
9 there has been an increasing emphasis on natural and healthy foods, making buckwheat  
10 an ingredient in functional food based on the low glycemic index. It has also been  
11 reported that buckwheat is a suitable food ingredient for different types of food such as  
12 noodles, pasta, biscuits and baking food (Yang, et al., 2019), showing good market  
13 potential in the functional and healthy food industry. Therefore, more research needs to  
14 be done on the food aspects of buckwheat.

15 Starch is mainly composed of amylose and amylopectin (Perez-Pacheco, et al.,  
16 2014), which can be used as a raw material or a food additive in developing food  
17 products or be applied as a delivery vehicle for substances of interest in the food and  
18 pharmaceutical industries (Ovando-Martinez, Bello-Perez, Whitney, Osorio-Diaz, &  
19 Simsek, 2011). Amylose is largely linear with a smaller molecular weight, while  
20 amylopectin is highly branched with relatively large molecular weight (Zhu, 2015). The  
21 amylose content and fine structure of amylopectin are critical for the physicochemical  
22 and functional properties of starch, thereby determining its application. Buckwheat

23 starch is the major component of the grain and appropriately accounts for 60-80% of  
24 the whole grain with about 25% amylose and 75% amylopectin (Qin, Wang, Shan, Hou,  
25 & Ren, 2010). Previous studies have shown that variations in amylose content and  
26 amylopectin chain length distribution of buckwheat starch result in differences in the  
27 light transmittance, swelling power, thermal and textural properties (Gao, et al., 2020;  
28 Hu, et al., 2022; Liu, et al., 2016). It has also been reported that the structure and  
29 properties of starch are critical for the quality of the buckwheat-based products (Zhu,  
30 2015). For example, amylose content is positively correlated with the elasticity of  
31 heated buckwheat dough due to the gelling capacity of amylose (Ikeda, Kishida, Kreft,  
32 & Yasumoto, 1997), and short chains of amylopectin is negatively correlated to the  
33 water solubility of buckwheat starch. Compared with maize and potato starch,  
34 buckwheat starch has the smallest granule size (3-14  $\mu\text{m}$ ) with lower water solubility  
35 and gelatinization enthalpy but higher gelatinization temperatures (Gao, et al., 2016).  
36 Kreft and Skrabanja (2002) have found that buckwheat starch had a slow glucose  
37 release rate and a large amount of resistant starch compared with other cereal starches,  
38 making it suitable for diabetic diets. The relationship between structure and properties  
39 of starch is a research hotspot. However, there have just few reports on the  
40 physicochemical properties of buckwheat starches, and these reports are on relatively  
41 fewer varieties. Therefore, understanding the relationships between the structural and  
42 physicochemical properties of buckwheat starches isolated from different varieties is  
43 essential for the development of starch-based products of buckwheat in food industry.

44 In this study, buckwheat starches with different amylose contents were isolated

45 from 12 varieties collected from 8 countries. The chemical composition, structural  
46 (morphological, crystalline structure, lamellar structure and short-range ordered  
47 structure) and physicochemical (water solubility, swelling power, light transmittance,  
48 pasting, thermal and rheological) properties were determined and compared, and the  
49 correlation between structural and physicochemical properties was investigated. The  
50 main aim of study was to reveal the relationship between structural and properties of  
51 buckwheat starch and provide useful information for starch production and utilization  
52 of buckwheat in food industry.

## 53 **2. Materials and methods**

### 54 *2.1 Materials*

55 A total of 12 buckwheat varieties collected from 8 countries were used in this study.  
56 All buckwheat varieties were planted in the Bottelare field (50°59'N, 3°49'E) of  
57 Belgium and harvested at maturity. An overview of the samples was listed in Table 1.

### 58 *2.2 Starch isolation*

59 Buckwheat starch was isolated from the hulled seeds following the method  
60 described by Hu, et al. (2022) with slight modifications. Buckwheat flour (500 g) was  
61 soaked in sodium hydroxide solution (0.2%, w/v) at a ratio of 1:3 and left at room  
62 temperature (25°C) for 16 h. Then, the samples were passed through a 150-mesh sieve  
63 and centrifuged at 4000 g for 10 min (the above step was repeated 3 times). Next, the  
64 supernatant was poured off and the white sediment from the bottom was washed with  
65 distilled water until it became clean. Finally, the samples were dried at 40°C, ground  
66 into powders and passed through a 100-mesh sieve.

67 *2.3 Analysis of chemical composition*

68 The moisture content was measured using the rapid moisture determination  
69 instrument (Sartorius, MA37-1). Ash content was determined according to standard  
70 method ICC no. 104/1. Protein content was measured using a Kjeldahl nitrogen  
71 analyzer and a factor of 6.25 was used to calculate the protein content. Amylose content  
72 and total starch content were quantified by the Amylose Assay Kit and Total Starch  
73 Assay Kit (Megazyme Ltd., Wicklow, Ireland), respectively.

74 *2.4 Cryo-scanning electron microscopy*

75 The starch samples were visualized using a JSM-7100F TTLS LV TFEG-SEM  
76 (Jeol Europe BV, Zaventem, Belgium). The starch powder was placed on a carbon  
77 double sided sticky tape that was fixed on an aluminum stub, vitrified in a nitrogen  
78 slush and transferred under vacuum conditions into a PP3010T cryo-preparation system  
79 (Quorum Technologies, East-Sussex, UK) conditioned at -140°C. Subsequently, the  
80 sample was sublimated for 10 min at -70°C to remove frost artefacts, sputter-coated  
81 with platinum using argon gas, transferred to the SEM stage at -140°C and electron  
82 beam targeted at 3 keV.

83 The images of buckwheat starch granules were further analyzed using ImageJ  
84 (National Institutes of Health, USA). The starch granules (50) with complete  
85 morphology were selected for labelling in each cryo-scanning electron microscopic  
86 image, and the obtained results of particle size were then made into the frequency  
87 distribution histograms.

88 *2.5 Granule size analysis*

89 The granule size of buckwheat starch was measured using a laser diffraction  
90 particle size analyzer (Malvern Instruments Ltd., Malvern, UK) equipped with a 300 F  
91 lens. Data analysis was conducted using the Mastersizer software, and the refractive  
92 index of real and imaginary particles was 1.45 and 0.1, respectively (Hellemans, et al.,  
93 2017). The granule size distribution was reported in terms of the volume distribution.

#### 94 *2.6 Wide angle X-ray scattering (WAXS)*

95 The WAXS pattern of buckwheat starch was determined using an X-ray scattering  
96 instrument (GeniX <sup>3D</sup> Cu HFL, Xenocs, France) following the method of Zhang, et al.  
97 (2018). The XRD patterns were recorded from 5° to 50° (2θ) with a scanning speed of  
98 1.2°/s. XSACT software (Xenocs, France) was used to normalize the results. The  
99 relative crystallinity (RC) was the ratio of the crystallinity area to the total diffraction  
100 area.

#### 101 *2.7 Small angle X-ray scattering (SAXS)*

##### 102 *2.7.1 SAXS measurement*

103 The SAXS test was conducted by a small-angle X-ray scattering instrument  
104 (GeniX <sup>3D</sup> Cu HFL, Xenocs, France). The optics and sample chamber were under  
105 vacuum to reduce air scattering. The 1D scattering curves were in the range of  $0 < q <$   
106  $0.3 \text{ \AA}^{-1}$  from the 2D scattering patterns.

##### 107 *2.7.2 SAXS analysis*

108 The obtained data was calibrated from the background scattering using the  
109 XSACT software (Xenocs, Sassenage, France). The data was further analyzed to  
110 calculate the lamellar parameters, through the normalized 1D correlation function as

111 described by Kuang, et al. (2017) based on the following equation (1):

$$112 \quad L(r) = \frac{\int_0^\infty I(q)q^2 \cos(qr) dq}{\int_0^\infty I(q)q^2 dq} \quad (1)$$

113 Where  $I(q)$ ,  $q$  and  $r$  were scattering intensity, scattering vector and the direction  
114 along the lamellar stack, respectively.

### 115 *2.8 Water solubility and swelling power*

116 The water solubility (WS) and swelling power (SP) of buckwheat starch were  
117 determined following our previous method (Gao et al., 2020). The WS (%) and SP (g/g,  
118 dry basis) was calculated as follows:

$$119 \quad \text{WS} = \text{mass of dried supernatant/mass of dry starch} \times 100\% \quad (2)$$

$$120 \quad \text{SP} = \text{sediment weight/mass of dry starch} \times (100 - \text{WS}) \quad (3)$$

### 121 *2.9 Light transmittance*

122 The starch sample (0.2 g) and distilled water (20 mL) were mixed and heated in  
123 boiling water for 30 min. After the samples were cooled to 25°C, the light transmittance  
124 (LT) was measured at 620 nm using a spectrophotometer with distilled water as a  
125 control (Gao, et al., 2020).

### 126 *2.10 Pasting properties*

127 Pasting profiles of buckwheat starch were measured using a Rheometer MCR 102  
128 (Anton Paar GmbH, Graz, Austria) through the method of Hellemans et al. (2017) with  
129 slight modifications. A 6% (w/v) starch-water suspension corrected for its moisture  
130 content was prepared for the measurements. After the pre-shearing, the suspension was  
131 held at 50°C for 1 min and then heated to 95°C at a rate of 5°C/min, held at 95°C for 5  
132 min, cooled to 50°C at the same rate and finally held at 50°C for 2 min. The pasting

133 parameters, including peak viscosity (PV), holding strength (HS), final viscosity (FV),  
134 breakdown (BD), setback from peak (SBP), setback from trough (SBT) and pasting  
135 temperature (PT), were automatically obtained through the RheoCompass software.

### 136 *2.11 Thermal properties*

137 The thermal properties of buckwheat starch were performed by differential  
138 scanning calorimetry (DSC) (Q1000, TA instruments, New Castle, DE, USA) following  
139 the method of Guo, et al. (2019) with slight modifications. Briefly, starch and distilled  
140 water were mixed into suspension at a ratio of 1:3, and the samples were sealed in an  
141 aluminum pan at 4°C overnight. Then, the samples were heated from 30 to 100°C at a  
142 rate of 10°C/min with an empty pan as a reference. The transition temperatures (onset,  
143  $T_o$ ; peak,  $T_p$  and conclusion,  $T_c$ ) and gelatinization enthalpy ( $\Delta H$ ) were obtained from  
144 the DSC curve.

### 145 *2.12 Rheological properties*

146 The rheological analysis was performed using a Rheometer MCR 102 (Anton Paar  
147 GmbH, Graz, Austria) following the method of Jiang, et al. (2020) with some  
148 modifications. The starch suspension (8%, w/v) was cooked in boiling water for 15 min.  
149 After the samples were cooled to room temperature (25°C), the starch gel was loaded  
150 onto the bottom plate at 25°C combined with a thin layer of silicone oil to reduce  
151 evaporation loss. The strain sweep test was carried out to determine the linear  
152 viscoelastic range (LVR).

#### 153 *2.12.1 Frequency sweep*

154 The frequency sweep of buckwheat starch was conducted from 0.1 to 100 rad/s at

155 1% strain that was within LVR. The storage modulus ( $G'$ ), loss modulus ( $G''$ ), complex  
156 viscosity ( $\eta^*$ ) and loss angle ( $\tan \delta = G''/ G'$ ) were recorded.

157 The obtained data could be analyzed by a Power law model with the following  
158 formula (Li, et al., 2021):

$$159 \quad G' = K' \times \omega^{n'} \quad (4)$$

$$160 \quad G'' = K'' \times \omega^{n''} \quad (5)$$

161 Where  $K'$  and  $K''$  represented model constants ( $\text{Pa/s}^n$ ),  $n'$  and  $n''$  were the frequency  
162 modulus exponents (dimensionless), and  $\omega$  was the frequency (rad/s).

### 163 *2.12.2 Creep-recovery test*

164 The creep-recovery test was studied with the constant stress of 1 Pa for 300 s. Then,  
165 the applied stress was removed and the performance was recorded for another 600 s.  
166 The obtained data was fitted using the four-parameter Burger's model (Zhao, Li, Wang,  
167 & Wang, 2022):

$$168 \quad J(t) = 1/G_0 + 1/G_1(1 - e^{-t/\lambda}) + t/\mu_0 \quad (6)$$

169 Where  $J$ ,  $G_0$ ,  $G_1$ ,  $\lambda$  and  $\mu_0$  was the creep compliance (1/Pa) at  $t$  time, the  
170 instantaneous elastic modulus (Pa), the retarded elastic modulus (Pa), the retardation  
171 time (s) and the viscous modulus (Pa s), respectively.

### 172 *2.13 Statistical analysis*

173 The results were expressed as means  $\pm$  standard deviations. One-way analysis of  
174 variance (ANOVA) and Duncan's multiple-range test ( $p < 0.05$ ) were conducted using  
175 SPSS software (v. 22.0, IBM, USA) for analyzing the significant difference among the  
176 data. Principal component analysis (PCA) and cluster dendrogram analysis based on

177 single-linkage were performed using the OriginPro software (v. 2021, Originlab, USA)  
178 to determine the similarities and differences among 12 buckwheat starches.

### 179 **3. Results and discussion**

#### 180 *3.1 Main chemical composition*

181 The main chemical compositions of buckwheat starch collected from different  
182 countries are summarized in Table 1. The yield of 12 buckwheat starches ranged from  
183 22.48 to 31.58%. There was significant difference in the total starch content ranging  
184 from 91.29 (BU9) to 95.11% (BU5), indicating that the purity of the sample was  
185 reasonably high (> 90%). The moisture content was between 8.43 and 13.65% with the  
186 lowest value in BU3 and the highest value in BU5, which was within the moisture level  
187 recommended for commercial starches (Soni, Sharma, & Gharia, 1993). Differences in  
188 moisture content among 12 buckwheat varieties may be due to the degree of starch  
189 drying. Significant differences were observed in ash content ranging from 0.12 (BU1)  
190 to 0.25% (BU5), which was similar to the results of quinoa starch (Jiang, et al., 2020)  
191 but slightly lower than that in sweet potato starch (Abegunde, Mu, Chen, & Deng, 2013).  
192 The protein content of 12 starch samples was significantly different ranging from 0.26  
193 (BU6) to 0.34% (BU5). Normally, the proteins in starch granules are mainly surface  
194 proteins and internal proteins. The former can be easily removed, while the removal of  
195 the latter requires the destruction of the starch granule structure (Swinkels, 1985). It has  
196 been reported that the isolation process can influence the protein content and that  
197 surface proteins can be removed from starch granules with NaOH solutions (Guo, et al.,  
198 2019), which can be used to explain the low protein content of buckwheat starch of this

199 study. In addition, the samples were low in ash and protein content, which met the  
200 experimental requirements for the absence of non-starch lipids and hydrated fine fibers  
201 (Jan, Panesar, Rana, & Singh, 2017). Significant differences were also observed in  
202 amylose content ranging from 29.55 to 36.13%, with the lowest value in BU3 and the  
203 highest value in BU8. The result of amylose content in this study was lower than that  
204 of previous results (Gao, et al., 2020), indicating that buckwheat variety can influence  
205 the amylose content of starch. It has been reported that amylose content has a crucial  
206 effect on the functional characteristics of starch, and the differences are mainly related  
207 to genotype background, growing environment, and measuring method (Zhang, et al.,  
208 2018).

### 209 *3.2 Morphological properties and particle size distribution*

210 The morphology of buckwheat starch granule was observed using cryo-scanning  
211 electron microscope at two different magnifications of 1000 and 5000 (Fig. 1). Most  
212 starch granules were irregular polygons with obvious edges and a few granules  
213 appeared spherical shape, which was consistent with previous studies on buckwheat  
214 starch of different varieties (Gao, et al., 2016; Gao, et al., 2020; Hu, et al., 2022). At  
215 high magnification, some hollows were observed on the surface of the starch granule,  
216 which could be explained by the fingerprints of the native protein bodies (Dura,  
217 Blaszcak, & Rosell, 2014). The image analysis showed that the granule sizes of 12  
218 buckwheat starches followed a normal distribution (Fig. 1), and there were significant  
219 differences in the granule size among different varieties with the maximum value in  
220 BU3 (7.29  $\mu\text{m}$ ) and the minimum value in BU8 (6.02  $\mu\text{m}$ ).

221 The volume distribution and standard average diameter can be obtained by using  
222 the laser diffraction particle size measuring instrument, assuming that the particles are  
223 spherical. As shown in Fig. 2 A, a smooth curve with two peaks was observed for the  
224 volume distribution of 12 starch samples with the weak peaks showing at 1  $\mu\text{m}$  and the  
225 strong peaks occurring at about 10  $\mu\text{m}$ . There were significant variations in the volume  
226 distribution among different varieties, showing the largest volume distribution in BU10  
227 and the smallest distribution in BU11. The size of most starch granules ranged from 3  
228 to 20  $\mu\text{m}$ , smaller than that of maize starch and sweet potato starch (Lin, et al., 2016;  
229 Zhang, et al., 2018). The D [4,3] ranged from 7.158 (BU8) to 8.576 (BU3)  $\mu\text{m}$  and the  
230 D [3,2] was in the range of 4.052 (BU12) to 4.583  $\mu\text{m}$  (BU3), slightly lower than the  
231 results of the previous study (Gao, et al., 2020). The d (0.1), d (0.5) and d (0.9) were in  
232 the range of 2.590-3.569, 7.251-8.307 and 10.883-16.558  $\mu\text{m}$ , respectively, with the  
233 maximum value in BU3 and the minimum value in BU8 (Table 2). It has been reported  
234 that starch granule size plays an important role in affecting the pasting behaviors of  
235 starch. Abegunde et al. (2013) have found that the granule size of sweet potato starch  
236 was positively correlated with the PV, BD and SB, which was consistent with the results  
237 of this study as shown in Fig. 6 A. The granule size of starch can be affected by the  
238 variety, growing condition and plant physiology (Guo et al., 2019). In this study, the 12  
239 buckwheat starch samples were planted in the same experimental site, indicating that  
240 the variations in particle size distribution of 12 starch samples due to the various  
241 genotype backgrounds.

### 242 3.3 WAXS

243 Generally, the X-ray scattering is widely used to study the helical structures of  
244 starch crystals at longer range scales (Kuang, et al., 2017). The XRD patterns of  
245 buckwheat starches and their relative crystallinities are shown in Fig. 2 B. The typical  
246 A-type crystalline structure can be observed with strong diffraction peak at around  $15^\circ$   
247 and  $23^\circ 2\theta$  and an unresolved peak at  $17^\circ$  and  $18^\circ 2\theta$ , which was consistent with the  
248 results of normal cereal starches (Cheetham & Tao, 1998). It has been reported that the  
249 peak intensity at  $2\theta = 5.4^\circ$  represents the B-type polymorphic form and the peak  
250 intensity at  $2\theta = 20^\circ$  corresponds to the amylose-lipid complex. In this study, slight  
251 difference was obtained in the peak positions of starch samples, which might be due to  
252 the genotypes and amylose content among different buckwheat varieties. As shown in  
253 Fig. 2 B, there were significant variations in the RC of 12 starch samples ranging from  
254 26.37% in BU8 to 35.21% in BU3, indicating that the BU3 presented more crystalline  
255 regions in comparison with other buckwheat varieties. The difference in the RC among  
256 12 buckwheat starches might be related to the variations in granule size and chemical  
257 compositions (Table 1). Compared with maize starch and bean starch (Lin, et al., 2016;  
258 Ovando-Martinez, et al., 2011), buckwheat starch showed the highest value in the RC,  
259 which could be related to the genotypes. The crystalline region of starch can be affected  
260 by the structure and content of amylopectin molecules, while the amorphous region is  
261 related to amylose molecules. In this study, the RC of buckwheat starch was negatively  
262 correlated with the amylose content (Fig. 6 A), which was similar to the results of maize  
263 starch (Cheetham, et al., 1998).

#### 264 3.4 SAXS

265 The variations of the lamellar structure of buckwheat starches were further  
266 determined through the small angle X-ray scattering (SAXS). The SAXS one-  
267 dimensional (1D) scattering intensity distribution of various starch samples are  
268 presented in Fig. 2 C. One “shoulder-like” scattering peak was observed around the  $q$   
269 value of 0.56-0.73  $\text{nm}^{-1}$  in each SAXS curves, exhibiting difference for the peak  
270 position among 12 buckwheat varieties. It has been reported that the scattering peak  
271 represented a long period in starch granules, and the position of the SAXS peak  
272 correlates with the average total thickness of the crystalline and amorphous regions in  
273 lamellar arrangements (Blazek & Gilbert, 2011). The scattering intensity is proportional  
274 to the square of electron density at the corresponding scale, that is, the peak intensity is  
275 related to the  $\Delta\rho$  and  $\Delta\rho_u$  (Tan, et al., 2015).  $\Delta\rho$  indicates the difference in electron  
276 density between the amylopectin crystalline lamella ( $\rho_1$ ) and amylose/amylopectin  
277 based amorphous region ( $\rho_3$ ), which is helpful to increase the overall intensity (Zhu,  
278 2015).  $\Delta\rho_u$  represents the difference in electron density between amylose background  
279 region ( $\rho_2$ ) and  $\rho_3$ , which is related to the low-angle intensity (Yu, et al., 2022). These  
280 results indicated that the electron density varied between different buckwheat starches.

281 Lorentz correction was used to clearly analyze the peak position, and the corrected  
282 SAXS curves were shown in Fig. 2 D. The peak intensity presented a “shoulder-like”  
283 position at around 0.6-0.7  $\text{nm}^{-1}$  with the maximum value in BU1 and the minimum  
284 value in BU7. The increased peak intensity indicated that there was also increase for  
285 the contrast of electron density, and the differences in 12 samples might be related to  
286 the variations in water absorption and swelling of the amorphous fraction and/or

287 leaching of amylose from the amorphous parts (Kuang, et al., 2017).

288 The correlation function can be used to analyze the starch aggregation structure  
289 and can provide the structure parameters of lamellar structures, including the crystalline  
290 layer thickness ( $d_c$ ), amorphous layer thickness ( $d_a$ ) and long period distance ( $d_{ac} = d_a$   
291  $+ d_c$ ) (Chen, et al., 2016). The normalized 1D correlation function is shown in Fig. 3  
292 and the lamellar structure parameters are summarized in Table 3. According to the  
293 Bragg's formula ( $d = 2\pi/q$ ), significant difference was observed in the thickness of the  
294 semi-crystalline layers ( $d_{Bragg}$ ) among 12 starch samples, ranging from 8.465 (BU9) to  
295 11.310 nm (BU8). There were also obvious variations in the  $d_{ac}$ , with BU8 having the  
296 largest value of 11.70 nm and BU9 having the lowest value of 8.56 nm, indicated that  
297 the  $d_{ac}$  from the correlation function had a proper fitting with  $d_{Bragg}$  from the Bragg's  
298 equation. The  $d_a$  was in the range of 3.20-4.85 nm with the largest amorphous thickness  
299 in BU8 and the lowest amorphous thickness in BU9. For the  $d_c$ , it was between 5.29  
300 and 6.85 nm with the order of BU8 > BU1 > BU5 > BU10 > BU3 > BU4 > BU11 >  
301 BU2 > BU12 > BU6 > BU9 > BU7. These results showed that there were significant  
302 variations in lamellar structure of buckwheat starch. Lan et al. (2017) have found that  
303 there was positive correlation between thickness layer and light transmittance but  
304 negative correlation between thickness layer and resilience of canna starch. Ma et al.  
305 (2022) have reported that the starch gelatinization of wheat starch can be affected by  
306 the lamellar structure. During cooking, the granular and lamellar structures of starch  
307 are fully gelatinized, thereby influencing the digestibility. Therefore, the suitable  
308 varieties of buckwheat should be selected based on the specific needs in food processing

309 and production. For example, buckwheat starches with low thickness are more suitable  
310 for the production of food additives, while starch samples with high thickness can be  
311 used to make adhesives.

### 312 *3.5 Water solubility and swelling power*

313 Water solubility can reflect the dissolution degree of starch during swelling and  
314 swelling power is used to measure the water holding capacity (Carcea & Acquistucci,  
315 1997). The WS and SP of buckwheat starches at different temperatures are summarized  
316 in Table S1. The results showed that the WS and SP values of 12 starch samples varied  
317 at different temperatures, and the values of WS and SP significantly increased with the  
318 increase of temperature. At 75°C, the WS and SP were both low (the average WS was  
319 6.04% and the average SP was 12.85 g/g). After 75°C, the WS and SP sharply increased  
320 with increasing temperature, with the average value of 10.76% and 19.41 g/g,  
321 respectively. When the temperature reached 95°C, both the WS and SP showed the  
322 maximum value, with an average WS of 12.52% and an average SP of 22.05 g/g,  
323 respectively. Similar change trend was observed in quinoa starch and sweet potato  
324 starch (Jiang, et al., 2020; Zhang, et al., 2018). The relationships between WS and SP  
325 of buckwheat starch showed a linear relationship, and the SP significantly increased  
326 with the increase of the WS at different temperatures (Fig. 4 A). In addition, the linear  
327 relationship was more significant at high temperatures, indicating that increasing  
328 temperature was helpful to promote the absorption and expansion of buckwheat starch  
329 granules. Generally, the water solubility and swelling power of cereal starches are used  
330 to study the interaction between water molecules and starch chains in crystalline and

331 amorphous regions during heating (Abegunde, et al., 2013). The extent of this  
332 interaction can be affected by the amylose content, amylose to amylopectin ratio and  
333 fine structure of amylopectin, resulting in variations in water solubility and swelling  
334 power (Kaur, Singh, McCarthy, & Singh, 2007). It has been concluded that amylose  
335 could inhibit the starch swelling, hinder the breakage of amylopectin double helix, and  
336 maintain the integrity of swollen granules (Lai, et al., 2016). In this study, the amylose  
337 content of buckwheat starch was negatively correlated with the WS ( $P = -0.4256$ ) and  
338 SP ( $P = -0.5027$ ) (Fig. 6 A), for example, the variety of BU3 with the lowest amylose  
339 content (Table 1) had the highest values of WS and SP (Table S1), which can be used  
340 to explain the variations in the WS and SP among 12 starch samples. In addition,  
341 differences in genetics and growing areas of buckwheat starch also contributed to the  
342 changes in the WS and SP.

### 343 *3.6 Light transmittance*

344 Light transmission can be used to indicate the clarity of the starch paste, reflecting  
345 the retrogradation process (Huang, et al., 2021). The results of the light transmittance  
346 (LT, %) of buckwheat starches are displayed in Fig. 4 B. It was clearly shown that there  
347 were significant differences in the LT among 12 starch samples, ranging from 18.48 to  
348 26.98%, with the largest value in BU3 and the lowest values in BU8, indicating that the  
349 starch granules of BU3 had the largest dispersion in water, leading to the highest light  
350 transmittance. In addition, the LT of all buckwheat starches was above 18%, which was  
351 significantly higher than the results of the previous study (Gao, et al., 2020), and this  
352 difference might be contributed to the varieties and growing conditions. It has been

353 reported that increasing the amorphous area could make it easier for water molecules  
354 to enter the starch and make starch granules expand and disperse better in water, thus  
355 reducing the light refraction and dispersion and increasing the light transmittance of the  
356 starch paste (Hu, et al., 2016). In this study, buckwheat starch with higher amorphous  
357 area showed lower value of the LT, and there was a significant negative correlation  
358 between the LT and amylose content (Fig. 6 A), which was consistent with the above  
359 conclusion. The LT of cereal starch can be influenced by the swelling power ability,  
360 arrangement of molecular structure and the ratio of amylose/amylopectin (Jacobson,  
361 Obanni, & Bemiller, 1997), which can be used to explain the variations in the LT of 12  
362 different buckwheat starches.

### 363 *3.7 Pasting properties*

364 Pasting behavior is helpful to determine the quality and utilization of starch  
365 (Abegunde, et al., 2013; Sun, et al., 2021). The viscosity profiles of buckwheat starches  
366 are presented in Fig. 4 C, and the pasting parameters are summarized in Table 4. It was  
367 shown that all starch samples exhibited a smooth curve with significant variations in  
368 their pasting behaviors among different buckwheat varieties (Fig. 4 C). PV ranged from  
369 601 to 862 mPa·s, showing the largest value in BU3 and the lowest value in BU8.  
370 Holding strength (HS) is the difference between peak viscosity and breakdown viscosity.  
371 The HS of 12 starch samples was between 535.70 and 753.47 mPa·s, with the maximum  
372 in BU1 and the minimum in BU8. Final viscosity (FV) is due to the reduced movement  
373 of water molecules surrounded by amylose and amylopectin as the temperature  
374 decreases and the viscosity increases again, reflecting the stability to swollen granule

375 structure. In this study, the FV was in the range of 1004.37-1537.33 mPa·s,  
376 corresponding to BU8 and BU3, respectively, which was slightly lower than the results  
377 of buckwheat starches reported by (Gao, et al., 2016). The difference might be related  
378 to the genotypes, starch purity and the interaction among starch components.  
379 Breakdown viscosity (BD) can reflect the heat resistance and shear resistance of starch  
380 paste, and starch with higher value means lower resistance to heat (Guo, et al., 2019).  
381 The BD ranged from 29.07 to 112.60 mPa·s with the lowest in BU6 and the largest in  
382 BU3, which indicated that BU3 contained lower resistance to heat and shear and was  
383 much easier to gelatinize during the heating process. Pasting temperature (PT) refers to  
384 the temperature where starch viscosity begins to rise. The PT of buckwheat starch  
385 significantly varied from 66.42 (BU3) to 71.13°C (BU8) with a mean value of 68.55°C.  
386 It has been reported that pasting temperature of starch is positively correlated with  
387 amylose content (Zhou, Shi, Meng, & Liu, 2013), which was similar to the results of  
388 this study as presented in Fig. 6 A. Differences in the PT of buckwheat starch might be  
389 related to the variations in granule size distribution as shown in Table 2. The results of  
390 the PT in this study were higher than the PT range (59.12-63.9°C) reported in previous  
391 studies (Gao, et al., 2016; Gao, et al., 2020; Liu, et al., 2016) but lower than that of  
392 quinoa starch (72.60°C) (Jiang, et al., 2020). In this study, the pasting properties of  
393 buckwheat starches were significantly different among various varieties, which might  
394 be related to the granule size, amylose content and chain length distribution of  
395 amylopectin. Therefore, the suitable buckwheat variety should be selected to achieve  
396 the desired properties in food industry.

397 *3.8 Thermal properties*

398 The thermal properties of buckwheat starches were analyzed using DSC, the  
399 thermograms are presented in Fig. 4 D, and the thermal parameters are summarized in  
400 Table 4. All starch samples showed smooth thermogram curves, while the positions of  
401 peaks and the degrees of peak openings were significantly different. Similar results  
402 have been reported in sweet potato starch (Guo, et al., 2019). Significant differences  
403 were also observed in the thermal parameters of this study (Table 4). Both the  $T_o$  and  
404  $T_c$  showed the lowest value in BU8 and the highest value in BU3, ranging from 55.70  
405 to 62.53°C and from 73.60 to 80.32°C, respectively. The  $T_p$  was in the range of 66.85-  
406 70.45°C, with the minimum value in BU12 and the maximum value in BU3. Starch  
407 with higher gelatinization transition temperatures would require higher heat of  
408 solubilization (Vasanthan, Bergthaller, Driedger, Yeung, & Sporns, 1999). The results  
409 observed in this study indicated that the BU12 was the easiest of the 12 buckwheat  
410 varieties to heat from the crystalline state to the gel state. Gelatinization enthalpy ( $\Delta H$ )  
411 reflects the melting of starch crystals, and a relatively high value means that much  
412 energy is needed to melt starch granules (Gao, et al., 2016). Higher crystallinity degree  
413 can lead to higher transition temperatures, making the starch granules more resistant to  
414 gelatinization (Uarrota, et al., 2013). In this study, the  $\Delta H$  varied from 6.44 (BU8) to  
415 8.92 J/g (BU3) with mean value of 7.43 J/g, suggesting that BU3 possessed more stable  
416 crystal structure and was more difficult to melt, which was consistent with the results  
417 of the RC and granule size distribution as shown above. The  $\Delta H$  values previously  
418 reported for buckwheat starches by Gao, et al. (2020) were similar to the values reported

419 in this study. Differences in the gelatinization parameters of starch are thought to be  
420 influenced by the main chemical compositions, granule size and the molecular structure  
421 of the crystalline region (Kaur, et al., 2007).

### 422 *3.9 Rheological properties*

#### 423 *3.9.1 Frequency sweep analysis*

424 Frequency sweep test was conducted to determine the viscoelastic properties of  
425 buckwheat starch. The storage modulus ( $G'$ ), loss modulus ( $G''$ ), complex viscosity ( $\eta^*$ )  
426 and loss angle ( $\tan \delta$ ) as a function of the angular frequency ( $\omega$ ) of buckwheat starch  
427 are displayed in Fig. 5 A-C. Generally,  $G'$  and  $G''$  represent the elastic behavior (solid-  
428 like system) and the viscous behavior (liquid-like system), respectively (Guo, Tao, Cui,  
429 & Janaswamy, 2019). If  $G''$  exceed  $G'$  (large  $\tan \delta$ ), the gel behaves more like a liquid  
430 because the energy used to deform the gel is viscous dissipated (Zhang, et al., 2022). In  
431 this study, the  $G'$  exceeded  $G''$  and without any intersection within the angular  
432 frequency range of 0.1-100 rad/s (Fig. 5 A), indicating the solid-like systems for  
433 buckwheat starch. In addition, there were significant differences in the  $G'$  and  $G''$  of the  
434 12 starch samples with the largest value in BU8 and the lowest value in BU3, which  
435 was probably related to the various sensitivity of buckwheat starch to frequency. As  
436 shown in Table S2, the  $G'$  and  $G''$  were well fitted by Power law model.  $K'$  and  $K''$   
437 reflects the viscous and elastic behaviors, respectively, and  $n$  is related to the frequency  
438 dependence. The fitted results showed that the gel of buckwheat starch exhibited a  
439 solid-like system. As can be seen from Fig. 5 B, the  $\eta^*$  decreased sharply within low  
440 angular frequencies and then gradually stabilized at high angular frequency. Loss angle

441 ( $\tan \delta$ ) reflects the relative contribution of the viscous part and elastic part to the  
442 viscoelastic response of starch samples (Ma, Zhang, Jin, Xu, & Xu, 2022). As shown  
443 in Fig. 5 C, the  $\tan \delta$  firstly increased and then decreased with the increase of angular  
444 frequency and ranged from 0.01 to 0.15 (lower than 0.4), indicating that all starch  
445 samples behaved as an elastic material. Han plot was used to further analyze the  
446 different buckwheat starch pastes (Fig. 5 D), and the Han plot firstly decreased and then  
447 significantly increased with the increase of  $G''$  and the relaxation mechanism of  
448 buckwheat starch was longer in the high  $G''$  region.

### 449 *3.9.2 Creep-recovery analysis*

450 To further analyze the variations in the viscoelastic behavior of buckwheat starch,  
451 creep-recovery test was conducted to record the strain and compliance over time. As  
452 shown in Fig. 5 E, when subjected to instantaneous stress, the strain of all starch  
453 samples increased with time (within 300 s), and then significantly decreased after the  
454 stress was removed. There were significant differences in the strain among 12  
455 buckwheat starches, showing the maximum value in BU4 and the minimum value in  
456 BU8, which indicated that BU4 possessed more viscous components, resulting in lower  
457 formability. After removing the stress, all samples exhibited stable strain, suggesting  
458 that the strain of buckwheat starch was well recovered.

459 As can be seen from Fig. 5 F, all samples showed a nonlinear response to stress.  
460 The compliance of buckwheat starches ranged from 0.01 to 0.03 J with the largest value  
461 in BU4 and the lowest value in BU8. The creep compliance can reflect the freedom  
462 extent of molecular movement and the strength of gel, and higher compliance values

463 correspond to a weak gel system with high deformability (Samutsri & Suphantharika,  
464 2012). The finding of this study suggested that the gel system of BU8 was the strongest  
465 of the 12 buckwheat varieties. Differences in various buckwheat varieties were  
466 probably due to the molecular weight and the shorted chain length of starch granules.  
467 In addition, the creep compliance curves were well fitted by Burger's model with the  
468 range of  $R^2$  from 0.915 to 0.996, and there were significant differences in the  $G_0$ ,  $G_1$ ,  
469  $\lambda$  and  $\mu_0$  among 12 buckwheat starches (Table S2). These results indicated that  
470 buckwheat starch showed solid and elastic gel network structure, which was consistent  
471 with the frequency sweep tests.

### 472 *3.10 Principal component analysis (PCA)*

473 The PCA is widely used to investigate the interrelationships between the structural  
474 properties of starch and the differences and similarities among starches collected from  
475 different sources (Kaur, et al., 2007). The relationship between the structural and  
476 physicochemical properties of 12 buckwheat starches were subjected to PCA, the  
477 loading and score plots are presented in Fig. 6 and the Pearson's correlation coefficients  
478 are shown in Fig. S1. The first principal component (PC1) and second principal  
479 component (PC2) are considered to explain the variance of data when multidimensional  
480 data is projected as one-dimensional data (Lee, Lee, & Chung, 2017). In this study, the  
481 PC1 and PC2 accounted for 65.92% and 12.30%, respectively. The correlations among  
482 the relative properties could be observed from the loading plot (Fig. 6 A). The curves  
483 that are close to each other on the plot are positively correlated, while those are in  
484 opposite directions are negatively correlated (Kaur, et al., 2007; Singh, McCarthy,

485 Singh, & Moughan, 2008). Among the main chemical compositions, Ts and Pr were  
486 loaded negatively on PC1 but positively on PC2, while Mo, As and Am were all in the  
487 negative direction of PC1 and PC2. Among the structural properties, RC was loaded  
488 positively on PC1 but negatively on PC2, while da, dc and dac were loaded negatively  
489 on PC1 but positively on PC2. Among the pasting properties, PV and HS were loaded  
490 positively on PC1 but negatively on PC2, PT was loaded negatively on PC1 but  
491 positively on PC2, while FV, BD, SBP and SBT were all in the positive direction of  
492 PC1 and PC2. Among the thermal properties, To, Tp and Tc were loaded positively on  
493 PC1 but negatively on PC2, while  $\Delta H$  were loaded positively on both PC1 and PC2.

494 The distance between any two starches on the score plot refers to the similarities  
495 and differences among the starches. A total of 12 buckwheat starches collected from 6  
496 countries are regularly distributed in the quadrants of the score plot with four groups  
497 based on the structural and physicochemical properties (Fig. 6 B). For example, BU1  
498 and BU3 had positive scores on PC1 and PC2, while opposite trend was observed on  
499 BU4, BU9 and BU12; BU5, BU8 and BU10 were loaded at the left of the score plot  
500 with positive scores on PC2, whereas BU2, BU6, BU7 and BU12 showed positive  
501 scores on PC1 but negative scores on PC2. Overall, the 12 buckwheat starches with  
502 different amylose content were clearly classified according to their different properties.

### 503 *3.11 Cluster analysis*

504 In order to compare the relationships of different buckwheat varieties, the  
505 hierarchical cluster was performed based on chemical compositions, granule size  
506 distribution, relative crystallinity, lamellar parameters, water solubility, swelling power,

507 light transmittance, pasting properties and thermal parameters. As shown in Fig. 6 C,  
508 the dendrogram consisted of two major clusters and these two clusters were separated  
509 by the distance of 3.35. One cluster just contained one buckwheat variety (BU3), and  
510 other 11 varieties were contained in other groups. It was obvious that the remained 11  
511 varieties could be further separated into two groups. One group included BU1, BU2,  
512 BU6, BU7, BU9 and BU11, and the other group contained BU4, BU5, BU8, BU10 and  
513 BU12. Furthermore, BU1 can be separated from the former group at the distance of  
514 2.10, and BU8 can be separated from the latter group at the distance of 1.85. These  
515 results indicated that buckwheat starches isolated from different varieties showed  
516 various structural and physicochemical properties, which was consistent with the results  
517 of the PCA analysis.

#### 518 **4. Conclusions**

519 Starch characteristics of 12 buckwheat varieties collected from 8 countries were  
520 investigated in this study. The results showed that the contents of moisture, ash, protein,  
521 amylose and total starch varied from 8.43 to 13.65%, 0.12 to 0.25%, 0.26 to 0.34%,  
522 29.55 to 36.13% and 91.29 to 95.11%, respectively. All starch samples showed irregular  
523 polygonal and spherical shapes and typical A-type crystalline structure, while had  
524 obvious differences in crystallinity ranging from 26.37% (BU8) to 35.21% (BU3).  
525 Among the 12 buckwheat starches, BU3 with the lowest value in amylose content  
526 presented higher values of water solubility, swelling power, light transmittance, pasting  
527 properties, thermal parameters and loss angle. However, starch with higher amylose  
528 content showed higher values of amorphous region, storage modulus, loss modulus and

529 complex viscosity. PCA analysis and cluster analysis showed that there were significant  
530 differences in structural and physicochemical properties among various buckwheat  
531 varieties. In all, buckwheat starches with high amylose content (such as BU8, BU10  
532 and BU12) were more suitable as a food packaging material or food additive, while the  
533 starch samples with low amylose content (such as BU1 and BU3) can be used as an  
534 adhesive, which would provide valuable information for the further utilization of  
535 buckwheat starch in food and non-food industries based on the specific genotypic  
536 sources.

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### 543 **Declaration of competing interest**

544 The authors declare that there is no conflict of interests regarding the publication  
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## 696 **Figure and Table legends**

697 **Fig. 1** Cryo-scanning electron microscopy images and relative image analysis of  
698 buckwheat starches from 12 varieties (magnification of 1000 and 5000).

699 **Fig. 2** The volume distribution (A), X-ray diffraction pattern (B), 1D SAXS curves (C)  
700 and Lorentz-corrected 1D SAXS profiles (D) of buckwheat starches from 12 varieties.

701 **Fig. 3** The normalized 1D correlation function of buckwheat starches from 12 varieties.

702 **Fig. 4** Physicochemical properties of buckwheat starches from 12 varieties. The  
703 relationship between water solubility and swelling power (A), light transmittance (B),  
704 pasting curve (C) and DSC thermogram (D).

705 **Fig. 5** Rheological properties of buckwheat starch from 12 varieties. Frequency sweep  
706 curves (A, B and C), Han plot (D), creep recovery curves (E) and creep compliance (F).

707 **Fig. 6** PCA analysis and cluster analysis based on the structural and physicochemical  
708 properties of buckwheat starches from 12 varieties. The loading plot (A), score plot (B)  
709 and dendrogram (C) (Ts: total starch; Mo: moisture; As: ash; Pr: protein; Am: amylose;  
710 Rc: relative crystallinity; dc: crystalline layer thickness; da: amorphous layer thickness;

711 dac: long period distance; WS75: water solubility at 75°C; WS85: water solubility at  
712 85°C; WS95: water solubility at 95°C; SP75: swelling power at 75°C; SP85: swelling  
713 power at 85°C; SP95: swelling power at 95°C; Lt: light transmittance; PV: peak  
714 viscosity; HS: holding strength; FV: final viscosity; BD: breakdown; SBP: setback from  
715 peak; SBT: setback from trough; PT: pasting temperature; To: onset temperature; Tp:  
716 peak temperature; Tc: conclusion temperature and  $\Delta H$ : gelatinization enthalpy).

717 **Table 1** The origin, yield and main chemical compositions of buckwheat starches from  
718 12 varieties.

719 **Table 2** The particle size distribution of buckwheat starches from 12 varieties.

720 **Table 3** The lamellar structure parameters of buckwheat starches from 12 varieties.

721 **Table 4** The pasting properties and thermal properties of buckwheat starches from 12  
722 varieties.